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Forebrain melanocortin signaling enhances the hindbrain satiety response to CCK-8

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Blevins JE, Morton GJ, Williams DL, Caldwell DW, Bastian LS, Wisse BE, Schwartz MW, Baskin DG. Forebrain melanocortin signaling enhances the hindbrain satiety response to CCK-8. Am J Physiol Regul Integr Comp Physiol 296: R476–R484, 2009. First published December 24, 2008; doi:10.1152/ajpregu.90544.2008.—Melanocortin 4 receptors (MC4R) are hypothesized to mediate the central nervous system actions of leptin to enhance the satiety effects of cholecystokinin (CCK). To further elucidate this mechanism, we confirmed that peripheral administration of CCK-8 is less effective in producing this effect in MC4R-deficient mice (MC4R−/−). Whereas intraperitoneal (ip) CCK-8 at 0.75 nmol/kg lean body mass (lbm) suppressed food intake in wild-type mice, CCK-8 doses of 7.5 nmol/kg lbm were required to attenuate food intake in MC4R−/− mice. To determine whether melanocortin signaling in the hypothalamic paraventricular nucleus (PVN) participates in regulating this CCK satiety response, we administered the MC3/MC4R antagonist, SHU9119, into the PVN of rats before ip CCK-8 administration. PVN administration of SHU9119 attenuated the ability of CCK-8 to reduce 30-min food intake by 20%. To determine whether MC4R are expressed by PVN neurons that project directly to hindbrain nuclei involved in the satiety response to ip CCK-8, the retrograde tracer fluorescent cholera toxin subunit B was injected into the nucleus tractus solitarius (NTS) of the hindbrain. After 4 days, labeled PVN neurons were collected by laser capture microdissection and found to express MC4R mRNA by quantitative RT-PCR analysis. These data provide evidence for a neuroanatomical link between hypothalamic melanocortin signaling in the PVN and NTS neurons that regulate food intake. These findings highlight the contribution of melanocortin signaling in the PVN toward regulating the satiety effects of CCK-8 while acknowledging that melanocortin-dependent pathways in other brain regions and/or melanocortin-independent mechanisms are also important in this mechanism.

paraventricular nucleus; retrograde tracer; cholecystokinin

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Among the many brain nuclei that contain MC4 receptors, the hypothalamic paraventricular nucleus (PVN) is especially well-positioned to mediate melanocortin regulation of CCK responsiveness. The PVN is richly supplied by axons that contain the endogenous melanocortin ligand, α-MSH (32), and PVN neuronal cell bodies express MC4 receptors (31). Moreover, both the endogenous MC3/MC4R agonist α-MSH and the synthetic MC3/MC4R agonist MTII inhibit food intake when injected into this nucleus (51).

Delineating the specific neuronal subsets that transduce input from melanocortins into heightened sensitivity to satiety signals will allow an improved understanding of energy homeostasis circuitry and may shed light on the pathogenesis of common forms of obesity. Toward this end, we sought to extend previous evidence that CNS melanocortin signaling is required for an intact satiety response induced by peripheral CCK in mice and rats. First, we compared the dose-response effects of peripheral CCK-8 on food intake in wild-type and MC4R−/− mice, which lack endogenous MC4R signaling. To test the hypothesis that endogenous melanocortin signaling in the PVN contributes to the intact satiety response to CCK, we measured the effects of PVN administration of the MC3/MC4R antagonist SHU9119 in combination with peripheral administration of CCK-8 on food intake in rats. To address the question of whether MC4R-expressing neurons in the PVN project directly to hindbrain nuclei sensitive to peripheral CCK, we injected a retrograde neuronal tracer, fluorescent choler toxin subunit B (CTB), into the nucleus tractus solitarius (NTS) of rats to label parvocellular PVN (pPVN) neurons that project directly to the NTS. We then used laser capture microdissection (LCM) in combination with quantitative RT-PCR to determine whether MC4R mRNA is expressed by pPVN neurons that project to the NTS.

