Excretion of fetal biliverdin by the rat placenta-maternal liver tandem

Oscar Briz, Rocío I. R. Macías, María J. Pérez, María A. Serrano, and José J. G. Marín. Excretion of fetal biliverdin by the rat placenta-maternal liver tandem. Am J Physiol Regul Integr Comp Physiol 290: R749–R756, 2006. First published November 3, 2005; doi: 10.1152/ajpregu.00487.2005.—Fetal liver immaturity is accompanied by active heme catabolism. Thus fetal biliary pigments must be excreted toward the mother by the placenta. To investigate biliverdin handling by the placenta-maternal liver tandem, biliverdin-IX was administered to 21-day pregnant rats through the jugular vein or the umbilical artery of an in situ perfused placenta. Jugular administration resulted in the secretion into maternal bile of both bilirubin and biliverdin (3:1). However, when biliverdin was administered to the placenta, most of it was transformed into bilirubin before being transferred to the maternal blood. Injecting Xenopus laevis oocytes with mRNA from rat liver or placenta enhanced their ability to take up biliverdin, which was inhibited by estradiol 17β-β-glucuronidase. The expression of three OATP isoforms in this system revealed that they have a varying degrees of ability to transport biliverdin (Oatp1/1a1 > Oatp2/1a4 > Oatp4/1b2). The abundance of their mRNA in rat trophoblast was Oatp1/1a1 > Oatp4/1b2 > Oatp2/1a4. The expression of biliverdin-IX reductase in rat placenta was detected by RT-PCR/sequencing and Western blot analysis. The relative abundance of biliverdin-IX reductase mRNA (determined by real-time quantitative RT-PCR) was fetal liver > placenta > maternal liver. Common bile duct ligation in the last week of pregnancy induced an upregulation of biliverdin-IX reductase in maternal liver but had no effect on fetal liver and placenta. In conclusion, several members of the OATP family may contribute to the uptake of fetal biliverdin by the rat placenta. Before being transferred to the mother, biliverdin is extensively converted into bilirubin by biliverdin-IX reductase, whose expression is maintained even though bilirubin excretion into maternal bile is impaired.

bilirubin; biliary pigment; heme; pregnancy; transport

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isoforms involved in the liver uptake of organic anions may play a role in the carrier-mediated uptake of fetal BV by the rat placenta and liver and by measuring the levels of expression of BVRRα in the three elements of the fetal liver-placenta-maternal liver trio involved in the handling of fetal BV in native form or biotransformed into BR.

**MATERIAL AND METHODS**

*Chemicals and animals.* Biliverdin (BV) IXα hydrochloride (more than 95% pure, as confirmed by HPLC, see below) was obtained from Frontier Scientific (Logan, UT), which is considered a reliable source than 95% pure, as confirmed by HPLC, see below) was obtained from PerkinElmer Life Sciences (Pacisa & Giralt, Madrid, Spain). Cholic acid (TC, 5 Ci/mmol) was obtained from PerkinElmer Life Sciences (Pacisa & Giralt, Madrid, Spain). Estradiol 17β-D-glucuronide (E217β/H9251) was obtained from Frontier Scientific (Logan, UT), which is considered a reliable source than 95% pure, as confirmed by HPLC, see below) was obtained from PerkinElmer Life Sciences (Pacisa & Giralt, Madrid, Spain).

Pregnant rats on day 21 of gestation (~370 g) from the University of Salamanca Animal House (Salamanca, Spain) and mature female frogs (*X. laevis*), purchased from Regine Olig (Hamburg, Germany), were used. The animals received humane care as outlined in the National Institutes of Health Guidelines for the Care and Use of Laboratory Animals (Institutional Animal Care and Use Committee Guidebook, 2nd ed., 2002). Experimental protocols were reviewed and approved by the Ethical Committee for Laboratory Animals of the University of Salamanca. Harvesting and preparation of oocytes from *X. laevis* were performed as previously described (6).

