

## CALL FOR PAPERS | *Physiology and Pharmacology of Temperature Regulation*

### Thermoregulation: some concepts have changed.

### Functional architecture of the thermoregulatory system

**Andrej A. Romanovsky**

*Systemic Inflammation Laboratory, Trauma Research, St. Joseph's Hospital and Medical Center, Phoenix, Arizona*

Submitted 21 September 2006; accepted in final form 23 September 2006

**Romanovsky AA.** Thermoregulation: some concepts have changed. Functional architecture of the thermoregulatory system. *Am J Physiol Regul Integr Comp Physiol* 292: R37–R46, 2007. First published September 28, 2006; doi:10.1152/ajpregu.00668.2006.— While summarizing the current understanding of how body temperature ( $T_b$ ) is regulated, this review discusses the recent progress in the following areas: central and peripheral thermosensitivity and temperature-activated transient receptor potential (TRP) channels; afferent neuronal pathways from peripheral thermosensors; and efferent thermoeffector pathways. It is proposed that activation of temperature-sensitive TRP channels is a mechanism of peripheral thermosensitivity. Special attention is paid to the functional architecture of the thermoregulatory system. The notion that deep  $T_b$  is regulated by a unified system with a single controller is rejected. It is proposed that  $T_b$  is regulated by independent thermoeffector loops, each having its own afferent and efferent branches. The activity of each thermoeffector is triggered by a unique combination of shell and core  $T_b$ s. Temperature-dependent phase transitions in thermosensory neurons cause sequential activation of all neurons of the corresponding thermoeffector loop and eventually a thermoeffector response. No computation of an integrated  $T_b$  or its comparison with an obvious or hidden set point of a unified system is necessary. Coordination between thermoeffectors is achieved through their common controlled variable,  $T_b$ . The described model incorporates Kobayashi's views, but Kobayashi's proposal to eliminate the term sensor is rejected. A case against the term set point is also made. Because this term is historically associated with a unified control system, it is more misleading than informative. The term balance point is proposed to designate the regulated level of  $T_b$  and to attract attention to the multiple feedback, feedforward, and open-loop components that contribute to thermal balance.

BY USING POWERFUL AUTONOMIC and even more powerful behavioral means, our species survives while being exposed to a wide range of ambient temperatures: from  $-110^\circ\text{C}$  (the surface of the Moon) to  $2,000^\circ\text{C}$  (the air around a space shuttle as it reenters the atmosphere). Even in these diversified thermal environments, we usually manage to maintain our deep (core) body temperature ( $T_b$ ) within a few tenths of a degree Celsius. In fact, an exodus of  $T_b$  from its usual range is so suggestive of a pathological condition, that  $T_b$  is monitored regularly in all hospitalized patients and reported in every medical history. Various aspects of  $T_b$  regulation have been discussed in fifty-one original articles (4, 7, 13, 22, 23, 29, 35–38, 40–42, 45, 46, 48–51, 54, 55, 59, 61, 62, 64–66, 71, 73, 76, 77, 79, 80, 85, 88,

99–101, 104, 106, 107, 110, 111, 115, 118, 120, 123–127), four reviews (20, 31, 32, 108), ten editorials (33, 68a, 70, 89, 90, 95, 109, 112, 113, 121) and one point (57)-counterpoint (11) exchange within the Special Call for Papers on Physiology and Pharmacology of Temperature Regulation published in several issues of the *American Journal of Physiology—Regulatory, Integrative and Comparative Physiology* over the past two years. While closing this Call, the present review summarizes the recent progress in our understanding of how body temperature is regulated. To give readers an idea of how remarkable this progress is, the new understanding (based on the latest developments) is compared with the information provided by a typical textbook chapter on thermoregulation. It seems that this chapter may need some updates, especially in its coverage of thermosensors, thermoeffectors, and the functional architecture of the thermoregulatory system.

#### THERMOSENSORS

##### *Central Thermosensors*

A typical textbook chapter would say that brain temperature is detected by central thermosensory neurons (central “thermoreceptors”). Most of them are warm-sensitive, that is, they increase their activity with an increase in brain temperature. The abundance of warm-sensitive central sensors is consistent with the following two facts. First, our thermal physiology is “asymmetrical:”  $T_b$  is positioned very closely, within just a few degrees Celsius, to the upper survival limit (which is possibly determined by the denaturation of regulatory proteins) but relatively far, a few tens of degrees, from the lower limit (which is likely determined by the freezing of water). Therefore, core overheating is much more dangerous than overcooling. Second, humans are endothermic animals (meaning that their principal source of heat is their own body), as opposed to ectothermic animals (that receive heat primarily from the environment). Hence, sensors for limiting heat gain have to be located inside the body. Although much less common, cold-sensitive neurons (i.e., those that increase their activity with a decrease in brain temperature) also exist. However, the cold sensitivity of most of them seems to be due to inhibitory synaptic input from nearby warm-sensitive neurons.

Thermoregulatory responses in a variety of animal species can be elicited by local thermal stimulation of various areas in the central nervous system, including several brain stem neu-

Address for reprint requests and other correspondence: A. A. Romanovsky, Trauma Research, St. Joseph's Hospital, 350 W. Thomas Rd., Phoenix, AZ 85013 (E-mail: aromano@chw.edu).

The costs of publication of this article were defrayed in part by the payment of page charges. The article must therefore be hereby marked “advertisement” in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

ronal groups [that used to be referred to as the reticular formation(s) of the medulla oblongata, pons, and midbrain; see Ref. 9] and the spinal cord, but thermosensitive neurons of the preoptic anterior hypothalamus (POA) are considered to be the most important for triggering autonomic thermoeffector responses. It bears mentioning that the locations of thermosensitive neurons triggering various thermoregulatory behaviors are largely unknown (see *Behavioral effectors*).

For a long time, it was assumed that the roles of cold- and warm-sensitive POA neurons are reciprocal and “symmetrical,” that is, that all thermoregulatory responses could be triggered by either activation of one class of the temperature-sensitive neurons or inhibition of the other (10); your textbook is likely to share this assumption. During the past decade, however, the idea of equally important roles of warm- and cold-sensitive neurons was put to rest. Elegant studies by Kanosue and colleagues (21, 128), involving thermal and chemical stimulation of POA cells, showed that both cold-defense and heat-defense autonomic responses are initiated by the corresponding changes in the activity of warm-sensitive neurons; increased activity of warm-sensitive POA neurons triggers heat-defense responses, while decreased activity triggers cold-defense responses.

Morphological identification of thermosensitive neurons has been another major achievement. Griffin et al. (43) showed that these cells are characterized by the horizontal orientation of their dendrites: toward the third ventricle medially and the medial forebrain bundle laterally. Because neurons conveying temperature signals from the body surface and viscera enter the hypothalamus via the periventricular stratum and medial forebrain bundle (24), such an orientation seems ideal for receiving input through both projections. The present Call for Papers has contributed to our understanding of the functional properties of thermosensitive neurons (11, 57, 123). Because warm-sensitive POA neurons display spontaneous membrane depolarization, as shown by Boulant and colleagues (123, 129), these cells are considered pacemakers; their thermosensitivity is due to currents that determine the rate of spontaneous depolarization between successive action potentials. It is true, however, that mechanisms of hypothalamic thermosensitivity continue to be disputed (57). The changes in ion currents that underlie both central (*vide supra*) and peripheral (*vide infra*) thermosensitivity are likely to involve temperature-dependent phase transitions (91).

### Peripheral Thermosensors

There are also peripheral thermosensory neurons (peripheral “thermoreceptors”) that detect shell temperatures in the skin and in the oral and urogenital mucosa. A typical textbook chapter would state that most superficial sensors are cold-sensitive. Because central thermosensors are concerned mainly with warmth, specialization of peripheral sensors in cold sensitivity is not that surprising. Skin cold sensors are located in or immediately beneath the epidermis. Their signals are conveyed by thin myelinated A $\delta$  fibers. The less common warm sensors are located slightly deeper in the dermis; their signals travel via unmyelinated C fibers. Peripheral thermosensors are not pacemakers, and mechanisms of peripheral thermosensitivity are thought to involve changes in the resting membrane potential. Importantly, the response of most peripheral thermosensors

shows a powerful dynamic (phasic) component: these cells are very active when the temperature is changing, but quickly adapt to a stable temperature. Such a response enables the organism to rapidly react to environmental changes. In addition to superficial cold- and warm-sensitive neurons, there are peripheral deep-body sensors, which respond to the core body temperature. They are located in the esophagus, stomach, large intra-abdominal veins, and other organs.

This basic information about peripheral thermosensors has been recently updated with three developments: 1) the discovery of a subclass of transient receptor potential (TRP) ion channels known as thermoTRP channels; 2) the progress in identifying thermoafferent pathways and their differential involvement in thermosensation and  $T_b$  regulation; and 3) the challenge to the old idea that a separate neuronal network computes some integrated  $T_b$  (from codes of local  $T_b$ s) and compares it to a set point  $T_b$  to form a thermal sensation and to send “orders” to thermoeffectors.