MATERIALS AND METHODS

**Experimental animals.** All experimental protocols were approved by the Institutional Animal Care and Use Committee of the VA Puget Sound Medical Center and the University of Washington. Adult male Wistar rats (272–417 g) from Charles River Laboratories (Wilmington, MA) were used in the rat studies. For mouse studies, adult male Wistar rats (272–417 g) from Charles River Laboratories (Wilmington, MA) were used in the rat studies. For mouse studies, adult male Wistar rats (272–417 g) from Charles River Laboratories (Wilmington, MA) were used in the rat studies. For mouse studies, adult male Wistar rats (272–417 g) from Charles River Laboratories (Wilmington, MA) were used in the rat studies. For mouse studies, adult male Wistar rats (272–417 g) from Charles River Laboratories (Wilmington, MA) were used in the rat studies. For mouse studies, adult male Wistar rats (272–417 g) from Charles River Laboratories (Wilmington, MA) were used in the rat studies. For mouse studies, adult male Wistar rats (272–417 g) from Charles River Laboratories (Wilmington, MA) were used in the rat studies.

**Materials and methods.** Experimental treatments. SHU9119 (Bachem/Peninsula, Belmont, CA; 0.08 nmol/0.5 μl) was solubilized in saline for infusion into the PVN 1 h before the start of the dark cycle. CCK-8 (Bachem/Peninsula) was dissolved in saline with 0.1% BSA and was given as an ip injection (1 ml/kg injection volume) immediately before the start of the dark cycle, when the animals normally begin eating and when CCK-8 has its most potent effect to reduce food intake. The four treatments groups for studies in rats were as follows: 1) PVN SHU9119 vehicle (saline) administered 1 h before a dose of CCK-8 vehicle (0.1% BSA, saline), 2) PVN SHU9119 vehicle 1 h before CCK-8, 3) PVN SHU9119 1 h before CCK-8 vehicle, and 4) PVN SHU9119 1 h before CCK-8. Doses of CCK-8 and SHU9119 were selected on the basis of preliminary studies from our laboratory and published reports (10, 50, 51). We intentionally selected a dose of SHU9119 that was subthreshold for feeding effects to avoid the confounding effect of an independent feeding response from the SHU9119. For 4V administration of SHU9119, SHU9119 was administered in saline 75 min before the start of the dark cycle. Food intake studies. Food intake was measured 30 min after animals received injections of CCK-8 or vehicle and given access to food at the onset of the dark cycle as previously described (9). Examination of cage bottoms revealed negligible spillage, and there were no visually apparent differences in spillage within or between treatment groups. The dose of CCK-8 used in determining the effect of PVN administration of SHU9119 on CCK-8–elicited inhibition of food intake in rats was 1.30 nmol/kg.
Behavioral assessment of 4V cannula placement. Assessment of 4V cannula placement was done with injections of 15 pmol/2.0 μl bombesin (Bachem/Peninsula) immediately before the start of the dark cycle in rats that were fasted for 6 h. All animals used in the subsequent analysis of the data reduced their food intake by at least 20% within 1 h, an established criterion for correct placement of these cannulas. Histological verification of injection sites. Brains were frozen by submerging for 10–15 s in isopentane and then covered with crushed dry ice. Coronal cryostat sections (60 μm) at the level of the ventral extent of the cannula track were mounted on microscope slides and dried 24 h before staining with Cresyl violet. A Nikon SMZ-U ×10 stereo microscope was used to locate the most ventral portion of the scar tract made by the injector needle within the PVN, defined as the injection site. Data from an animal were excluded when the injection site extended 0.3 mm beyond the PVN boundary.

A subset of animals (n = 5) that received unilateral CTB injections into the NTS was also analyzed to validate injection site location. Coronal cryostat sections (14 μm) at the level of the ventral extent of the cannula track were mounted on microscope slides. Slides were analyzed with a Zeiss Axioplan fluorescence microscope and all injection sites were analyzed using a ×20 objective lens. Identification of anatomic landmarks was assisted by staining cell nuclei with Hoechst 33258 (Sigma, St. Louis, MO), which was added to the mounting medium and observed with a conventional DAPI filter set. CTB was clearly visible in the dorsal vascular complex of all five brains that were examined. As expected, CTB diffused throughout the dorsal vascular complex in all five of these brains and was not restricted solely to the NTS.

