Follicle-stimulating hormone, interleukin-1, and bone density in adult women

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Although the average age at menopause is ~51 years, changes in hormonal concentrations and cycle regularity may begin more than 15 years earlier (18). Follicular-phase serum follicle-stimulating hormone (FSH) concentrations can increase by more than twofold between the ages of 33 and 45 years (43). Klein et al. (23) have reported that serum FSH concentrations of 40- to 45-yr-old women were higher throughout the menstrual cycle, and ~70% higher in the early follicular phase, than in 20- to 25-yr-old women, but estradiol concentrations were similar between the two age groups. Beyond age 45, early follicular FSH concentrations increase as much as fivefold, reaching ~50–70 mIU/ml by the time of the final menstrual period, whereas estradiol concentrations fall by ~50–60% (6, 38). Several studies have indicated that bone mineral density (BMD) also begins to change years before the last menstrual period (17, 38, 41). Reductions in BMD have been correlated with increases in serum FSH concentrations in perimenopausal women, but not with any changes in estradiol during this period (42).

Several cytokines influence the balance between the bone-resorbing activity of osteoclasts and the bone-forming activity of osteoblasts. In vitro studies have shown that interleukin (IL)-1 stimulates the differentiation and bone-resorbing activity of osteoclasts (21). IL-1 also affects osteoblasts by inhibiting their bone matrix synthetic activity (11) and stimulating their secretion of IL-6 (26). IL-6, in turn, promotes development of osteoclasts and stimulates bone resorption (29). Tumor necrosis factor (TNF)-α promotes the fusion of monocytes into osteoclasts while inhibiting differentiation of osteoblasts from progenitor cells (32). In vivo, administration of either the naturally occurring IL-1 receptor antagonist (IL-1Ra) or soluble TNF receptors inhibited bone resorption in mice made osteopenic by ovariectomy (22). Antibodies against IL-6 blocked osteoid development in marrow cultures taken from ovariectomized mice (20), whereas IL-6-deficient mice were not susceptible to osteoporosis by ovariectomy (35).

Both progesterone and estradiol have biphasic dose-response influences on cytokine production. In the concentration ranges of the normal menstrual cycle, exogenous progesterone enhanced, and exogenous estradiol inhibited, IL-1β and TNF-α secretion in vitro (14, 31, 34). Furthermore, spontaneous IL-1β secretion from isolated human monocytes was positively related to endogenous progesterone and negatively related to endogenous estradiol concentrations of the women from whom the cells were isolated (7). The influence of gonadotropins on cytokine secretion has not been as extensively studied, but there are reports that FSH enhanced IL-1β and IL-2 secretion by isolated peripheral blood mononuclear cells (24, 25).

Based on these observations, we hypothesized that FSH influences BMD, in part, by affecting the activity of bone-resorbing cytokines, either by inducing their secretion or by altering their receptor expression. Therefore, the purpose of the present study was to determine in adult women if 1) endogenous FSH concentrations were related to cytokine concentrations and cytokine receptor expression in vivo, 2) if cytokine secretion or receptor shedding were inducible by FSH in vitro, and 3) if any of these measures of cytokine status were associated with BMD. Physical activity, lean and fat mass, and the adipocyte-derived hormone leptin were also assessed to control for other factors that may affect BMD.

MATERIALS AND METHODS

Subjects

Thirty-six women between 20 and 50 yr of age who were experiencing natural menstrual cycles (although these cycles may have become variable) were recruited. Exclusion criteria included smoking.
pregnancy, lactation, or a history of serious diseases such as diabetes, liver disease, cardiovascular disease, or cancer. Additional exclusion criteria included use of any hormone treatment or prescription drug in the past 3 mo.

Subjects reported to the laboratory between 7:30 and 9:00 A.M., having fasted for at least 8 h. Information regarding their menstrual history was collected using the baseline interview questionnaire developed for the longitudinal, multisite “Study of Women’s Health Across the Nation” (SWAN) (44). Physical activity was assessed using the International Physical Activity Questionnaire (9). This was followed by blood sample collection and body composition analysis. All procedures were approved by the Human Investigation Review Board at the Medical College of Georgia.