### ThermoTRP Channels

The mammalian TRP superfamily consists of ~30 channels divided in six subfamilies known as the TRPC (canonical), TRPV (vanilloid), TRPM (melastatin), TRPML (mucolipin), TRPP (polycystin), and TRPA (ankyrin). Of these, the heat-activated TRPV1-V4, M2, M4, and M5 and the cold-activated TRPM8 and A1 are often referred to as the thermoTRP channels. Involvement of these recently cloned and characterized channels in thermoregulation has been studied intensively. In the present Call for Papers, these studies are represented by the original articles by Ni et al. (80) and Wechselberger et al. (123), the invited review by Caterina (20), and, to a certain extent, the point-counterpoint exchange between Kobayashi et al. (57) and Boulant (11). While referring the reader to the review by Caterina (20) and several other recent reviews (30, 87, 102) for more detailed information, I would like to emphasize a few points.

First, activation of all thermoTRP channels results in an inward nonselective cationic current and, consequently, in an increase in the resting membrane potential. This mechanism agrees more readily with a role for these channels in peripheral thermosensitivity (83) rather than hypothalamic thermosensitivity (11, 129). Furthermore, the TRPV4 channel (which is likely to play a physiological role in thermoregulation; *vide infra*) does not seem to be expressed in the bodies of POA neurons (39), but it is expressed by the neuronal bodies of dorsal root ganglia (44).

Second, although each thermoTRP channel is activated within a relatively narrow temperature range, the range that they cover cumulatively is very wide: from noxious cold to noxious heat (Fig. 1). Furthermore, they cover this temperature range in an overlapping fashion, and their activities have different sensitivities to temperature. These features make the thermoTRP channels well suited to the job of peripheral thermosensors. It is important to note, however, that the temperature ranges shown in Fig. 1 were obtained *in vitro*. *In vivo*, a specific cell type on which a thermoTRP channel is expressed and concomitant activation with chemical ligands affect the temperature range in which the channel is activated and, in some cases, bring it closer to physiological values of deep  $T_b$ . Indeed, although the TRPV1 channel is widely thought to be

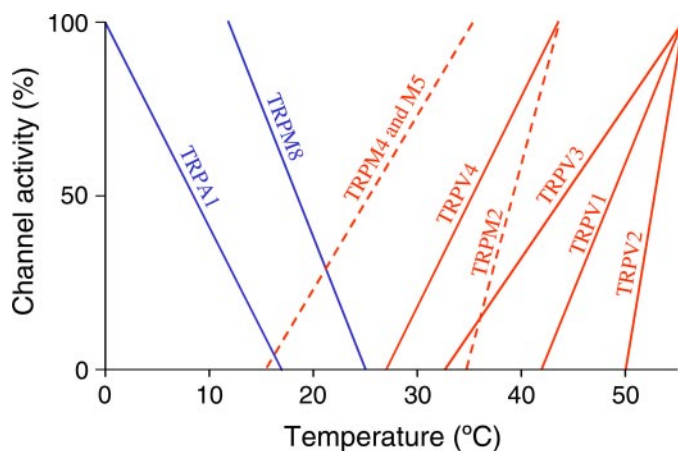


Fig. 1. Schematic representation of the dependence of the activity of cold-activated (blue) and heat-activated (red) thermoTRP channels on temperature. The thresholds of activation and temperatures of maximal activation are based on the activity of the channels in heterologous systems; some of the thresholds are means of values obtained in several studies. The figure is adapted from Patapoutian et al. (87). Information on the TRPM2 is added based on Togashi et al. (119); information on the TRPM4 and M5 is added based on Talavera et al. (114); the added lines are dashed. Please note that quantitative aspects of the relationships shown should be looked upon with great skepticism, as the figure does not account for several important factors (81). Nevertheless, this figure illustrates how well thermoTRP channels are suited for detecting physiologically relevant temperatures both in the shell and core.

activated at pain-inducing temperatures of >42°C (20, 30), Ni et al. (80) showed that increasing temperature within the normal physiological range (perhaps as low as ~34.5°C) can exert a direct stimulatory effect on pulmonary sensory neurons, and this effect is likely mediated through the activation of the TRPV1 channel and other subtypes of TRPV channels. For some TRPV channels (namely, the V3 and V4), likely physiological roles have been established (63, 74). Other thermoTRP channels are currently under investigation.

Afferent Pathways

**Discriminative sensation.** There is little doubt that afferent pathways start with primary thermosensory neurons. The bod-

ies of these bipolar cells are located in the dorsal root ganglia, and the central axons project to the dorsal horn of the spinal cord (mostly lamina I), where they synapse with secondary monopolar neurons. Axons of these secondary neurons cross the midline and ascend in the lateral funiculus of the spinal cord. It was believed for a long time that the secondary neurons project directly to the ipsilateral ventrobasal complex of the thalamus, from where their signals are conveyed to the ipsilateral somatosensory cortex (postcentral gyrus) by tertiary neurons, and your textbook may hold this to be true. According to Craig (27, 28), however, this pathway, which is involved in tactile sensation, is uninvolved in temperature sensation. Instead, the lamina-I neurons carry temperature signals to the insular cortex (the island of Reil) with a relay in the posterolateral thalamus (specifically, the posterior part of the ventromedial nucleus) or with two relays (in the parabrachial nucleus and the basal part of the ventromedial nucleus of the thalamus) (Fig. 2). These two branches of the spino-thalamo-cortical pathway are involved in discriminative temperature sensation. A functional magnetic resonance imaging study in humans by Hua et al. (48) published in this Call for Papers shows that this pathway is organized topically, as evident from the topical representation of skin temperature in the insula. This pathway allows for sensing temperature at a high spatial resolution (e.g., temperature of the surface under the tip of an individual finger can be assessed). Therefore, this pathway is important for making decisions about a wide range of issues related to interactions with the environment, but it appears to have little to do with  $T_b$  regulation, that is, with triggering thermoeffector responses to defend deep  $T_b$ . It should also be noted that the spino-thalamo-cortical pathway (as described here for humans) is not the same in other vertebrates, as several interspecies differences have been noticed (27).

**Homeostatic control.** Thermoeffector responses are triggered by thermal exposures massive enough to affect heat exchange between the body and the environment. Temperature-generated signals from large areas of the shell are sent to the brain through the spino-reticulo-hypothalamic pathway (Fig. 2), in which the secondary lamina-I neurons project to the reticular formation (medullar, pontine, and midbrain neuronal

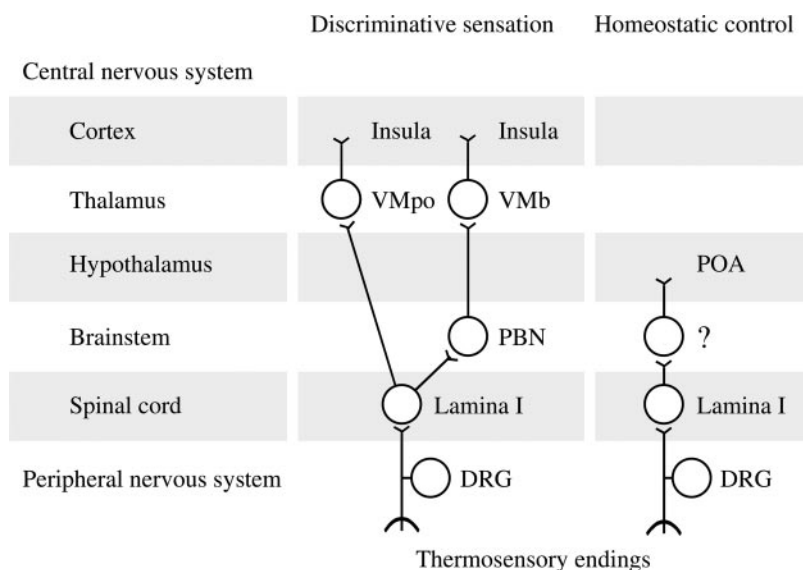


Fig. 2. Afferent neuronal pathways for discriminative sensation/localization of a thermal stimulus and for homeostatic control of body temperature. DRG, dorsal root ganglion; PBN, parabrachial nucleus; POA, preoptic anterior hypothalamus; VMb, basal part of the ventromedial nucleus of the thalamus (formerly known as the parvicellular part of the ventroposterior medial nucleus); VMpo, posterior part of the ventromedial nucleus of the thalamus; ?, unknown location(s) within the medulla, pons, and midbrain.

groups). In this pathway, thermosensory “lines” converge at the level of lamina-I neurons (i.e., a single lamina-I cell receive inputs from multiple thermosensory neurons; see Ref. 82) and possibly at other levels, so that downstream neurons (e.g., brain stem neurons in thermoeffector pathways) can collect signals from large thermoreceptive fields (5, 6). The tertiary neurons of the afferent spino-reticulo-hypothalamic pathway project to hypothalamic structures (including the POA) either via the periventricular stratum passing along the wall of the third ventricle or via the medial forebrain bundle, which passes more laterally. As described above, warm-sensitive POA neurons have a dendrite orientation ideal for collecting information from both the periventricular stratum and the medial forebrain bundle. More precise delineation of this homeostatic, spino-reticulo-hypothalamic pathway, especially of the “reticulo” part of it, requires further study.