Laser capture microdissection and RT-PCR of cells in containing retrograde tracer. Slide-mounted cryostat sections (from unfurled rat brains) were thawed briefly, followed by graded dehydration in consecutive 1-min immersions of 75, 95, and 100% (2 ×) ethanol and xylene (1 ×) for 5 min, then air-dried for 10 min. The sections were then visualized on the Arcturus AutoPix Fluorescent laser capture microdissection (LCM) System (Molecular Devices, Union City, CA). To obtain PVN cells for PCR measurement of MC4R mRNA, slide-mounted cryostat sections prepared from unfurled brains that had previously been injected 4 days earlier with CTB retrograde tracer into the cmNTS were dehydrated and air-dried as described above. In the pPVN, ~250 neurons that were labeled with Alexa-488-labeled CTB (cells that projected to the NTS) were collected from three to four anatomically matched coronal sections of the PVN from each rat. As a negative control, 250 cells were picked by LCM from the suprachiasmatic nucleus (SCN), a region that expresses low levels of oxytocin (24) and corticotropin-releasing hormone (CRH) as well as MC4 receptors (31). In addition, ~250 neurons that did not contain fluorescent CTB were also collected from the pPVN. RNA was extracted from the captured cells using an Arcturus Picopure RNA Isolation Kit (Molecular Devices) followed by reverse transcription into cDNA using a High Capacity cDNA Archive Kit (Applied Biosystems, Foster City, CA). Analysis for MC4R, oxytocin, and CRH mRNA in the RNA extracts was performed by quantitative RT-PCR on an Applied Biosystems Prism 7000 Sequence Detection System. Glyceraldehyde-3-phosphate dehydrogenase (GAPDH) mRNA in each sample was measured as an internal standard. Each sample was measured in triplicate. Primers and probes were designed using Primer Express (version 2.0.0) from TaqMan (Applied Biosystems). The primer sequences used for RT-PCR were rat GAPDH forward primer: 5'-GCCACGCTCTGCTCATAGACA-T3'; rat GAPDH reverse primer: 5'-GTCCGATAAGCCGCAATTC-3'; rat GAPDH probe primer: VIC-5'-ATGGTGAAAGGCTTTGGT3'-; rat oxytocin, forward: 5'-TGACCTCCCGCTGTCACTC-3'; rat oxytocin reverse: 5'-AGGGAAGACATGTCGATATC-3'; rat oxytocin probe primer: 6-carboxyfluorescein (FAM)-5'-CTGGGGCCGAAGAG-3', rat melanocortin-4 receptor forward primer: 5'-CAACATGAAAGGGCG-CAATT-3'; rat melanocortin-4 receptor reverse primer: 5'-GCCCAGACGACACAAACACT-3'; rat melanocortin-4 receptor probe primer: FAM-5'-CCTTGACCTTCGATTG-3'. The probe and primer for rat CRH were acquired from Applied Biosystems (cat. no. Rn0146237_m1). PVN and SCN expression levels of MC4R, oxytocin, and CRH were normalized to GAPDH mRNA content.

The mRNA measurements in the SCN were a control to determine whether PCR data produced by LCM picking of the subpopulation of PVN cells that contained the CTB tracer from the NTS could detect meaningful levels of MC4R mRNA rather than just background levels. MC4R are known to be expressed at low levels in the PVN relative to the SCN. Thus, oxytocin and CRH mRNA, which like MC4R are highly expressed in the PVN but not in the SCN, were also probed as they would also be expected to show lower mRNA levels in the SCN, if the MC4R differences produced by the LCM/PCR procedure are valid.

Statistics. The data for each group are expressed as means ± SE. Comparisons between multiple groups in a within-subjects design were made using a two-way repeated-measures ANOVA followed by Fisher’s least-significant-difference test as a post hoc test. Comparisons between multiple groups as a between-subjects design also were made using a two-way ANOVA followed by Fisher’s least-significant-difference test as a post hoc test. Comparisons in a within-subjects design were made using a one-way repeated-measures ANOVA followed by Fisher’s least-significant-difference test as a post hoc test. Analyses were performed using the statistical program SYSTAT (Richmond, CA) and Statistica (StatSoft, Tulsa, OK). Differences were considered significant if P < 0.05. Animals that did not meet the acceptance criterion for proper cannula placement were eliminated from the analysis.