Body Composition

Body composition was measured using dual-energy X-ray absorptiometry (Hologic Discovery W Densitometer; Hologic, Bedford, MA). Each scan was compartmentalized and analyzed using Hologic whole body software for total BMD, fat-free soft tissue, and percent body fat. Separate scans at the hip and lumbar spine were performed and analyzed for BMD using site-specific Hologic Scan/Analysis software protocols.

Blood Collection and Leukocyte Isolation

Blood was drawn by venipuncture of the antecubital vein and collected into a 7-ml untreated tube for serum and a 6-ml EDTA-treated tube for plasma. The serum and plasma were aliquotted and frozen at −70°C for later analysis. Three 10 ml sodium heparin-treated tubes were drawn, and the mononuclear cells were isolated by density gradient centrifugation for flow cytometry and cell culture.

Reproductive Hormone Assays

Serum concentrations of FSH, luteinizing hormone (LH), progesterone, estradiol, inhibin-A, and inhibin-B were performed by the CLASS laboratory: the core laboratory for SWAN. The CLASS laboratory is located in the Department of Epidemiology, University of Michigan, Dr. Daniel S. McConnell, Director.

Serum FSH concentrations were measured with a two-site chemiluminescence (sandwich) immunoassay using two antibodies specific for the intact FSH molecule. Results were determined from a calibration curve using standards obtained from Bayer Diagnostics, referenced to the World Health Organization 2nd IRP 78/549 standard. The minimum detectable concentration was 0.3 mIU/ml. Inter- and intra-assay coefficients of variation (CV) were 8.1 and 3.5%, respectively.

Serum LH concentrations were measured with a two-site chemiluminescence immunoassay by Bayer diagnostics. Two monoclonal antibodies, one directed at the α-subunit and the other at the β-subunit, with specificity for intact LH are employed. The minimum detectable concentration is 0.07 mIU/ml. Inter- and intra-assay CV were 6.8 and 2.8%, respectively.

Serum estradiol concentrations were measured with a competitive chemiluminescence immunoassay [ACS: 180 (E2–6), Bayer Diagnostics, Tarrytown, NY]. This assay is standardized analytically and confirmed by gas chromatography–mass spectrometry (GC-MS). The minimum detectable concentration is 1 pg/ml. Closely related compounds such as estriol show insignificant cross-reactivity. Inter- and intra-assay CV are 13.8 and 8.5%, respectively. The serum progesterone assay is also a competitive chemiluminescence immunoassay. Progesterone standards have been confirmed by GC-MS (interassay CV: 7.30%, intra-assay CV: 3.0%).

Serum inhibin-A and inhibin-B concentrations were measured with enzyme-linked immunosorbent assays (ELISAs) from Diagnostic Systems Laboratories. For the inhibin-A assay, the minimum detectable concentration is 10 pg/ml, the interassay CV is 13.4%, and the intra-assay CV is 11.2%. For the inhibin-B assay, the minimum detectable concentration is 7 pg/ml, the interassay CV is 6.7%, and the intra-assay CV is 4.6%.

In Vitro Cytokine Secretion

Mononuclear cells were isolated by density gradient centrifugation using ficoll-hypaque (Sigma), washed three times with sterile saline (0.9% NaCl), and resuspended in phenol red-free RPMI-1077 medium supplemented with 2 mM of t-glutamine, 100 U/ml penicillin, 100 μg/ml streptomycin, 1% (by volume) 1 M HEPES buffer (all from Sigma, St. Louis, MO), and 1% heat-inactivated autologous plasma. Cells were then distributed in 24-well polystyrene plates (Corning Glass Works, Corning, NY) at 2.5 × 10⁶ cells/well and incubated with 0, 10, 50, and 100 mIU/ml of highly purified pharmaceutical grade human FSH (Bravelle; Ferring Pharmaceuticals, Suffern, NY) for 18 h at 37°C in a humidified 5% CO2 atmosphere. After incubation, cell supernatants were collected and centrifuged at 10,000 rpm for 1 min to remove any aspirated cells, then aliquotted and frozen at −70°C for later cytokine analysis.