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To carry out these experiments, the rats were anesthetized with sodium pentobarbital (Nembutal N.R.; Abbot, Madrid, Spain) by intraperitoneal injection (50 mg/kg body wt). When needed, polyethylene catheters were inserted into the maternal left jugular vein (for infusion) and the left carotid artery (for sampling). Bile flowing through a polyethylene catheter implanted into the common bile duct was collected in preweighed vials placed on ice and protected from light to prevent the degradation of bilirubin. Immediately after surgical preparation, the animals were left for an equilibration period. To carry out these experiments, the rats were anesthetized with sodium pentobarbital (Nembutal N.R.; Abbot, Madrid, Spain) by intraperitoneal injection (50 mg/kg body wt). When needed, polyethylene catheters were inserted into the maternal left jugular vein (for infusion) and the left carotid artery (for sampling). Bile flowing through a polyethylene catheter implanted into the common bile duct was collected in preweighed vials placed on ice and protected from light to prevent the degradation of biliary pigments. Immediately after surgical preparation, the animals were left for an equilibration period of 20 min before starting sample collection.

In situ single-pass perfusion of rat placenta was carried out as previously described in detail (4). Briefly, one of the placentas was exposed and perfused through the umbilical artery using a heparinized saline solution (0.05% heparin, 150 mM NaCl) at 37°C at a constant inflow of 500 μl/min with a peristaltic pump. An incision in the umbilical vein was performed to permit free outflow. Perfusion pressure was measured as an indicator of placental perfusion resistance and was considered to be appropriate when it remained relatively constant and lower than 20 cm H2O throughout the experimental period.

To compare BV handling by the placenta-maternal liver tandem vs. the maternal liver alone or the placenta alone, four separate sets of experiments were carried out, in which BV in 150 mM NaCl at pH 7.8 was administered: 1) 0.1 μmol BV was administered to the mother as a bolus (50 μl/min over 5 min) through the jugular vein. 2) 1.0 μmol BV was single-pass-perfused (500 μl/min over 5 min) through the umbilical artery, followed by continuous perfusion of this placenta with heparinized saline solution throughout the rest of the experimental period. 3) To prevent the biotransformation of the BV by the maternal liver and kidneys once it had been transferred to the mother through the placenta, immediately before administering 1.0 μmol BV through the umbilical artery, the maternal liver and kidneys were rapidly isolated from the maternal circulation using previously employed ligatures. The viability of these preparations during the experimental period was checked by measuring steady-state concentrations of urea in serum (7) and glucose in blood (Accutrend Sensor kit, Roche Diagnostics, Barcelona, Spain). 4) To investigate the profile of spontaneous accumulation of biliary pigments in serum in this model, similar experiments were carried out, except that vascular isolation of the liver and kidneys was not followed by administration of BV through the umbilical artery. Samples of maternal serum and/or bile were collected periodically to carry out analytical measurements.

To investigate the effect of blocking the maternal biliary excretion of biliary pigments on the expression of BVRRα in the fetal liver-placenta-maternal liver trio, complete obstruction of the maternal common bile duct was imposed on day 14 of gestation, and samples of the above tissues were collected on day 21, as described elsewhere (34).

**Measurement of gene expression levels.** To determine mRNA levels by real-time RT-PCR, tissue samples were immediately immersed in the RNA stabilization reagent RNAlater (Qiagen, Izasa, Barcelona, Spain) and stored at −80°C until total RNA was isolated using RNAeasy spin columns from Qiagen (Izasa, Barcelona, Spain). After treatment with RNase-free DNase I (Roche, Barcelona, Spain), RNA was quantified fluorometrically with the RiboGreen RNA Quantitation kit (Molecular Probes). cDNA was synthesized from 2 μg of total RNA using random hexamers and avian myeloblastosis virus RT (Enhanced Avian RT-PCR kit; Sigma-Genosys, Cambridge, UK), according to the instructions supplied by the vendor.

Real-time quantitative PCR was then performed using AmpliTaq Gold polymerase (Applied Biosystems, Madrid, Spain) in an ABI Prism 5700 Sequence Detection System (Applied Biosystems). The thermal cycling conditions were as follows: a single cycle at 95°C for 10 min followed by 40 cycles at 95°C for 15 s and at 60°C for 60 s. Detection of amplification products was carried out using SYBR Green I. Nonspecific products of PCR, as detected by 2.5% agarose gel electrophoresis or melting temperature curves, were not found in any case. The results of mRNA abundance for each target gene in each sample were normalized on the basis of its 18S RNA content, which was measured with the TaqMan Ribosomal RNA Control Reagents kit (Applied Biosystems).