RESULTS

Effect of CCK-8 on food intake in MC4R null mice. To extend previous findings suggesting that mice lacking MC4R have reduced sensitivity to the satiety effect to a single dose of CCK on standard mouse chow (20), we determined the effects of multiple doses of CCK-8 in chow-fed wild-type and MC4R−/− mice. Our aim was to evaluate the dose-response function within each of the two groups relative to the baseline of the respective genotypes, rather than comparing the baseline differences between groups. MC4R−/− mice consumed nearly 26% less food over a 30-min period compared with wild-type mice after a 24-h fast (P < 0.05). CCK-8 produced a 22, 29, and 43% suppression of 30-min food intake in wild-type mice at doses of 0.75 (P < 0.05), 2.5 (P < 0.01), and 7.5 (P < 0.01) nmol/kg Ibm compared with the same animals that received vehicle alone (Fig. 1A). There was a significant main effect of CCK-8 to reduce 30-min food intake in wild-type mice [F(3,35) = 6.92, P < 0.01]. When administered to MC4R−/− mice, CCK-8 did not suppress 30-min food intake at 0.75 nmol/kg (P = NS), 2.5 nmol/kg (P = NS), but CCK-8 produced a 36% suppression of 30-min food intake at 7.5 nmol/kg Ibm (P < 0.01) compared with the same animals that received vehicle alone (Fig. 1B). There was a significant main effect of CCK-8 to reduce 30-min food intake in the MC4R−/− mice [F(3,33) = 4.83, P < 0.01]. Two-way repeated-measures ANOVA revealed a significant main effect on 30-min food intake for strain [F(1,16) = 7.94, P < 0.05] and CCK-8 [F(3,48) = 10.52, P < 0.01], but there was no significant interaction between strain×CCK (P = NS). Overall, the data showed that there is a decreased sensitivity to CCK-8 in MC4R−/− mice at doses that were effective in wild-type mice,
but the response to a large dose of CCK-8 in MC4R<sup>−/−</sup> mice is not attenuated.

**Effect of PVN SHU9119 and CCK-8 on food intake.** To determine whether endogenous melanocortin signaling through MC3/MC4Rs in the PVN contributes to the normal feeding response to CCK, we measured the feeding response to PVN injections of SHU9119 in combination with ip injections of CCK-8. Due to technical challenges inherent in targeting the PVN in mice, the effects of SHU9119 were examined in the PVN in rats, where the feasibility of placing bilateral cannulas in the PVN is well-established. In the presence of PVN pretreatment with SHU9119, the ability of CCK-8 to inhibit 30-min food intake was attenuated by 20% compared with controls that received PVN pretreatment with vehicle ($P < 0.05$; Fig. 2). CCK-8 inhibited 30-min food intake by 61% compared with the same animals that received PVN injections of vehicle alone ($P < 0.05$). SHU9119 alone did not increase 30-min food intake relative to animals that received PVN injections of vehicle alone ($P = \text{NS}$). Two-way repeated-measures ANOVA revealed a significant main effect of CCK-8 to inhibit 30-min food intake [$F(1,48) = 56.397, P < 0.01$], without a main effect of SHU9119 on 30-min food intake [$F(1,48) = 0.624, P = \text{NS}$]. There was a significant interactive effect of SHU9119 and CCK-8 on 30-min food intake [$F(1,48) = 4.516, P < 0.05$]. A representative photomicrograph of the injection site in the PVN and the distribution of injection sites within the PVN are shown in Figs. 3 and 4, respectively. One rat whose injection site was beyond the boundary of the PVN did not show a decreased response to CCK-8 following PVN injections of SHU9119 compared with CCK-8 alone. Two additional animals who showed an enhanced response to CCK-8 following injections of SHU9119 were deleted from the data analysis because the histological verification was compromised from bad morphology.

**Effect of 4V SHU9119 on 30-min food intake.** Our aim was to determine whether doses of SHU9119 found ineffective at stimulating 30-min food intake in the PVN were effective at stimulating 30-min food intake in nuclei outside the PVN. To test this aim, we examined the effects of 4V administration of SHU9119 at doses (0.2, 0.4 nmol) that were 1.5- to 2.5-fold higher than the total subthreshold dose of SHU9119 given bilaterally into the PVN (0.08 nmol/side or 0.16 nmol total). SHU9119 administration (0.2, 0.4 nmol) into the 4V was ineffective at stimulating 30-min food intake ($P = \text{NS}$; Table 1).