Cytokine Analysis

Concentrations of IL-1β, TNF-α, IL-6, and soluble cytokine receptors (IL-1R type I & II, TNFR type I & II, IL-6Rα and gp130) were measured in plasma, serum, and supernatants using a cytometric bead array (BD Biosciences, San Jose, CA). This format employs multiple populations of microbeads, each with unique fluorescence characteristics. Each bead population is coated with capture antibodies raised against specific cytokines or soluble receptors. The beads are then mixed into the sample along with detection antibodies raised against each analyte that are labeled with compounds that fluoresce at a different wavelength than the beads. After incubation and washing, the beads are resuspended and analyzed with a BD FACSArray flow cytometer. Analytes are differentiated by bead fluorescence characteristics, and analyte concentration is measured by the fluorescence intensity of the labeled detection antibody, which is compared with standard curves generated with recombinant analytes. All assays had detection limits (based on 95% confidence intervals over blank) of <5 pg/ml, except gp130 (80 pg/ml). The intra-assay CVs were between 4 and 10%; the interassay CVs were between 4 and 15%.

Plasma leptin concentrations were determined by two-site ELISA using antibodies (MAB398, BAMD398) from R & D Systems (Minneapolis, MN). The detection limit was 0.06 ng/ml, and inter- and intra-assay variability was 5% and 2%, respectively.

Flow Cytometric Analysis of Receptor Expression

FSH receptors. Mononuclear cells were washed, resuspended to 1 × 10⁷ cells/ml, blocked with 1 mg/ml human IgG (Sigma I8640) for 30 min, and then stained with polyclonal rabbit anti-FSHR (PAI-23545;}

<table>
<thead>
<tr>
<th>Table 1. Subject physical characteristics and hormonal concentrations</th>
<th>All Subjects</th>
<th>≤45 yr old</th>
<th>&gt;45 yr old</th>
<th>P Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>FSH, mIU/ml</td>
<td>27.3 ± 46.4</td>
<td>11.1 ± 3.4</td>
<td>94.7 ± 77.0</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>LH, mIU/ml</td>
<td>9.0 ± 10.7</td>
<td>5.7 ± 0.6</td>
<td>22.7 ± 18.4</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>Estradiol, pg/ml</td>
<td>80.1 ± 112</td>
<td>91.8 ± 121</td>
<td>31.8 ± 30.8</td>
<td>0.002</td>
</tr>
<tr>
<td>Progesterone, ng/ml</td>
<td>0.59 ± 0.36</td>
<td>0.61 ± 0.34</td>
<td>0.53 ± 0.46</td>
<td>0.37</td>
</tr>
<tr>
<td>Inhibin-B, pg/ml</td>
<td>53.8 ± 36.4</td>
<td>68.0 ± 33.4</td>
<td>&lt;10</td>
<td>0.007</td>
</tr>
<tr>
<td>Height, cm</td>
<td>163.3 ± 64.0</td>
<td>1.64 ± 0.01</td>
<td>1.62 ± 0.02</td>
<td>0.40</td>
</tr>
<tr>
<td>Weight, kg</td>
<td>69.3 ± 3.1</td>
<td>69.1 ± 3.0</td>
<td>72.2 ± 9.5</td>
<td>0.16</td>
</tr>
<tr>
<td>Body fat, %</td>
<td>3.8 ± 8.2</td>
<td>3.4 ± 7.7</td>
<td>5.0 ± 10.5</td>
<td>0.66</td>
</tr>
<tr>
<td>LBMI, kg/m²</td>
<td>15.6 ± 0.37</td>
<td>15.5 ± 2.0</td>
<td>16.0 ± 3.1</td>
<td>0.58</td>
</tr>
<tr>
<td>LTPA, min</td>
<td>159 ± 172</td>
<td>155 ± 171</td>
<td>174 ± 187</td>
<td>0.80</td>
</tr>
</tbody>
</table>

Values are means ± SE. FSH, follicle-stimulating hormone; LH, luteinizing hormone; LBMI, lean body mass index = lean body mass (kg)/height² (m); LTPA, leisure time physical activity. The P value pertains to the difference between the two age groups.
Affinity BioReagents, Golden, CO) followed by phycoerythrin (PE)-conjugated F(ab")_2 fragment goat anti-rabbit IgG (Jackson ImmunoResearch Laboratories, West Grove, PA). Rabbit IgG (Jackson ImmunoResearch Laboratories) followed by the PE-conjugated secondary reagent, was used to test for nonspecific staining. Monocytes were identified with an allophycocyanin-conjugated anti-human CD14 (BD Biosciences, San Diego, CA) that was added to the cells at the same time as the rabbit anti-FSHR. After being washed, the cells were fixed in 5% formalin/PBS, pH 7.3, and stored overnight at 4°C. The following day, flow cytometric analysis was performed with a Becton Dickinson two-laser, four-detector FACSCalibur Flow Cytometer. Cell counts were restricted to intact cells identified by a dual parameter plot of forward scatter vs. side scatter. Multicolor flow cytometry was performed with appropriate one-color controls for setting compensation for spectral overlap. Additional isotype controls were run to test for nonspecific binding.