#### *Thermosensors or Thermostats?*

Our views on how temperature is sensed have been challenged recently by Kobayashi and colleagues (56, 58, 83). The pre-Kobayashi models of  $T_b$  regulation assumed that thermoreceptors code temperatures of different parts of the body (into neuronal activity codes) and that these codes of local temperatures are then integrated by a separate network (that consists of several neurons with different roles) into some mean temperature. The location of the integrating network was unclear (8a). Nevertheless, it was often assumed that this integrating network also compares the integrated temperature with an external or internal reference signal and, based on such a comparison, somehow generates individual orders to thermoeffectors.

Kobayashi and colleagues proposed a different scenario (56, 58, 83). According to them, a sensory neuron is wired through a number of neurons to an effector cell. When the temperature to which the temperature-sensitive part of a sensory neuron is exposed reaches the activation threshold of this neuron, the neuron fires and, through its connections, sends a signal to the effector cell. If a large number of sensory neurons sends signals to their effector cells, a thermoeffector response occurs. In this model, a decision to trigger an effector is made “automatically” (principally, by sensory neurons) and involves neither a separate decision-making network nor operations with temperature codes (computation of a mean  $T_b$ ). According to this model, a sensation is a “side product” of the activation of those neurons that are wired to cause certain effector responses (i.e., feeling cold means having activated those neurons that trigger cold-defense responses). The same principle is used by Kobayashi and colleagues (58) to explain how we sense other modalities as well.

Kobayashi’s model also explains how deep  $T_b$ s and peripheral (e.g., skin)  $T_b$ s contribute to thermoregulation. Deep  $T_b$ s are regulated variables, and they are very stable. From the point of view of the control theory, they serve as feedback signals. Peripheral  $T_b$ s, on the other hand, are not regulated; they are highly variable (97). They are feedforward signals that, according to the control theory, allow the body to respond to a thermal load “in advance,” that is, before deep  $T_b$ s start changing. In Kobayashi’s model, both deep and peripheral  $T_b$ s drive effector responses in a similar way. Which central and peripheral sensory neurons are wired to a particular thermoeffector

determines which combinations of core and shell  $T_b$ s activate this effector (see *Thermoeffector Loops*).

Despite its conceptual strength, Kobayashi’s model has not become widely accepted yet, perhaps because the author favors rather drastic terminological changes: he proposes to call sensors *thermostats* (or *comparators*). Kobayashi is certainly right that, from the engineering point of view, the role of thermoreceptors in his model is that of thermostats, and not of sensors. However, in the minds of the majority of medical and biological scientists and students, the current terms (sensor, thermosensor, sensory, somatosensory, etc.) are associated with no particular engineering analog. Hence, biologists feel no urge whatsoever to get rid of the entire family of widely used biological terms or to start translating them to the engineering language. (Later in this review, I will discuss a different term, set point, which is associated with a false idea in the minds of most biologists and physicians and, therefore, must be replaced.) In the case of thermosensors, a more productive approach might be to save the term, but to provide it with a different meaning.

## THERMOEFFECTORS

### *Effectors*

Your textbook is likely to name various autonomic thermoeffectors and possibly some behavioral ones, to discuss their anatomy, mechanisms of physiological control, and effects on heat balance. Various aspects of thermoregulatory skin vasoconstriction (29, 118) and vasodilation (4, 55, 71, 127), thermogenesis in brown adipose tissue (BAT) (79), shivering (62), and thermoregulatory behavior (41) have been addressed in the present Call for Papers. Some interactions between thermogenesis, energy metabolism, and body temperature regulation have been analyzed in several original articles (37, 45, 49, 124), two editorials (112, 113), and the invited review by Diepvens et al. (31). I would also like to refer the reader to the fundamental review on BAT by Cannon and Nedergaard (17) published in *Physiological Reviews*. What your textbook is unlikely to cover is the efferent neuronal pathways to thermoeffectors.

### *Efferent Pathways*

*Autonomic effectors.* Efferent pathways to thermoeffectors have not been well characterized in humans, although this topic has recently started receiving attention (34). Even in the rat, the most studied laboratory species, not all thermoeffector pathways have been mapped. Some of the neuronal circuitries connecting the warm-sensitive hypothalamic neurons to autonomic thermoeffectors in the rat, particularly the BAT and skeletal muscles (heat-production effectors) and the vasculature of the tail (a specialized heat-exchange organ), have been characterized during the past decade (Fig. 3). Studies of these pathways have been propelled by the development of transsynaptic retrograde tracing techniques using pseudorabies virus, along with the refinement of techniques for discrete lesioning and pharmacological stimulation and inhibition of neural structures. The revealed pathways appeared complex, and their detailed description is beyond the scope of the present paper; see the invited review by DiMicco and Zaretsky (32) in the present issue, as well as the reviews by Nagashima et al. (78)

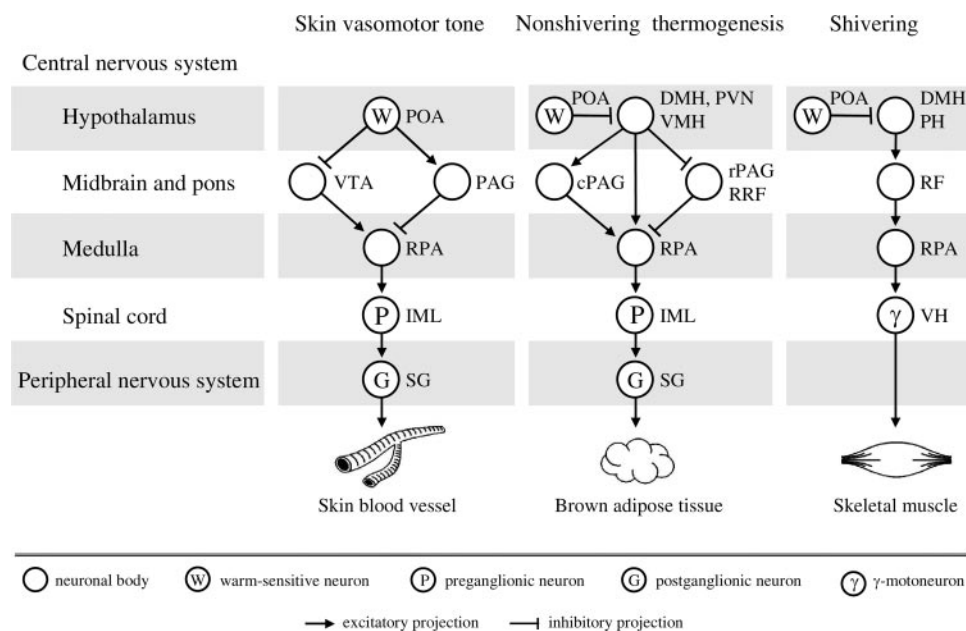


Fig. 3. Efferent neuronal pathways for control of skin vasomotor tone, nonshivering thermogenesis in brown adipose tissue, and shivering in the rat. The concept was taken from Nagashima et al. (78); the figure was substantially modified and published in Romanovsky (93) by permission from Elsevier. The Romanovsky (93) version is reproduced here with a minor modification and by permission from both Elsevier and Blackwell Publishing. DMH, dorsomedial hypothalamus; IML, intermediolateral column; PAG, periaqueductal gray matter; cPAG, caudal PAG; rPAG, rostral PAG; PH, posterior hypothalamus; PVN, paraventricular nucleus; RF, reticular formation; RPA, raphé/peripyramidal area; RRF, retrorubral field; SG, sympathetic ganglia; VH, ventral horn; VMH, ventromedial hypothalamus; VTA, ventral tegmental area.

and Morrison (75) published elsewhere. However, a few points deserve elaboration.

As shown in Fig. 3, both the BAT and skin vasculature are controlled by sympathetic ganglia, with the bodies of preganglionic neurons located in the intermediolateral column of the spinal cord. These spinal neurons receive direct input from cells located primarily in the raphé/peripyramidal area of the medulla. These medullary cells are under the control of hypothalamic (dorsomedial and paraventricular nuclei), midbrain (periaqueductal gray matter, retrorubral field, and ventral tegmental area), and possibly pontine (locus coeruleus) neurons that receive input from warm-sensitive POA cells (8, 18, 84). Although the efferent pathways for skin vasomotor tone and nonshivering thermogenesis have some similarities, they are not identical, and both differ substantially from the pathway controlling shivering. Within the shivering pathway,  $\gamma$  and  $\alpha$  motoneurons of the ventral horn receive direct and indirect inputs from the midbrain and brain stem, including the raphé/peripyramidal area of the medulla (116). Axons of the midbrain neurons descend the spinal cord with the reticulospinal and rubrospinal tracts. These midbrain neurons are under control of posterior hypothalamic neurons, which, in turn, receive inhibitory input from warm-sensitive POA cells.