**MC4R expression in pPVN cells that project to the NTS.** To determine whether components of the MC4R signaling pathway exist in projections from the pPVN to the NTS, we initially injected retrograde fluorescently labeled (Alexa 488) tracer (CTB) into the cmNTS and waited 4 days for retrograde transport.
transport of the CTB to the PVN (10). pPVN cells that contained the CTB (representing pPVN neurons that had direct projections to the hindbrain) were collected by LCM (Fig. 5, A–C) and analyzed for MC4R mRNA content by quantitative RT-PCR. Analysis of oxytocin, and CRH mRNA expression, peptides found in neurons that project from the pPVN to the NTS (39, 40) were also analyzed as a positive control to confirm collection of pPVN neurons. Neurons in the SCN (CTB/H11002) were also collected by LCM and analyzed similarly, as a control nucleus that was expected to express relatively low levels of each of these transcripts (24, 31). In addition, unlabeled cells from the pPVN were collected and screened for MC4R mRNA, oxytocin mRNA, and CRH mRNA.

CTB+ cells in the pPVN that project to the NTS contained nearly five-, six-, and eightfold greater levels of mRNAs for MC4R (Fig. 6A), oxytocin (Fig. 6B), and CRH (Fig. 6B), respectively, compared with the SCN (P < 0.01). There were no significant differences in MC4R, oxytocin, and CRH mRNA between CTB-negative cells from the pPVN and CTB+ cells in the pPVN (P = NS; data not shown), consistent with the earlier studies reporting the pPVN to be enriched in these transcripts (27, 31, 37, 39). These findings provide evidence that pPVN neurons that directly project to the NTS contain MC4R mRNA, and mRNA for oxytocin and CRH.

**DISCUSSION**

As a first step to clarify the role of melanocortin signaling in the satiety response to CCK, we sought to extend earlier findings performed in MC4R−/− mice (20, 48). We showed here that suppression of satiety in mice required higher doses of CCK-8 in the absence of melanocortin signaling. CCK-8 administration (ip) at doses of 0.75 and 2.5 nmol/kg lbm was ineffective in reducing 30-min food intake in MC4R−/− mice, whereas the same doses of ip CCK-8 reduced food intake by 24 and 29%, respectively, in wild-type mice. In this study, only the 7.5-nmol/kg lbm dose of CCK-8 was effective in the MC4R−/− mice, resulting in a 38% suppression of 30-min food intake, comparable to the 42% suppression produced by this dose in wild-type mice.

Moreover, here we show that administration of SHU9119 into the PVN decreased the effectiveness of CCK-8-induced satiety in rats. Finding this effect now in rats as well as mice lends credence to the possible existence of a physiological mechanism in which endogenous melanocortin signaling through melanocortin-receptive neurons in the PVN contributes to the magnitude of the satiety responses to CCK that is released from the intestines during meals. Furthermore, based on their expression of MC4R mRNA, pPVN neurons that project to the region of the cmNTS are strong candidates for an anatomical link between melanocortin circuitry in the hypothalamus and hindbrain nuclei sensitive to peripheral CCK. While not ruling out the contribution of other hypothalamic circuits or the contribution of PVN projections to hindbrain areas outside the NTS [dorsal motor nucleus of the vagus (DMV), area postrema (AP)] labeled during the CTB injection process, these data support the hypothesis that melanocortin-sensitive neurons in the PVN play a physiological role to regulate the satiety effects of CCK. The present findings highlight the contribution of melanocortin signaling in the PVN toward regulating the magnitude of the satiety effects of CCK-8, but the satiety response to CCK-8 is undoubtedly also regulated by melanocortin-dependent pathways in other brain regions and/or melanocortin-independent mechanisms.