Cytokine receptors. IL-1 receptor type I (IL-1RI, CD121a) and type II (IL-1RII, CD121b), tumor necrosis factor receptor type I (TNFRI, CD120a) and type II (TNFRII, CD120b), IL-6 receptor (IL-6R, CD126), and the IL-6 receptor (gp130, CD130) were identified by Fig. 2. Bone mineral density (BMD) vs. serum FSH concentration. Solid line, regression for all subjects; broken line, regression for subjects ≥45 years of age. A: total BMD, all subjects: \( r = -0.38, P = 0.023, \geq 45 \text{ yr}; r = -0.44, P = 0.02, \text{yr}; r = -0.41, P = 0.01, \leq 45 \text{ yr}; r = -0.42, P = 0.03, \text{yr}; r = -0.36, P = 0.03, \geq 45 \text{ yr}; r = -0.37, P = 0.05, \text{yr}; r = -0.52, P = 0.001, \leq 45 \text{ yr}; r = -0.31, P = 0.10. \) Women with regular cycles tested within the first 8 days of the follicular phase are indicated by circles; perimenopausal women with irregular cycles are indicated by triangles.
one-color flow cytometry using PE-conjugated monoclonal antibodies (BD Biosciences). Monocytes were differentiated from lymphocytes in the isolated mononuclear cell preparations by forward- and side-scatter characteristics.

Because of unavoidable variations in blood collection volumes and isolated cell yields, not all analyses were possible in all subjects.

Data Analyses

All statistical analyses were performed using Statview statistical software (SAS, Cary, NC). Hormone and cytokine concentrations were normalized by log transformation as indicated. Results are reported as means ± SE. Standardized regression coefficients (b) are reported for independent factors in multiple regressions. A P value of <0.05 was considered statistically significant.

RESULTS

Subject Characteristics

Physical characteristics and hormonal concentrations for the subjects are summarized in Table 1, and the relationship of reproductive hormone concentrations to age is shown in Fig. 1. Overall, serum FSH concentrations correlated positively with age (r = 0.65, P < 0.0001). These concentrations increased at a rate of 0.13 mIU·ml⁻¹·yr⁻¹ up to age 45 (r = 0.37, P = 0.047, regression line in Fig. 1A), and increased at a much steeper rate after that. Serum LH did not exhibit any significant change up to age 45, but, because of the increased concentrations in women >45 yr of age, a statistically significant increase was detected overall (r = 0.42, P = 0.01). All but two women were tested within the first 8 days of their follicular phase. These two women had prolonged/irregular cycles but were judged to be in follicular phase based on serum progesterone concentrations (shown in Figs. 1 and 2, triangles).

Serum estradiol concentration exhibited an age-related increase up to age 45 (r = 0.51, P = 0.004, Fig. 1B) and decreased after that age. Serum progesterone concentrations exhibited no association with age. Mediators of ovarian feedback control of pituitary FSH secretion, inhibins A and B, were also measured. Inhibin A, the secretory product of developing antral follicles during the luteal phase, was undetectable in the majority of samples from the follicular phase women of this study. Inhibin B, the secretory product of granulosa cells in early follicular development, tended to decrease with age. Although this trend was not statistically significant by linear regression, the concentrations were significantly lower in women over 45, compared with younger women (Table 1).

FSH and Bone Density

Serum FSH concentration was inversely related to each of four measures of BMD by univariate regression (Fig. 2). This figure also shows that correlations with total BMD and femoral neck were statistically significant even if the seven subjects >45 yr of age, with extremely high FSH levels (>25 mIU/ml), were excluded from analysis. The serum concentrations of leptin correlated with total BMD (r = 0.37, P = 0.04), femoral neck BMD (r = 0.39, P = 0.02), and with percent body fat (r = 0.76, P < 0.0001).