The primer oligonucleotide sequences and conditions for measuring the absolute abundances of Oatp1/1a1 (gene symbol Slc21a1/Sclo1a1), Oatp2/1a4 (Slc21a5/Sclo1a4), and Oatp4/1b2 (Slc21a10/Sclo1b2) and rat placental lactogen type II, a specific marker for trophoblastic cells, have been described elsewhere (34). The primer oligonucleotide sequences for BVRRx were forward primer (position 138–140, GeneBank Accession Number BC078766) 5′-TTG GAG TGG TAG TGG TTG GTG TT-3′ and reverse primer (position 288–266) 5′-TCC AAA GAA ATC TGC GTG ACT TC-3′. All primers were designed with the assistance of Primer Express software (Applied Biosystems), and their specificity was checked using BLAST. They were obtained from Sigma-Genosys. Sequencing of the amplified fragments of rat BVRRx cDNA in maternal liver, fetal liver, and placenta resulted in a similarity of 100% with a previously described sequence (GeneBank Accession Number BC078766).

Western blot analyses on liver and placenta homogenates were carried out in 7.5% SDS-polyacrylamide gels using equal protein loading (150 μg per lane) as confirmed by staining of membranes after transfer with Ponceau Red-S (Sigma-Aldrich). Rabbit polyclonal antibody OSA-400 against BVRRα was from Stressgen Bioreagents (bioNova Cientifica, Madrid, Spain). Donkey anti-rabbit IgG horseradish peroxidase-conjugated antibody NA934 and enhanced chemiluminescence reagents were from Amersham Pharmacia Biotech (Freiburg, Germany).

**Uptake studies using *X. laevis* oocytes.** Using the PolyATract mRNA Isolation System purchased from Promega (Innogenetix, Barcelona, Spain), poly(A) mRNA to be injected into the oocytes was isolated from total RNA, obtained from rat liver or placenta as described above. The synthesis of cRNA of rat Oatpl/1a1, Oatpl/1a4, and Oatp4/1b2 for injection into oocytes was performed using recombinant plasmids containing the open reading frame DNA of these transporters, all kindly supplied by Dr. Peter J. Meier, Dr. Bruno Stieger, and Dr. Bruno Hagenbuch (Zurich University Hospital, Switzerland). These plasmids were isolated from *Escherichia coli* using...
the Qiagen Plasmid Mini Kit (Izasa) and further linearized with restriction enzymes. Capped and poly(A) tailed cRNAs were synthesized using the T7 mMessage mMachine Ultra kit (Ambion; bioNova Cientifica).

X. laevis oocytes were microinjected with TE buffer (1 mM EDTA, 10 mM Tris, pH 8.0) either alone, for determining nonspecific uptake and binding of the substrates, or containing the following amounts of RNA: 25 ng mRNA from rat liver or placenta or 7 ng cRNA of each OATP. Oocytes were used 2 days after RNA injection, when, on the basis of preliminary experiments on the time course of the functional expression for these carriers, the uptake rate was highest (data not shown).

Uptake studies were carried out using groups of five oocytes per data point. The data represent the results of at least three separate sets of RNA injections. Oocytes were washed with substrate-free uptake medium (100 mM choline chloride, 2 mM KCl, 1 mM CaCl2, 1 mM MgCl2, and 10 mM HEPES/Tris, pH 7.0) and incubated with 100 μl of uptake medium containing 50 μM [3H]TC or 100 μM BV at 25°C for 1 h. Uptake was stopped by the addition of 4 ml ice-cold uptake medium. The oocytes were washed three more times before being collected and were placed in vials individually (to measure radioactive substrate by liquid scintillation) or in groups of two oocytes (to increase the absorbance signal in HPLC analysis of BV uptake).

Analytical and statistical methods. Bile flow was determined gravimetrically. BR concentrations in bile were determined by a modification of the diazotized sulfanilic acid coupling method (18) and BV by HPLC as described below. To determine BV and BR in serum, both pigments were extracted (2), and subsequently dissolved in methanol. The mixture was treated to transform BR glucuronides into the corresponding methyl esters, as described elsewhere (5), evaporated under nitrogen and dissolved in 200 μl lysis/extraction solution (methanol containing 50 pmol ROX as internal standard for HPLC) to be analyzed by HPLC using 1 ml/min flow rate in a solvent gradient system. The initial mobile-phase composition was 100% phosphate buffer (2.5 mM, pH 7.2). After 1 min, the solvent was progressively changed to 50% methanol and 50% phosphate buffer over 2 min. This elution system was maintained for 1 min before starting a 5-min linear change to 100% methanol, which was maintained until the end of the runtime (20 min). Measurement of absorbances at 378, 450, or 567 nm wavelength for BV, BR, or ROX, respectively, were used for peak detection.