**Table 1. Effects of SHU9119 administration into the 4V on 30-min food intake**

<table>
<thead>
<tr>
<th>Treatment</th>
<th>0.2 nmol</th>
<th>0.4 nmol</th>
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<tr>
<td>Food intake</td>
<td>4.2±0.27</td>
<td>4.3±0.25</td>
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Values are means ± SE. Rats received SHU9119 injections 75 min before access to food and start of the dark cycle (n = 19/dose). Food intake was measured at 30 min following the start of the dark cycle. Each animal received each treatment at 48-h intervals.
Understanding how melanocortins influence food intake potentially has important therapeutic ramifications. Farooqi and O’Rahilly suggested that mutations in the MC4R gene underlie the most common form of monogenic human obesity (21). Consistent with previous reports (2, 20, 46, 53), the present findings strengthen the emerging hypothesis that melanocortin signaling decreases meal size through increasing the CNS responsivity to satiety signals such as CCK in both rats and mice, and raise the possibility that this may represent a generalized mechanism for regulating food intake in mammals. Furthermore, the adipocyte hormone leptin also reduces food intake via a mechanism that involves increased sensitivity to satiety signals (6, 17, 18) as well as activation of hypothalamic melanocortin-containing neurons that project to the PVN (43). Thus, the mechanism linking changes of leptin signaling to satiety signals (6, 17, 18) as well as activation of hypothalamic melanocortin-containing neurons that project to the PVN (43). This response, therefore, appears to be a poorly understood phenomenon. Our findings reveal that the MC4R-/- mice consumed nearly 26% less food over a 30-min period compared with wild-type mice after a 24-h fast, but these findings do not provide a definitive resolution to the conflicting literature. It is possible that a hyperphagic response in the MC4R-/- mice would be difficult to detect in 30 min because of the relatively small amount of food normally consumed in that time. Moreover, if the animals are eating continuously during that period, the amount of food consumed may not have exceeded 0.5–0.6 g of chow under any treatment. The present studies were not designed to address this particular issue and thus future work is required to better understand this response in MC4R-/- mice.

Although the response to other satiety-inducing stimuli in the MC4R-/- mice was not examined in the current studies, others reported intact or even heightened responses to bombesin and PYY(3–36) in these animals (20, 48). Thus, MC4R-/- mice appear to be responsive to other satiety-inducing agents and these effects appear to be relatively specific to CCK-8.

The possibility that all or a portion of the inhibition of food intake observed at the highest dose of CCK-8 (7.5 nmol/kg lbm) in both wild-type and MC4R-/- mice could be secondary to discomfort or malaise (34) cannot be ruled out. Comparable doses of CCK-8 have been reported to inhibit food intake (36) and activate neurons in the area postrema (13, 22), which has a leaky blood-brain barrier (52), contains CCK-1 receptors (22, 24), and activates areas that project to the NTS. Data represent means ± SE. A: relative MC4R mRNA levels following analysis of Alexa 488-conjugated CTB-labeled pPVN neurons by quantitative RT-PCR, with the data normalized to GAPDH mRNA levels (44). Relative MC4R mRNA levels in CTB-labeled pPVN neurons were compared with levels in suprachiasmatic nucleus (SCN) as negative control. B: relative oxytocin- and CRH-mRNA following analysis of Alexa 488-conjugated CTB-labeled pPVN neurons by quantitative RT-PCR, with the data normalized to GAPDH mRNA levels. Relative oxytocin- and CRH-mRNA levels were compared with levels in SCN as a negative control. †P < 0.01 vs. SCN.
Although injections of the Alexa 488-conjugated CTB fluorescent tracer (0.5 μl) potentially diffused to areas surrounding the NTS (DMV, AP), any particular labeled PVN neurons likely projected to the NTS, DMV, AP, or immediate surrounding area that was positioned to uptake the CTB fluorescent tracer. The findings support the conclusion that some of the labeled PVN neurons project to the NTS and that this is consistent with literature showing that PVN in this area has direct projections to the NTS, and that the NTS has a rich innervation of oxytocin and CRH neurons from the PVN (37, 39, 40).

The key findings of this study support a mechanism by which endogenous melanocortin signaling to the PVN contributes to the normal satiety response to CCK through activation of a descending PVN-NTS projection. The PVN contains a population of MC4R-expressing cells (31) which receives endogenous melanocortin signaling through proopiomelanocortin (POMC) projections from the ARC (15). In addition, MC4Rs are expressed by pPVN neurons that project to the cmNTS, providing an anatomical link between hypothalamic MC4Rs and hindbrain nuclei sensitive to peripheral CCK. Moreover, PVN administration of MTII and α-MSH reduces food intake at doses that do not cause an aversive response (51); this indicates MC4R activation in the PVN is sufficient to inhibit food intake. Restoration of MC4R into the PVN of mice that do not express MC4R attenuates their hyperphagia and obesity (5). Thus, the findings presented here fit well with the emergent hypothesis that melanocortin signaling through MC4Rs in the PVN may be a critical component of CNS circuitry that regulates the magnitude of normal satiety responses to CCK.