The efferent pathways described are to a large extent inhibitory. Consequently, thermoeffector activation involves disinhibition of tonically inhibited neurons. Importantly, thermoeffectors are controlled relatively independently of each other (69, 78), and certain portions of the pathways may be recruited in a thermoregulatory response in a stimulus-specific fashion. The latter speculation is supported by findings that the paraventricular nucleus (16, 47, 67) and locus coeruleus (3) seem to mediate a thermogenic response to bacterial lipopolysaccharide or prostaglandin E<sub>2</sub> but not cold-induced thermogenesis or a nocturnal T<sub>b</sub> rise in rodents. In the present Call for Papers, neural circuitries of autonomic thermoeffector responses have been subjects of original articles by Nakamura and Morrison (79), Ootsuka and McAllen (85), Tanaka and McAllen (115)

and editorial foci by DiMicco and Zaretsky (33) and McAllen (68a).

**Behavioral effectors.** Evidence (mostly from stimulation experiments) suggests that different thermoregulatory behaviors in the rat (e.g., relaxed postural extension, thermoregulatory grooming, and locomotion) use distinct neural circuitries (92). However, the neuroanatomic substrate of no thermoregulatory behavior has been studied extensively, and little is known about the neuroanatomy of behavioral thermoregulation (78). This situation is likely to change, as behavioral thermoregulation is becoming a subject of keen attention (1, 2, 59, 60, 68). In the present issue, Konishi et al. (59) report that neurons in the median preoptic nucleus are involved in the intensification of an operant thermoregulatory behavior (moving to a reward zone during heat exposure to trigger a breeze of cold air) caused by hypertonic saline. For a different behavioral response (moving to a cold environment during bacterial lipopolysaccharide-induced shock), two other substrates have been recently identified by Almeida et al. (2): neurons of the dorsomedial hypothalamic nucleus and fibers passing through the paraventricular hypothalamic nucleus. By studying warmth- and cold-seeking behaviors of rats in six different tests, Almeida et al. (2) also showed that these behaviors do not require an intact POA, whereas autonomic thermoregulatory responses do.

#### FUNCTIONAL ARCHITECTURE OF THE THERMOREGULATORY SYSTEM

*“Think simple” as my old master used to say—meaning reduce the whole of its parts into the simplest terms, getting back to first principles.*

—Frank Lloyd Wright (1868–1959)

*“The ability to simplify means to eliminate the unnecessary so that the necessary may speak.”*

—Hans Hofmann (1880–1966)

### *Thermoeffector Loops*

Thermoregulatory pathways form distinct thermoeffector loops. The efferent parts of the loops clearly differ, because each effector has its own efferent pathway (Fig. 3); the article by Ootsuka and McAllen (85) in this Call for Papers provides further support for this thesis. The afferent parts are also not identical, as each effector receives a unique combination of signals from peripheral and central thermosensors. The question as to which temperatures (thermosensors in which parts of the body) trigger which effectors has been recently revisited in several studies (14, 116), including the one by Nakamura and Morrison (79) in the present issue.

In general (although fully realizing that generalization may not be the best approach in this particular case), behavioral responses depend more on signals from peripheral thermosensors (shell temperatures) than central thermosensors (core temperature) (92), whereas deep  $T_b$  is relatively more important for triggering autonomic responses (52, 103). Such an organization reflects the fact that behavioral responses are often aimed at escaping the forthcoming thermal insult. In contrast, autonomic cold-defense responses (energetically expensive) and heat-defense responses (water-consuming) are often recruited only when deep  $T_b$  starts changing because behavioral mechanisms were ineffective or could not be used (e.g., due to competing behavioral demands). Even within the autonomic responses, different thermoeffector responses are triggered by different combinations of peripheral and central  $T_b$ s. Because peripheral thermosensors are mostly cold sensors, information from peripheral sensors is relatively more important for triggering cold-defense effectors (14, 79, 103, 116) than heat-defense ones (103). Because central thermosensors are mostly warm sensors, information from central sensors is relatively more important for triggering heat-defense responses (103).

Combinations of shell and core  $T_b$ s that trigger the same thermoeffector response in different species are also likely to differ. The great thermal inertia of large animals makes transient thermal exposures less threatening, which decreases the importance of feedforward regulation. Indeed, peripheral thermosensitivity is relatively more important in smaller animals, whereas central thermosensitivity is relatively more important in larger animals (72). Therefore, caution should be exercised while extrapolating results obtained in rats (this section of the present review is based primarily on such results) to human thermophysiology.

### *Thermoeffector Coordination*

Usually, the recruitment of thermoeffectors into a thermoregulatory response looks like a highly coordinated event. Those effectors that affect heat balance in the opposite directions are typically not activated simultaneously. Energetically expensive and water-consuming responses are typically triggered after those that do not consume a lot of energy or water. For a long time, such coordination between thermoeffectors was explained with the help of a complex neuronal network (coordinator) within a single integrated system, the same (or a similar) network that was thought to be responsible for the formation of thermal sensations (see *Thermosensors or Thermostats?*). However, the coordinator has never been found experimentally. Furthermore, each effector was found to be

controlled by a distinct group of neurons (see *Thermoeffector Loops*).

It is also accepted now that the anatomically distinct thermoeffectors function largely independently (105), and a body of experimental data has been accumulated showing that different effectors sometimes defend drastically different levels of  $T_b$  (for a review, see Ref. 94). An example of differential responses of thermoeffectors is endotoxin shock: it is accompanied by a large ( $2^\circ\text{C}$ ) decrease in the threshold hypothalamic temperature for activation of cold-induced thermogenesis, but a small (a few tenths of a degree) or no decrease in the threshold hypothalamic temperature for triggering tail skin vasodilation (98). Data showing independent effector responses are often questioned (e.g., Refs. 15 and 19) based on the fact that a thermoeffector can be recruited by another homeostatic system to meet a competing demand and, hence, can become unavailable for thermoregulation. In the example with endotoxin shock (98), skin vasoconstriction is needed to maintain blood pressure, thus preventing thermoregulatory skin vasodilation. The real question, however, is not whether an "unusual" thermoeffector response has a compelling teleological explanation, but whether the thresholds of different thermoeffectors can change independently of each other. The data accumulated (reviewed in Ref. 94) clearly show that thermoeffector thresholds often change independently, and this is a rather strong argument against a unified model of the thermoregulatory system with a single controller. Models of the thermoregulation system with multiple controllers (relatively independent thermoeffector loops) to replace the unified system have been proposed (e.g., Ref. 53).

Furthermore, it has been realized that any regulatory system can exist, using the words of Partridge (86), as a group of relatively autonomous controllers, acting in a common environment with generally compatible rules, but at any particular time, operating with only limited active coordinating linkages, and at no time acting as a unified system with a single controller. In fact, almost any regulated variable in the body, for example, arterial blood pressure (39), is likely to be an emergent product of a decentralized control system.

Coming back to the thermoregulatory system, basic coordination between thermoeffectors is likely to be achieved through their dependence on a common variable:  $T_b$ . Such coordination can be explained, in a simplified way, as follows. When one thermoeffector is activated, its activity changes  $T_b$ , which changes the position of  $T_b$  relative to the thresholds of other thermoeffectors, which, in turn, triggers activation or cessation of other thermoeffector mechanisms (for a more detailed description, see Ref. 93).

### *What To Do with the Term Set Point?*

On a related issue, there has been a recent upsurge of interest in the term set point (12, 15, 19, 93, 94, 96). The original meaning of the term—a physical (thermal, electric, etc.), externally originated reference signal in a unified control system—is now considered invalid almost unanimously. However, the term is still used widely, mostly in the following three meanings. First, it is used to designate some internal property of the unified thermoregulatory system that substitutes for an external reference signal. Needless to say, such a usage is obviously wrong as it refers to the same unified system.

Second, it is used to designate some internal characteristics (usually thresholds) of individual thermoeffector loops or their subcomponents (e.g., Kobayashi's "thermostats" or individual thermoTRP channels). There is nothing wrong with such a usage of the term, except that it creates confusion: this usage refers to a set point as a property of an individual component, whereas all definitions of thermoregulatory responses (see next paragraph) use the term set point while referring to the entire system.

Third, many colleagues use this term to designate a regulated level of  $T_b$  (e.g., Refs. 12, 15, 19). Such a usage is in accordance with the most recent thermophysiological glossary (25), and many definitions of thermoregulatory responses (fever, anaprexia, hypothermia, hyperthermia) are built upon this definition of the set point. However, in the minds of biologists, physicians, and students, the term set point is strongly, perhaps inseparably, associated with the reference signal of a unified thermoregulatory system. Even today, it is not unusual for scientists outside the thermoregulation field to talk about the set point temperature (117) or about neurons that detect the error signal (26). Even for scientists within the thermoregulation field, it is rather typical to seemingly accept the relative mutual independence of individual thermoeffector loops, but to use (whether intentionally or not) the unified model with a single controller while describing how  $T_b$  is regulated (12, 15), to still conclude that all thermoeffectors operate according to a common plan (15), and to propose a neuronal model of a set point of a unified thermoregulation system (12).