To measure BV in oocytes, these were placed in vials (in groups of two oocytes) and treated with 150 μl lysis/extraction solution and sonicated for 5 min in an ice-chilled bath. Then, oocyte lysates were centrifuged at 20,000 g for 5 min, and the supernatants were used to measure BV by HPLC.

Results are expressed as means ± SD. Regression lines were calculated by the least-squares method. To calculate the statistical significance of the differences between groups, the paired Student’s t-test or the Bonferroni method for multiple-range testing were used, as appropriate.

RESULTS

To evaluate the overall handling of BV by the placenta-maternal liver tandem, 1.0 μmol BV was administered through the umbilical artery of in situ perfused rat placenta in a single-pass perfusion system (Fig. 1). Only 1.6% of the pigment was recovered in maternal bile—almost all of it (99%) biotransformed into BR—over the following 90 min [15.5 (SD 3.2) nmol BR plus 0.13 (SD 0.05) nmol BV]. Moreover, in a different set of experiments, 1.0 μmol BV was administered through the umbilical artery of in situ perfused rat placenta, just after vascular isolation of the maternal liver and kidneys (Fig. 2). The result was that a peak in serum BV concentrations appeared before the general deterioration, which occurred after 20 min, as monitored by increasing levels of urea and decreasing glucose concentrations in maternal serum. A transient peak of BV concentrations was followed by a delayed (5–10 min), and progressive, increase in unconjugated BR, but not in conjugated BR (data not shown), concentrations in maternal
serum (Fig. 2). The magnitude of the increase in serum concentrations for BV and BR is consistent with the amount of BR secreted into bile when BV was administered to animals with intact liver and kidney circulation. No detectable spontaneous accumulation of any of these pigments was observed in animals up to 20 min after vascular isolation of the liver and kidneys when BV was not administered through the umbilical artery (BV was not detected and BR concentrations remained below 0.3 μM) (Fig. 2).

To measure the proportion of BV converted into BR by the maternal liver after crossing the placenta, a dose of 10% of that administered through the umbilical artery (i.e., 0.1 μmol) was administered directly into the maternal circulation by injection through the jugular vein (Fig. 3). Approximately 80% of this amount was eliminated in bile over the following 90 min as a mixture of BR and BV. Most of the secreted pigments (80%, 61.7 (SD 3.6) nmol) were found to be BV biotransformed into BR, but a smaller, although considerable, amount (20%, 15.7 (SD 4.0 nmol) was secreted in bile as BV.

To study the ability of several OATP isoforms to transport BV, these transporters were expressed in X. laevis oocytes, and BV uptake was measured in a HPLC system. ROX was used as an internal standard, because BV and ROX showed similar (98%) recoveries from oocytes undergoing the lysis and extraction protocol (data not shown), similar retention time in HPLC, and low interferences between them in the recording of absorbance chromatograms (Fig. 4A). Using this method, the
presence of endogenous BV in X. laevis oocytes was detected. This is not surprising because this pigment has been found previously in eggs from very different species (22). The technique permitted linear detection of BV over a large concentration range (Fig. 4B) and a clear discrimination between non-specific uptake in noninjected oocytes (Fig. 4C) and transport-mediated uptake. Indeed, injection of oocytes with mRNA from rat placenta resulted in an enhanced uptake of BV (Fig. 4D). When carrier-mediated uptake (uptake in injected oocytes minus that in noninjected oocytes) was calculated, the ability of three members of the OATP family, involved in the uptake of organic anions by the rat liver to induce sodium-independent uptake of TC (Oatp4/1b2 > Oatp1/1a1 > Oatp2/1a4) was observed (Fig. 5). This was more marked for BV (Oatp1/1a1 > Oatp2/1a4 > Oatp4/1b2) (Fig. 5). A similar ability to enhance overall uptake of TC was induced by injection of mRNA from either rat placenta or maternal liver, and both were similarly inhibited by E217β (Fig. 6), a substrate shared by several OATP isoforms (10). Moreover, mRNA from placenta induced a significantly higher BV uptake than that from maternal liver (Fig. 6). E217β also induced a dramatic inhibition of BV uptake in both cases (Fig. 6). It should be noted that in experiments carried out with oocytes injected with total mRNA, overall uptake was the result of an uptake-efflux balance, as export pumps were probably also expressed.