Stepwise multiple regressions were performed to test FSH in context with other independent variables that might influence BMD, namely estradiol, progesterone, LH, age, lean body mass index, percent body fat, and leisure time physical activity. Separate regressions were performed for BMD at each skeletal site. Using this approach, serum FSH was significantly related to total and lumbar BMD (Table 2). In addition, lean body mass was positively associated with total and femoral neck BMD, namely estradiol, progesterone, LH, age, lean body mass index, percent body fat, and leisure time physical activity.

Table 2. Relationships of several measures of bone mineral density (dependent variables) with serum reproductive hormone concentrations, body composition, and physical activity by stepwise multiple regression

<table>
<thead>
<tr>
<th>Dependent Variable</th>
<th>FSH</th>
<th>LH</th>
<th>P</th>
<th>E</th>
<th>InhB</th>
<th>Age</th>
<th>LBMI</th>
<th>Fat, %</th>
<th>LTPA</th>
<th>Model (r², P)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total BMD</td>
<td>-0.26</td>
<td>0.34</td>
<td></td>
<td>0.44</td>
<td>0.52</td>
<td>0.65, &lt;0.0001</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total hip</td>
<td>-0.41</td>
<td>0.32</td>
<td></td>
<td>0.32</td>
<td>0.23, 0.01</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Femoral neck</td>
<td>-0.42</td>
<td>0.32</td>
<td></td>
<td>0.31, 0.002</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total lumbar</td>
<td>-0.52</td>
<td></td>
<td></td>
<td></td>
<td>0.27, 0.001</td>
<td></td>
<td></td>
<td></td>
<td></td>
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</tr>
</tbody>
</table>

Values are standardized regression coefficients for each statistically significant independent variable. The last column of the table provides the r² and P values for each of the overall regression models. Independent variables: FSH, LH, progesterone (P), estradiol (E), inhibin-B (InhB), age, LBMI, %fat, and LTPA.
BMD, and leisure time physical activity was positively associated with total and hip BMD (Table 2). Estradiol, LH, and inhibin-B were not significantly related to any of the measures of BMD.

**FSH-Induced Cytokine Secretion**

The ability of FSH to induce secretion of cytokines and soluble receptors was tested by incubating isolated mononuclear cells with 0, 10, 50, or 100 mIU/ml of FSH. As the dose-response curves in Fig. 3 show, FSH-induced increases in IL-1β and TNF-α secretion were observed inconsistently (IL-6 secretion was similar, but not shown). When a response was observed, it was usually biphasic with a maximally effective FSH dose of 10 or 50 mIU/ml, with a diminished response at 100 mIU/ml. The secretion rates of soluble receptors for IL-1, TNF, and IL-6 were not significantly affected by FSH in vitro.

Further analysis indicated that responsiveness to FSH was related to FSH receptor expression on CD14 monocytes. Representative flow cytometry data are shown in Fig. 4. The percentage of monocytes expressing FSH receptors ranged from 5 to 48% and was not related to age, individual reproductive hormone concentrations, or the anthropomorphic characteristics of the women. However, FSH receptor expression was related to an interaction between serum FSH and estradiol concentrations (log FSH: \( b = -0.84, P = 0.015 \); estradiol: \( b = -1.99, P = 0.008 \); log FSH × estradiol: \( b = 1.80, P = 0.006 \); overall: \( R^2 = 0.31, P = 0.04 \)). The positive correlations between FSH receptor expression and FSH-induced cytokine secretion (quantified as the area under the dose-response curves) are shown in Fig. 5. Consistent with these in vitro data, endogenous (serum) concentrations of FSH correlated with circulating concentrations of IL-1β (\( r = 0.40, P = 0.02 \); Fig. 6), TNF-α (\( r = 0.38, P = 0.02 \)), and IL-6 (\( r = 0.35, P = 0.04 \)).