In other words, the intrinsic association of the term set point with nonexistent physical signals (a computed mean  $T_b$  and reference  $T_b$ ) within the nonexistent unified thermoregulatory system complicates the usage of this term. It provides reference to the engineering analogies that are more misleading than informative. To eliminate this often unintended reference, I have proposed to use the term balance point when talking about the regulated level of  $T_b$  (93, 94, 96). The balance point-based definitions work for all cases where the set point-based definitions of thermoregulatory responses work (25). More importantly, they also work for all cases where the set point-based definitions may not work (94). As an added benefit of such a substitution, the term balance point redirects the scientific search from looking for the location of the set point (or building a new model of it) to studying the multiple feedback, feedforward, and open-loop components that contribute to thermal balance in the thermoregulatory system operating as a federation of independent thermoeffector loops. Interestingly, many scientists in the field are already avoiding using the term set point altogether or replacing it with different terms (e.g., Ref. 10). In his editorial about the regulation of body fat content, Wade (122) suggests to put the notions of lipostats and set points behind us and to move on. Applying Wade's suggestion to  $T_b$  regulation, it is time to free the thermophysiological terminology of remnants of the unified control system and to focus our research on studying independent thermoeffector loops.

#### INSTEAD OF CONCLUSIONS

In this review, I have tried to make a point that thermoeffectors can coordinate their activities and regulate  $T_b$  while functioning within relatively independent loops, without a

unified control system. To understand how thermoeffector loops work, we need to study, among other things, thermosensors (including the thermoTRP channels; Fig. 1), as well as the afferent (Fig. 2) and efferent (Fig. 3) portions of thermoeffector loops. Figures 1–3 represent an attempt to summarize the recent progress achieved in these three areas. I invite the readers to use these figures in the classroom to complement the thermoregulation chapter they are currently using. I encourage viewing these figures as drafts and asking your students to correct and update them. A major shortcoming of these figures is that they follow the organization of the same unified thermoregulatory system that is extensively criticized in this review. Although the central controller is eliminated, different thermoeffector loops are cut across to represent the same level of all loops in each cross section: sensors, afferent pathways, and efferent pathways. To correct this shortcoming would require constructing a figure for each thermoeffector that would represent the entire loop. Inspirational examples of such constructs can be found in the article of Nakamura and Morrison (79) in the present issue. Investigations of the roles of individual thermoTRP channels in the control of different thermoeffectors have just been started, for example, by Almeida et al. (1, 2), who used TRPV1 and TRPM8 agonists to cause thermoregulatory locomotion.

#### ACKNOWLEDGMENTS

I thank all of the past and present members of my laboratory, especially Drs. Alexandre A. Steiner, Victoria F. Turek, and M. Camila Almeida, for helping me with the work on which the current review was partially built, for educating me on various aspects of thermoregulation, and for discussing with me early drafts of this manuscript and the figures. My views on how  $T_b$  is regulated have been influenced by enlightening discussions with Drs. Kazuyuki Kanosue and the late Lloyd D. Partridge. A draft of Fig. 3 was discussed with Drs. Shaun F. Morrison and Christopher J. Madden. Dr. Thomas M. Hamm and F. E. Farmer read an early version of this review and provided important feedback.

This review closes the Special Call for Papers on Physiology and Pharmacology of Temperature Regulation of the *American Journal of Physiology—Regulatory, Integrative and Comparative Physiology* (volumes 288–292, 2005–2007). As Guest Editor for this Call, I thank Dr. Pontus B. Persson, Editor-in-Chief, for accepting the proposal for this project. I also thank the authors of the more than 100 manuscripts submitted in response to this Call, as well as the many colleagues in the field who expertly reviewed these submissions. Special thanks go to Olivia Kaferly, Assistant to the Editor-in-Chief. Like a good mother takes care of her children, Olivia took an excellent care of the editors, authors, and referees involved. She successfully led us through the project and made sure we would not fall in various organizational, technical, ethical, and other potholes.

#### GRANTS

The author's research reviewed in this paper has been supported by grants from the National Institute of Neurological Disorders and Stroke (NS41233), Arizona Biomedical Research Commission (8016), and St. Joseph's Foundation.

#### REFERENCES

1. Almeida MC, Steiner AA, Branco LGS, and Romanovsky AA. Cold-seeking behavior as a thermoregulatory strategy in systemic inflammation. *Eur J Neurosci* 23: 3359–3367, 2006.
2. Almeida MC, Steiner AA, Branco LGS, and Romanovsky AA. Neural substrate of cold-seeking behavior in endotoxin shock. *PLoS one* 1: e1, 2006.
3. Almeida MC, Steiner AA, Coimbra NC, and Branco LGS. Thermoeffector neuronal pathways in fever: a study in rats showing a new role of the locus coeruleus. *J Physiol* 558: 283–294, 2004.
4. Aoki K, Stephens DP, Zhao K, Kosiba WA, and Johnson JM. Modification of cutaneous vasodilator response to heat stress by daytime