To investigate the relevance of OATPs as transporters responsible for BV uptake by the rat placenta, their expression levels in this organ were determined by measurement of their absolute abundance of mRNA (Fig. 7A). In all cases, the expression levels were markedly lower in placenta than in maternal liver. Moreover, the most abundant isoform in the placenta was Oatp1/1a1. When these values were corrected by the amount of trophoblast, using the rat placental lactogen type II mRNA as a marker, the abundance of mRNA for Oatp1/1a1 was 7- to 8-fold higher than that for Oatp2/1a4 and Oatp4/1b2 (Fig. 7B).

To study the potential contribution of fetal and maternal livers and the placenta to the biotransformation of fetal BV into BR, the relative abundance of BVRα in these organs was determined by real-time quantitative RT-PCR, after confirming that no nontarget products were generated during amplification, as revealed by the appearance of a single peak in the DNA melting curves and 2.5% agarose gel electrophoresis of amplification products (Fig. 8A).

To compare the relative expression of this enzyme in different tissues, the abundance of its mRNA and protein was considered per gram of tissue (Fig. 8B) and by Western blot analysis per fixed amount of total protein (Fig. 8B, inset), respectively. The highest relative expression was found in fetal liver, followed by the placenta and the maternal liver. To
evaluate the total relevance of each organ in the overall activity of this enzyme in the whole animal, the relative abundance of mRNA was corrected by the weight of each organ (Fig. 8C). The highest total amount of mRNA was found in maternal liver, followed by the fetal liver and the placenta.

Cholestasis imposed in pregnant rats during the last week of pregnancy induced a marked increase in maternal serum total BR concentrations from 0.14 (SD 0.03) mg/dl in controls (n = 6) to 3.02 (SD 0.51) mg/dl in the group with cholestasis (n = 6; P < 0.05), together with an upregulation of BVRα in maternal liver (Fig. 8, B and C). However, the repercussion on fetal serum BR concentrations was less marked [from 0.31 (SD 0.08) mg/dl in the group with cholestasis (n = 12; P < 0.05)] and was not accompanied by any effect on the expression of BVRα in fetal liver (Fig. 8, B and C). The expression of BVRα in the placenta, which is exposed to blood from both compartments, was not affected by maternal cholestasis either (Fig. 8, B and C).

**DISCUSSION**

Many studies have been conducted recently to shed light on the role of plasma membrane transporters in the rat and human placental handling of endogenous substances and xenobiotics (16, 20, 29). The present study provides direct evidence that several of the OATPs expressed in rat placenta and maternal liver are able to take up BV from the fetal and maternal circulations. Among them, Oatp1/1a1 and Oatp4/1b2 are abundantly expressed in the maternal liver, and, together with the contribution to a lesser extent of Oatp2/1a4, may play an important role in BV uptake from the maternal blood. As regards the placenta, Oatp1/1a1, owing to its ability to transport BV and its high expression compared with that of Oatp2/1a4 and Oatp4/1b2, is probably the most important of these three transporters contributing to BV uptake by this organ from fetal blood. Nevertheless, other transporters may also participate in this function.

The observation that when mRNA from liver or placenta was injected in *X. laevis* oocytes, the uptake by these cells was higher for BV than for TC, is likely due to the fact that Na⁺-independent uptake, which was the type measured in the present study, could be more important for BV than for TC. It should be noted that at the basolateral surface of rat hepatocytes, TC uptake is mainly governed by the very efficient Na⁺-taurocholate cotransporting polypeptide (Ntcp, gene symbol Scl10a1) (11). Moreover, the expression of export pumps may differently reduce the net amount of TC and BV found in the cells after 60 min incubation.

In in vivo experiments, only a minor part of the amount of BV that was taken up by the rat placenta was rapidly released toward the mother, followed by a delayed release of BR. This suggests that the transport of BV from the trophoblast toward the mother is very poor and/or that placental biotransformation of BV into BR is very efficient. Indeed, our results confirm the
expression in rat placenta of BVRα, whose enzymatic activity has previously been reported (25). Moreover, expression levels of this enzyme in the placenta, which were higher than that in maternal liver, are consistent with the observed extensive biotransformation of BV into BR during placental transit. The fact that the expression of BVRα in fetal liver was even higher than in the placenta, suggests that an important part of endogenous BV produced by the fetus could be transformed into BR by the fetal liver before being taken up by the placenta.