The findings reported here may be relevant to understanding the CNS mechanism by which leptin reduces food intake because the melanocortin signaling in the hypothalamus is implicated as a link in the transduction of hypothalamic leptin action to the activation of descending neuronal pathways to the NTS, where meal size is regulated. It has been established that leptin activates POMC neurons in the ARC (42), some of which have projections to the PVN (13, 16) while others project to the NTS (32, 53). Electrophysiological studies indicate that individual neurons in the pPVN respond similarly to both α-MSH and MTII, and the specificity of these effects was verified by blockade by AGRP and SHU9119, respectively (13). Benoit et al. (7) reported that administration of either the selective MC4R agonist R027–3225 or MTII into the third ventricle produced a similar pattern of neuronal activation in the PVN, consistent with a role for MC4R in the actions of these ligands within the PVN. Evidence that POMC projections from the ARC also directly innervate the NTS (32, 53) raises the possibility that other melanocortin circuits, in addition to those originating from the PVN, may also play a role in regulating the hindbrain response to satiety signals as well as the interaction of these inputs with leptin signaling to the CNS.

The observation in the present study that SHU9119 administration into the PVN did not completely block the food intake response to CCK-8 to the levels seen in control animals can most likely be explained by the fact that melanocortin signaling through MC4R in other brain nuclei such as the NTS is also important in regulating the satiety response to CCK (20, 46, 53). Furthermore, the dose of SHU9119 that was bilaterally injected into the PVN was unlikely to have accessed all MC4Rs in the entire PVN; some PVN MC4Rs were likely unaffected or not reached by the injected compounds. In addition, the regulation of CCK-induced satiety mechanisms undoubtedly involves melanocortin-independent pathways that would not have been directly affected by these interventions. However, these effects are unlikely to be explained by the MC3/MC4R antagonist acting in areas outside the PVN, including the hindbrain, which also contains MC4Rs, as administration of SHU9119 administered into the 4V at doses higher than delivered to the PVN failed to stimulate food intake.

If melanocortin signaling through pPVN neurons links the ability of 3V leptin to interact with CCK-sensitive neurons at the level of the hindbrain, oxytocin (10, 25), CRH (27), thyrotropin-releasing hormone (TRH) (1), and gastrin-releasing peptide (GRP) (26) neurons in the pPVN would be logical candidates to mediate this effect. TRH neurons and GRP neurons were reported to be activated after leptin administration into the 3V (1, 26). We reported previously that 3V leptin treatment increased CRH mRNA content in rat PVN (41). Moreover, 3V infusion of a CRH receptor antagonist blocked the effect of leptin to reduce food intake (47) and body weight. Recent findings link MC4R expression to CRH neurons in the pPVN (27), some of which have known direct projections to the NTS and DMV as well as spinal cord (39).

Perspectives

These data are consistent with the hypothesis that melanocortin signaling regulates the satiety effects of CCK and that under defined experimental conditions, MC4R neurons in the PVN contribute to the satiety response to a pharmacological dose of CCK in rats. Furthermore, these data indicate that the satiety effects of CCK-8 are also regulated by melanocortin-dependent mechanisms in mice. The present findings highlight the contribution of melanocortin signaling in the PVN toward regulating the magnitude of the satiety effects of CCK-8, but the satiety response to CCK-8 is undoubtedly also regulated by melanocortin-dependent pathways in other brain regions and/or melanocortin-independent mechanisms. The present findings also provide evidence that MC4R mRNA is expressed by pPVN neurons that project to the cmNTS, providing a functional circuit to link hypothalamic melanocortin-responsive neurons that are sensitive to peripheral CCK-8 and putatively influence meal size. Since the hypothalamic melanocortin system is implicated as a key mediator of leptin’s anorexigenic action in the brain, the present work suggests that melanocortin-sensitive neurons that project to the NTS regions from the medial parvocellular region of the PVN contribute to leptin’s ability to enhance the satiety effect of CCK.

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