- exogenous melatonin administration. *Am J Physiol Regul Integr Comp Physiol* 291: R619–R624, 2006.
5. **Asami A, Asami T, Hori T, Kiyohara T, and Nakashima T.** Thermally-induced activities of the mesencephalic reticulospinal and rubrospinal neurons in the rat. *Brain Res Bull* 20: 387–398, 1988.
  6. **Asami T, Hori T, Kiyohara T, and Nakashima T.** Convergence of thermal signals on the reticulospinal neurons in the midbrain, pons and medulla oblongata. *Brain Res Bull* 20: 581–596, 1988.
  7. **Atanackovic D, Pollok K, Faltz C, Boeters I, Jung R, Nierhaus A, Braumann K-M, Hossfeld DK, and Hegewisch-Becker S.** Patients with solid tumors treated with high-temperature whole body hyperthermia show a redistribution of naive/memory T-cell subtypes. *Am J Physiol Regul Integr Comp Physiol* 290: R585–R594, 2006.
  8. **Bamshad M, Song CK, and Bartness TJ.** CNS origins of the sympathetic nervous system outflow to brown adipose tissue. *Am J Physiol Regul Integr Comp Physiol* 276: R1569–R1578, 1999.
  - 8a. **Berner NJ and Heller HC.** Does the preoptic anterior hypothalamus receive thermoafferent information? *Am J Physiol Regul Integr Comp Physiol* 274: R9–R18, 1998.
  9. **Blessing WW.** Inadequate frameworks for understanding bodily homeostasis. *Trends Neurosci* 20: 235–239, 1997.
  10. **Bligh J.** A theoretical consideration of the means whereby the mammalian core temperature is defended at a null zone. *J Appl Physiol* 100: 1332–1337, 2006.
  11. **Boulant JA.** Counterpoint: heat-induced membrane depolarization of hypothalamic neurons: an unlikely mechanism of central thermosensitivity. *Am J Physiol Regul Integr Comp Physiol* 290: R1481–R1484, 2006; Discussion R1484, 2006.
  12. **Boulant JA.** Neuronal basis of Hammel's model for set-point thermoregulation. *J Appl Physiol* 100: 1347–1354, 2006.
  13. **Bradford CD, Cotter JD, Thorburn MS, Walker RJ, and Gerrard DF.** Exercise can be pyrogenic in humans. *Am J Physiol Regul Integr Comp Physiol* 292: R143–R149, 2007.
  14. **Bratincsak A and Palkovits M.** Evidence that peripheral rather than intracranial thermal signals induce thermoregulation. *Neuroscience* 135: 525–532, 2005.
  15. **Cabanac M.** Adjustable set point: to honor Harold T. Hammel. *J Appl Physiol* 100: 1338–1346, 2006.
  16. **Caldeira JC, Franci CR, and Pela IR.** Bilateral lesion of hypothalamic paraventricular nucleus abolishes fever induced by endotoxin and bradykinin in rats. *Ann NY Acad Sci* 856: 294–297, 1998.
  17. **Cannon B and Nedergaard J.** Brown adipose tissue: function and physiological significance. *Physiol Rev* 84: 277–359, 2004.
  18. **Cano G, Passerin AM, Schiltz JC, Card JP, Morrison SF, and Sved AF.** Anatomical substrates for the central control of sympathetic outflow to interscapular adipose tissue during cold exposure. *J Comp Neurol* 460: 303–326, 2003.
  19. **Caputa M.** Comments on "Do fever and anapyrexia exist? Analysis of set point-based definitions". *Am J Physiol Regul Integr Comp Physiol* 289: R281, 2005; Reply R281–R282, 2005.
  20. **Caterina MJ.** Transient receptor potential ion channels as participants in thermosensation and thermoregulation. *Am J Physiol Regul Integr Comp Physiol* 292: R64–R76, 2007.
  21. **Chen XM, Hosono T, Yoda T, Fukuda Y, and Kanosue K.** Efferent projection from the preoptic area for the control of non-shivering thermogenesis in rats. *J Physiol* 512: 883–892, 1998.
  22. **Chevrier C, Bourdon L, and Canini F.** Cosignaling of adenosine and adenosine triphosphate in hypobaric hypoxia-induced hypothermia. *Am J Physiol Regul Integr Comp Physiol* 290: R595–R600, 2006.
  23. **Chu AL, Jay O, and White MD.** The effects of hyperthermia and hypoxia on ventilation during low-intensity steady-state exercise. *Am J Physiol Regul Integr Comp Physiol* 292: R195–R203, 2007.
  24. **Cliffer KD, Burstein R, and Giesler GJ Jr.** Distributions of spinothalamic, spinohypothalamic, and spinothalamic fibers revealed by anterograde transport of PHA-L in rats. *J Neurosci* 11: 852–868, 1991.
  25. **Commission for Thermal Physiology of the International Union of Physiological Sciences.** Glossary of terms for thermal physiology. *Jpn J Physiol* 51: 245–280, 2001.
  26. **Costa AC, Stasko MR, Stoffel M, and Scott-McKean JJ.** G-protein-gated potassium (GIRK) channels containing the GIRK2 subunit are control hubs for pharmacologically induced hypothermic responses. *J Neurosci* 25: 7801–7804, 2005.
  27. **Craig AD.** How do you feel? Interoception: the sense of the physiological condition of the body. *Nat Rev Neurosci* 3: 655–666, 2002.
  28. **Craig AD.** Interoception: the sense of the physiological condition of the body. *Curr Opin Neurobiol* 13: 500–505, 2003.
  29. **DeGroot DW and Kenney WL.** Impaired defense of core temperature in aged humans during mild cold stress. *Am J Physiol Regul Integr Comp Physiol* 292: R103–R108, 2007.
  30. **Dhaka A, Viswanath V, and Patapoutian A.** TRP ion channels and temperature sensation. *Annu Rev Neurosci* 29: 135–161, 2006.
  31. **Diepvens K, Westerterp KR, and Westerterp-Plantenga MS.** Obesity and thermogenesis related to the consumption of caffeine, ephedrine, capsaicin, and green tea. *Am J Physiol Regul Integr Comp Physiol* 292: R77–R85, 2007.
  32. **DiMicco JA and Zaretsky DV.** The dorsomedial hypothalamus: a new player in thermoregulation. *Am J Physiol Regul Integr Comp Physiol* 292: R47–R63, 2007.
  33. **DiMicco JA and Zaretsky DV.** The mysterious role of prostaglandin E<sub>2</sub> in the medullary raphe: a hot topic or not? *Am J Physiol Regul Integr Comp Physiol* 289: R1589–R1591, 2005.
  34. **Egan GF, Johnson J, Farrell M, McAllen R, Zamarripa F, McKinley MJ, Lancaster J, Denton D, and Fox PT.** Cortical, thalamic, and hypothalamic responses to cooling and warming the skin in awake humans: a positron-emission tomography study. *Proc Natl Acad Sci USA* 102: 5262–5267, 2005.
  35. **Fabricio ASC, Rae GA, Zampronio AR, D'Orléans-Juste P, and Souza GEP.** Central endothelin ET<sub>B</sub> receptors mediate IL-1-dependent fever induced by preformed pyrogenic factor and corticotropin-releasing factor in the rat. *Am J Physiol Regul Integr Comp Physiol* 290: R164–R171, 2006.
  36. **Fabricio ASC, Tringali G, Pozzoli G, Melo MC, Vercesi JA, Souza GEP, and Navarra P.** Interleukin-1 mediates endothelin-1-induced fever and prostaglandin production in the preoptic area of rats. *Am J Physiol Regul Integr Comp Physiol* 290: R1515–R1523, 2006.
  37. **Fahlman A, Schmidt A, Handrich Y, Woakes AJ, and Butler PJ.** Metabolism and thermoregulation during fasting in king penguins, *Aptenodytes patagonicus*, in air and water. *Am J Physiol Regul Integr Comp Physiol* 289: R670–R679, 2005.
  38. **Feleder C, Perlik V, Tang Y, and Blatteis CM.** Putative antihyperpyretic factor induced by LPS in spleen of guinea pigs. *Am J Physiol Regul Integr Comp Physiol* 289: R680–R687, 2005.
  39. **Fink GD.** Hypothesis: the systemic circulation as a regulated free-market economy. A new approach for understanding the long-term control of blood pressure. *Clin Exp Pharmacol Physiol* 32: 377–383, 2005.
  40. **Ganta CK, Helwig BG, Blecha F, Ganta RR, Cober R, Parimi S, Musch TI, Fels RJ, and Kenney MJ.** Hypothermia-enhanced splenic cytokine gene expression is independent of the sympathetic nervous system. *Am J Physiol Regul Integr Comp Physiol* 291: R558–R565, 2006.
  41. **Gilbert C, Le Maho YL, Perret M, and Ancel A.** Body temperature changes induced by huddling in breeding male emperor penguins. *Am J Physiol Regul Integr Comp Physiol* 292: R176–R185, 2007.
  42. **Gray DA, Maloney SK, and Kamerling PR.** Lipopolysaccharide-induced fever in Pekin ducks is mediated by prostaglandins and nitric oxide and modulated by adrenocortical hormones. *Am J Physiol Regul Integr Comp Physiol* 289: R1258–R1264, 2005.
  43. **Griffin JD, Saper CB, and Boulant JA.** Synaptic and morphological characteristics of temperature-sensitive and -insensitive rat hypothalamic neurons. *J Physiol* 537: 521–535, 2001.
  44. **Guler AD, Lee H, Iida T, Shimizu I, Tominaga M, and Caterina M.** Heat-evoked activation of the ion channel, TRPV4. *J Neurosci* 22: 6408–6414, 2002.
  45. **Gutman R, Choshniak I, and Kronfeld-Schor N.** Defending body mass during food restriction in *Acomys russatus*: a desert rodent that does not store food. *Am J Physiol Regul Integr Comp Physiol* 290: R881–R891, 2006.
  46. **Helwig BG, Parimi S, Ganta CK, Cober R, Fels RJ, and Kenney MJ.** Aging alters regulation of visceral sympathetic nerve responses to acute hypothermia. *Am J Physiol Regul Integr Comp Physiol* 291: R573–R579, 2006.
  47. **Horn T, Wilkinson MF, Landgraf R, and Pittman QJ.** Reduced febrile responses to pyrogens after lesions of the hypothalamic paraventricular nucleus. *Am J Physiol Regul Integr Comp Physiol* 267: R323–R328, 1994.
  48. **Hua LH, Strigo IA, Baxter LC, Johnson SC, and Craig AD.** Antero-posterior somatotopy of innocuous cooling activation focus in human