Although several human OATPs, namely OATP-C/1B1 and OATP8/1B3, are able to transport BR (6, 8) and could play a role in BR uptake by the placenta (6), little is known about the fate of this pigment, or that of BR, during the rest of transplacental transit (19).

Inside the trophoblast, BR is probably partly bound to lipids and proteins such as glutathione S-transferase (39). Functional studies have suggested that from there, BR would be exported across the apical pole of the trophoblast via an ATP-dependent mechanism (35). Whether one or several isoforms of multidrug resistance-associated proteins (MRPs), such as MRP1 and MRP2, which are expressed in human (38) and rat placenta (34, 37), are involved in this process is not known. Canalicular MRP2 plays a key role in the secretion into bile of mono- and di-glucuronides of BR (12). However, owing to the low UDP-glucuronosyltransferase activity of the fetal liver and the absence of placental biotransformation of unconjugated BR during transplacental transfer, when this compound is administered through the umbilical artery of in situ perfused rat placentas (5), MRP2 is not expected to play an important role in BR transfer across the placenta. Moreover, the excretion of dicarboxylate organic anions related to BV into rat bile is also mediated by MRP2-independent mechanisms (24), suggesting that this export pump is not responsible for the poor release of BR from the trophoblast into the maternal blood across the apical pole of this epithelium.

Like MRP2, MRP1 is also able to perform ATP-dependent transport of bilirubin glucuronides (12). Although indirect evidence has suggested that MRP1 may also be involved in the efflux of unconjugated BR from human choriocarcinoma BeWo cells (28), its role in BR and BV transport across the placenta remains to be elucidated.

An interesting observation of the present study was that the impairment in maternal bile secretion induced an upregulation of BVRα in the maternal liver. This probably enhances the ability of this organ to generate BR from BV as part of the redox cycle, BR being an important defense mechanism against oxidative stress (1), such as that caused by maternal cholestasis (30). However, probably because of the lower exposure of fetal liver to compounds accumulated in maternal blood (17), the expression of BVRα in fetal liver was not affected by maternal cholestasis. The absence of changes in BVRα expression in paired placentas is consistent with the concept that hepatobiliary-like excretory mechanism of the placenta is for the most part unidirectional (5), and therefore constitutes an efficient barrier that excludes, at least in part, cholephilic compounds from the trophoblast.

As we have previously reported that several OATPs (Oatp1/1a1, Oatp2/1a4, and Oatp4/1b2) and MRPs (Mrp1, Mrp2, and Mrp3) are indeed upregulated in rat placenta in response to maternal cholestasis (34), the fact that BVRα expression in maternal liver, but not in placenta, was sensitive to maternal cholestasis suggests that the placenta would be equipped with different mechanisms for regulating gene expression in response to accumulation in the maternal compartment of cholephilic organic anions. Indeed, although several endogenous, as well as exogenous substances, may activate the nuclear receptors—the pregnane X receptor and the constitutive androstane receptor—resulting in changes in the expression of genes such as Slco1a4, Abcc2 (for Mrp2) and Abcc3 (for Mrp3) in the rat liver (13, 36), these receptors are poorly expressed in rat placenta (37). Nevertheless, the expression of several OATPs and MRPs can be induced in rat placenta by maternal cholestasis (34), which in addition to these events also causes enhanced oxidative stress and apoptosis (31). This results in an overall impairment of the ability of this organ to transfer cholephilic organic anions, such as bile acids, from the fetus to the mother (34).

In conclusion, our results suggest that among transporters involved in fetal BR uptake by the rat placenta, several OATPs, in particular Oatp1/1a1, may be involved. Once in the placenta, and before transfer to the mother, BV is extensively converted into BR by BVRα, which is highly expressed in this organ. The small amount of BV that reaches the maternal blood is efficiently taken up, probably in part by Oatp1/1a1, Oatp2/1a4, and Oatp4/1b2, and biotransformed into BR, which joins fetal BR transferred as such by the placenta, to be eliminated mainly through secretion into bile by the maternal liver.

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