- dorsal posterior insular cortex. *Am J Physiol Regul Integr Comp Physiol* 289: R319–R325, 2005.
49. **Hübschle T, Mütze J, Mühlradt PF, Korte S, Gerstberger R, and Roth J.** Pyrexia, anorexia, adipisia, and depressed motor activity in rats during systemic inflammation induced by the Toll-like receptors-2 and -6 agonists MALP-2 and FSL-1. *Am J Physiol Regul Integr Comp Physiol* 290: R180–R187, 2006.
  50. **Ivanov AI, Steiner AA, Patel S, Rudaya AY, and Romanovsky AA.** Albumin is not an irreplaceable carrier for amphipathic mediators of thermoregulatory responses to LPS: compensatory role of  $\alpha_1$ -acid glycoprotein. *Am J Physiol Regul Integr Comp Physiol* 288: R872–R878, 2005.
  51. **Jay O, Gariépy LM, Reardon FD, Webb P, Ducharme MB, Ramsay T, and Kenny GP.** A three-compartment thermometry model for the improved estimation of changes in body heat content. *Am J Physiol Regul Integr Comp Physiol* 292: R167–R175, 2007.
  52. **Jessen C.** Independent clamps of peripheral and central temperatures and their effects on heat production in the goat. *J Physiol* 311: 11–22, 1981.
  53. **Kanosue K, Romanovsky AA, Hosono T, Chen XM, and Yoda T.** “Set point” revisited. In: *Thermal Physiology 1997*, edited by Nielsen Johanssen B and Nielsen R. Copenhagen: The August Krogh Institute, 1997, p. 39–43.
  54. **Kenny GP, Jay O, Zaleski WM, Reardon ML, Sigal RJ, Journeay WS, and Reardon FD.** Postexercise hypotension causes a prolonged perturbation in esophageal and active muscle temperature recovery. *Am J Physiol Regul Integr Comp Physiol* 291: R580–R588, 2006.
  55. **Kenny GP, Murrin JE, Journeay WS, and Reardon FD.** Differences in the postexercise threshold for cutaneous active vasodilation between men and women. *Am J Physiol Regul Integr Comp Physiol* 290: R172–R179, 2006.
  56. **Kobayashi S.** Temperature-sensitive neurons in the hypothalamus: a new hypothesis that they act as thermostats, not as transducers. *Prog Neurobiol* 32: 103–135, 1989.
  57. **Kobayashi S, Hori A, Matsumura K, and Hosokawa H.** Point: heat-induced membrane depolarization of hypothalamic neurons: a putative mechanism of central thermosensitivity. *Am J Physiol Regul Integr Comp Physiol* 290: R1479–R1480, 2006. Discussion R1484, 2006.
  58. **Kobayashi S, Okazawa M, Hori A, Matsumura K, and Hosokawa H.** Paradigm shift in sensory system—animals do not have sensors. *J Therm Biol* 31: 19–23, 2006.
  59. **Konishi M, Kanosue K, Kano M, Kobayashi A, and Nagashima K.** The median preoptic nucleus is involved in the facilitation of heat-escape/cold-seeking behaviour during systemic salt loading in rats. *Am J Physiol Regul Integr Comp Physiol* 292: R150–R159, 2007.
  60. **Konishi M, Nagashima K, Asano K, and Kanosue K.** Attenuation of metabolic heat production and cold-escape/warm-seeking behaviour during a cold exposure following systemic salt loading in rats. *J Physiol* 551: 713–720, 2003.
  61. **Kozak W, Wrotek S, and Kozak A.** Pyrogenicity of CpG-DNA in mice: role of interleukin-6, cyclooxygenases, and nuclear factor- $\kappa$ B. *Am J Physiol Regul Integr Comp Physiol* 290: R871–R880, 2006.
  62. **Kvadsheim PH, Folkow LP, and Blix AS.** Inhibition of shivering in hypothermic seals during diving. *Am J Physiol Regul Integr Comp Physiol* 289: R326–R331, 2005.
  63. **Lee H, Iida T, Mizuno A, Suzuki M, and Caterina MJ.** Altered thermal selection behavior in mice lacking transient receptor potential vanilloid 4. *J Neurosci* 25: 1304–1310, 2005.
  64. **Leite LH, Lacerda ACR, Marubayashi U, and Coimbra CC.** Central angiotensin AT<sub>1</sub>-receptor blockade affects thermoregulation and running performance in rats. *Am J Physiol Regul Integr Comp Physiol* 291: R603–R607, 2006.
  65. **Li Z, Perlik V, Feleder C, Tang Y, and Blatteis CM.** Kupffer cell-generated PGE<sub>2</sub> triggers the febrile response of guinea pigs to intravenously injected LPS. *Am J Physiol Regul Integr Comp Physiol* 290: R1262–R1270, 2006.
  66. **Lim CL, Wilson G, Brown L, Coombes JS, and Mackinnon LT.** Preexisting inflammatory state compromises heat tolerance in rats exposed to heat stress. *Am J Physiol Regul Integr Comp Physiol* 292: R186–R194, 2007.
  67. **Lu J, Zhang YH, Chou TC, Gaus SE, Elmquist JK, Shiromani P, and Saper CB.** Contrasting effects of ibotenate lesions of the paraventricular nucleus and subparaventricular zone on sleep-wake cycle and temperature regulation. *J Neurosci* 21: 4864–4874, 2001.
  68. **Maruyama M, Nishi M, Konishi M, Takashige Y, Nagashima K, Kiyohara T, and Kanosue K.** Brain regions expressing Fos during thermoregulatory behavior in rats. *Am J Physiol Regul Integr Comp Physiol* 285: R1116–R1123, 2003.
  - 68a. **McAllen RM.** The cold path to BAT. *Am J Physiol Regul Integr Comp Physiol* 292: R124–R126, 2007.
  69. **McAllen RM.** Preoptic thermoregulatory mechanisms in detail. *Am J Physiol Regul Integr Comp Physiol* 287: R272–R273, 2004.
  70. **McAllen RM and McKinley MJ.** Personal body maps. *Am J Physiol Regul Integr Comp Physiol* 289: R317–R318, 2005.
  71. **McCord GR, Cracowski J-L, and Minson CT.** Prostanoids contribute to cutaneous active vasodilation in humans. *Am J Physiol Regul Integr Comp Physiol* 291: R596–R602, 2006.
  72. **Mercer JB and Simon E.** A comparison between total body thermosensitivity and local thermosensitivity in mammals and birds. *Pflügers Arch* 400: 228–234, 1984.
  73. **Mochizuki T, Klerman EB, Sakurai T, and Scammell TE.** Elevated body temperature during sleep in orexin knockout mice. *Am J Physiol Regul Integr Comp Physiol* 291: R533–R540, 2006.
  74. **Moqrich A, Hwang SW, Earley TJ, Petrus MJ, Murray AN, Spencer KS, Andahazy M, Story GM, and Patapoutian A.** Impaired thermosensation in mice lacking TRPV3, a heat and camphor sensor in the skin. *Science* 307: 1468–1472, 2005.
  75. **Morrison SF.** Central pathways controlling brown adipose tissue thermogenesis. *News Physiol Sci* 19: 67–74, 2004.
  76. **Mouihate A, Ellis S, Harre E-M, and Pittman QJ.** Fever suppression in near-term pregnant rats is dissociated from LPS-activated signaling pathways. *Am J Physiol Regul Integr Comp Physiol* 289: R1265–R1272, 2005.
  77. **Mouihate A, Horn TF, and Pittman QJ.** Oxyresveratrol dampens neuroimmune responses in vivo: a selective effect on TNF- $\alpha$ . *Am J Physiol Regul Integr Comp Physiol* 291: R1215–R1221, 2006.
  78. **Nagashima K, Nakai S, Tanaka M, and Kanosue K.** Neuronal circuitries involved in thermoregulation. *Auton Neurosci* 85: 18–25, 2000.
  79. **Nakamura K and Morrison SF.** Central efferent pathways mediating skin cooling-evoked sympathetic thermogenesis in brown adipose tissue. *Am J Physiol Regul Integr Comp Physiol* 292: R127–R136, 2007.
  80. **Ni D, Gu Q, Hu H-Z, Gao N, Zhu MX, and Lee L-Y.** Thermal sensitivity of isolated vagal pulmonary sensory neurons: role of transient receptor potential vanilloid receptors. *Am J Physiol Regul Integr Comp Physiol* 291: R541–R550, 2006.
  81. **Nilius B, Talavera K, Owsianik G, Prenen J, Droogmans G, and Voets T.** Gating of TRP channels: a voltage connection? *J Physiol* 567: 35–44, 2005.
  82. **Nomoto S, Shibata M, Iriki M, and Riedel W.** Role of afferent pathways of heat and cold in body temperature regulation. *Int J Biometeorol* 49: 67–85, 2004.
  83. **Okazawa M, Takao K, Hori A, Shiraki T, Matsumura K, and Kobayashi S.** Ionic basis of cold receptors acting as thermostats. *J Neurosci* 22: 3994–4001, 2002.
  84. **Oldfield BJ, Giles ME, Watson A, Anderson C, Colvill LM, and McKinley MJ.** The neurochemical characterisation of hypothalamic pathways projecting polysynaptically to brown adipose tissue in the rat. *Neuroscience* 110: 515–526, 2002.
  85. **Ootsuka Y and McAllen RM.** Comparison between two rat sympathetic pathways activated in cold defense. *Am J Physiol Regul Integr Comp Physiol* 291: R589–R595, 2006.
  86. **Partridge LD.** The good enough calculi of evolving control systems: evolution is not engineering. *Am J Physiol Regul Integr Comp Physiol* 242: R173–R177, 1982.
  87. **Patapoutian A, Peier AM, Story GM, and Viswanath V.** ThermoTRP channels and beyond: mechanisms of temperature sensation. *Nat Rev Neurosci* 4: 529–539, 2003.
  88. **Perlik V, Li Z, Goorha S, Ballou LR, and Blatteis CM.** LPS-activated complement, not LPS per se, triggers the early release of PGE<sub>2</sub> by Kupffer cells. *Am J Physiol Regul Integr Comp Physiol* 289: R332–R339, 2005.
  89. **Persson PB.** Temperature control: from molecular insights, regulation in king penguins and diving seals, to studies in humans. *Am J Physiol Regul Integr Comp Physiol* 291: R512–R514, 2006.
  90. **Pittman QJ.** Endothelin—an emerging role in proinflammatory pathways in brain. *Am J Physiol Regul Integr Comp Physiol* 290: R162–R163, 2006.

91. **Pollack GH.** *Cells, Gels and the Engines of Life: A New, Unifying Approach to Cell Function.* Seattle: Ebner, 2001.
92. **Roberts WW.** Differential thermosensor control of thermoregulatory grooming, locomotion, and relaxed postural extension. *Ann NY Acad Sci* 525: 363–374, 1988.
93. **Romanovsky AA.** Temperature regulation. In: *Lecture Notes on Human Physiology*, edited by Petersen O. Oxford: Blackwell, 2006, chap. 23, p. 603–615.
94. **Romanovsky AA.** Do fever and anapyrexia exist? Analysis of set point-based definitions. *Am J Physiol Regul Integr Comp Physiol* 287: R992–R995, 2004.
95. **Romanovsky AA.** Vioxx, Celebrex, Bextra . . . do we have a new target for anti-inflammatory and antipyretic therapy? *Am J Physiol Regul Integr Comp Physiol* 288: R1098–R1099, 2005.
96. **Romanovsky AA, Almeida MC, Aronoff DM, Ivanov AI, Konsman JP, Steiner AA, and Turek VF.** Fever and hypothermia in systemic inflammation: recent discoveries and revisions. *Front Biosci* 10: 2193–2216, 2005.
97. **Romanovsky AA, Ivanov AI, and Shimansky YP.** Selected contribution: ambient temperature for experiments in rats: a new method for determining the zone of thermal neutrality. *J Appl Physiol* 92: 2667–2679, 2002.
98. **Romanovsky AA, Shido O, Sakurada S, Sugimoto N, and Nagasaka T.** Endotoxin shock: thermoregulatory mechanisms. *Am J Physiol Regul Integr Comp Physiol* 270: R693–R703, 1996.
99. **Rudaya AY, Steiner AA, Robbins JR, Dragic AS, and Romanovsky AA.** Thermoregulatory responses to lipopolysaccharide in the mouse: dependence on the dose and ambient temperature. *Am J Physiol Regul Integr Comp Physiol* 289: R1244–R1252, 2005.
100. **Rummel C, Barth SW, Voss T, Korte S, Gerstberger R, Hübschle T, and Roth J.** Localized vs. systemic inflammation in guinea pigs: a role for prostaglandins at distinct points of the fever induction pathways? *Am J Physiol Regul Integr Comp Physiol* 289: R340–R347, 2005.
101. **Saha S, Engstrom L, Mackerlova L, Jakobsson P-J, and Blomqvist A.** Impaired febrile responses to immune challenge in mice deficient in microsomal prostaglandin E synthase-1. *Am J Physiol Regul Integr Comp Physiol* 288: R1100–R1107, 2005.
102. **Saito S and Shingai R.** Evolution of thermoTRP ion channel homologs in vertebrates. *Physiol Genomics* In press.
103. **Sakurada S, Shido O, Fujikake K, and Nagasaka T.** Relationship between body core and peripheral temperatures at the onset of thermoregulatory responses in rats. *Jpn J Physiol* 43: 659–667, 1993.
104. **Sasaki K, Taniguchi M, Miyoshi M, Goto O, Sato K, and Watanabe T.** Are transcription factors NF- $\kappa$ B and AP-1 involved in the ANG II-stimulated production of proinflammatory cytokines induced by LPS in dehydrated rats? *Am J Physiol Regul Integr Comp Physiol* 289: R1599–R1608, 2005.
105. **Satinoff E.** Neural organization and evolution of thermal regulation in mammals. *Science* 201: 16–22, 1978.
106. **Schmidt A, Alard F, and Handrich Y.** Changes in body temperature in king penguins at sea: the result of fine adjustments in peripheral heat loss? *Am J Physiol Regul Integr Comp Physiol* 291: R608–R618, 2006.
107. **Shabtay A and Arad Z.** Reciprocal activation of HSF1 and HSF3 in brain and blood tissues: is redundancy developmentally related? *Am J Physiol Regul Integr Comp Physiol* 291: R566–R572, 2006.
108. **Simon A and van der Meer JWM.** Pathogenesis of familial periodic fever syndromes or hereditary autoinflammatory syndromes. *Am J Physiol Regul Integr Comp Physiol* 292: R86–R98, 2007.
109. **Simon E.** Ion channel proteins in neuronal temperature transduction: from inferences to testable theories of deep-body thermosensitivity. *Am J Physiol Regul Integr Comp Physiol* 291: R515–R517, 2006.
110. **Steiner AA, Chakravarty S, Robbins JR, Dragic AS, Pan J, Herkenham M, and Romanovsky AA.** Thermoregulatory responses of rats to conventional preparations of lipopolysaccharide are caused by lipopoly-saccharide per se—not by lipoprotein contaminants. *Am J Physiol Regul Integr Comp Physiol* 289: R348–R352, 2005.
111. **Steiner AA, Rudaya AY, Robbins JR, Dragic AS, Langenbach R, and Romanovsky AA.** Expanding the febrigenic role of cyclooxygenase-2 to the previously overlooked responses. *Am J Physiol Regul Integr Comp Physiol* 289: R1253–R1257, 2005.
112. **Székely M.** Orexins, energy balance, temperature, sleep-wake cycle. *Am J Physiol Regul Integr Comp Physiol* 291: R530–R532, 2006.
113. **Szelényi Z.** Neuronal CCK and thermoregulation: two receptors with different functions. *Am J Physiol Regul Integr Comp Physiol* 292: R109–R111, 2007.
114. **Talavera K, Yasumatsu K, Voets T, Droogmans G, Shigemura N, Ninomiya Y, Margolske RF, and Nilius B.** Heat activation of TRPM5 underlies thermal sensitivity of sweet taste. *Nature* 438: 1022–1025, 2005.
115. **Tanaka M and McAllen RM.** A subsidiary fever center in the medullary raphe? *Am J Physiol Regul Integr Comp Physiol* 289: R1592–R1598, 2005.
116. **Tanaka M, Owens NC, Nagashima K, Kanosue K, and McAllen RM.** Reflex activation of rat fusimotor neurons by body surface cooling, and its dependence on the medullary raphe. *J Physiol* 572: 569–583, 2006.
117. **Tankersley CG, Irizarry R, Flanders SE, Rabold R, and Frank R.** Unstable heart rate and temperature regulation predict mortality in AKR/J mice. *Am J Physiol Regul Integr Comp Physiol* 284: R742–R750, 2003.
118. **Thompson CS, Holowatz LA, and Kenney WL.** Cutaneous vasoconstrictor responses to norepinephrine are attenuated in older humans. *Am J Physiol Regul Integr Comp Physiol* 288: R1108–R1113, 2005.
119. **Togashi K, Hara Y, Tominaga T, Higashi T, Konishi Y, Mori Y, and Tominaga M.** TRPM2 activation by cyclic ADP-ribose at body temperature is involved in insulin secretion. *EMBO J* 25: 1804–1815, 2006.
120. **Vallone D, Frigato E, Vernesi C, Foà A, Foulkes NS, and Bertolucci C.** Hypothermia modulates circadian clock gene expression in lizard peripheral tissues. *Am J Physiol Regul Integr Comp Physiol* 292: R160–R166, 2007.
121. **Van Someren EJW.** Thermoregulation and aging. *Am J Physiol Regul Integr Comp Physiol* 292: R99–R102, 2007.
122. **Wade GN.** Regulation of body fat content? *Am J Physiol Regul Integr Comp Physiol* 286: R14–R15, 2004.
123. **Wechselberger M, Wright CL, Bishop GA, and Boulant JA.** Ionic channels and conductance-based models for hypothalamic neuronal thermosensitivity. *Am J Physiol Regul Integr Comp Physiol* 291: R518–R529, 2006.
124. **Weiland TJ, Voudouris NJ, and Kent S.** CCK<sub>2</sub> receptor nullification attenuates lipopolysaccharide-induced sickness behavior. *Am J Physiol Regul Integr Comp Physiol* 292: R112–R123, 2007.
125. **Wernstedt I, Edgley A, Berndtsson A, Fäldt J, Bergström G, Wallenius V, and Jansson J-O.** Reduced stress- and cold-induced increase in energy expenditure in interleukin-6-deficient mice. *Am J Physiol Regul Integr Comp Physiol* 291: R551–R557, 2006.
126. **Whyte DG and Johnson AK.** Lesions of the anteroventral third ventricle region exaggerate neuroendocrine and thermogenic but not behavioral responses to a novel environment. *Am J Physiol Regul Integr Comp Physiol* 292: R137–R142, 2007.
127. **Widmer RJ, Laurinec JE, Young MF, Laine GA, and Quick CM.** Local heat produces a shear-mediated biphasic response in the thermoregulatory microcirculation of the Pallid bat wing. *Am J Physiol Regul Integr Comp Physiol* 291: R625–R632, 2006.
128. **Zhang YH, Yanase-Fujiwara M, Hosono T, and Kanosue K.** Warm and cold signals from the preoptic area: which contribute more to the control of shivering in rats? *J Physiol* 485: 195–202, 1995.
129. **Zhao Y and Boulant JA.** Temperature effects on neuronal membrane potentials and inward currents in rat hypothalamic tissue slices. *J Physiol* 564: 245–257, 2005.