**Cytokine Ligands, Receptors, and Bone Density**

None of the individual measures of cytokine receptor expression, secretion, or circulating concentrations were significantly related to any measure of BMD. However, the ratio of plasma IL-1β/IL-1Ra was inversely related to BMD if two apparent “outliers” were excluded (Fig. 7). These two women reported leisure time physical activity well above the mean value of 159 min/wk. Monocytes isolated from these and other women in the top tertile of physical activity (Fig. 7) expressed 20% lower levels of the signal-transducing IL-1RI and 20% higher levels of the nonsignaling (decoy) IL-1RII. These differences were not statistically significant themselves, but the difference in the ratio of IL-1RI/IL-1RII (signaling/decoy) receptors was significant (\( P = 0.01 \), Fig. 8). The univariate correlations between IL-1β/IL-1Ra ratio and BMD at all four skeletal sites (with inclusion and exclusion of the most active women) are presented in Table 3. In addition, the rate of IL-1 receptor shedding in vitro correlated significantly with the level of physical activity (Fig. 9). Analyses of TNF-α and IL-6 in combination with their receptors (both membrane-
Associations With Body Fat

A very strong correlation between plasma IL-1Ra and percent body fat was observed ($r = 0.78$, $P < 0.0001$). A similar correlation was observed between plasma IL-1Ra and plasma leptin ($r = 0.70$, $P < 0.0001$; Fig. 10). This corresponded with leptin, a secretory product of adipose cells, correlating very highly with percent body fat ($r = 0.76$, $P < 0.0001$).

DISCUSSION

Hormonal Changes With Age

The steady increase in serum FSH concentrations up to age 45, followed by a steeper increase in later years (Fig. 1), is consistent with a 14-yr longitudinal study by Sowers et al. (43), who identified an inflection point in the age-related rise in follicular-phase FSH concentrations 7 yr before the final menstrual period, at a mean chronological age of 43.6 yr. Although FSH concentrations change significantly over the course of the menstrual cycle, increased concentrations of FSH in the follicular phase of older women are accompanied by an upward displacement of FSH concentrations at all other phases of the menstrual cycle as well (23, 40).

FSH and Bone Density

The inverse relation between BMD and serum FSH (Fig. 2) is in accord with previous observations in pre- and perimenopausal women (42, 44). In addition, a meta-analysis of ten prospective studies found that the rate of spinal BMD loss during perimenopause was greater than the rate of loss in the years following menopause, when estradiol concentrations were much lower (37). Although these observations imply a dominant influence of FSH (relative to estradiol) on BMD during the menopausal transition, osteoporosis can also occur.

Table 3. Associations of bone mineral density with $IL-1\beta$ to $IL-Ra$ ratio

<table>
<thead>
<tr>
<th></th>
<th>All Subjects</th>
<th>LTPA $&lt;180$ min/week*</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>$r$</td>
<td>$P$</td>
</tr>
<tr>
<td>Total</td>
<td>$-0.11$</td>
<td>$0.55$</td>
</tr>
<tr>
<td>Hip</td>
<td>$-0.18$</td>
<td>$0.31$</td>
</tr>
<tr>
<td>Femoral neck</td>
<td>$-0.30$</td>
<td>$0.09$</td>
</tr>
<tr>
<td>Lumbar</td>
<td>$-0.36$</td>
<td>$0.04$</td>
</tr>
</tbody>
</table>

*Subjects reporting leisure time physical activity of $<180$ min per week.
in other situations (such as hypothalamic amenorrhea and administration of gonadotropin-releasing hormone antagonists) where FSH concentrations are low (reviewed in Ref. 36).

An influence of FSH on bone homeostasis has also been observed in animal studies. Sun et al. (45) reported that mice lacking the β-subunit of the dimeric FSH molecule, or lacking FSH receptors, were severely hypogonadal but maintained bone mass, indicating that a loss of estrogen did not induce bone resorption. In the same study, these authors detected FSH receptors on osteoclast precursor cells derived from the bone marrow of FSH receptor-replete mice as well as human monocyte-derived osteoclasts and found that FSH stimulated osteoclast differentiation and bone resorption by these cells in vitro. We have previously reported that immunoreactive FSH receptor extracted from human mononuclear cells migrated in gel electrophoresis in parallel with purified FSH receptor isolated from rat ovaries, with a molecular weight of ~75 kDa (8).

**Monocyte sensitivity to FSH.** The magnitude of FSH-induced cytokine secretion in vitro was proportional to the level of FSH receptor expression detected on monocytes by flow cytometry (Fig. 5). The variation in FSH receptor expression was not related to any individual hormone, but to the interaction between FSH and estradiol, which is in keeping with the rather complex hormonal regulation of FSH receptor expression described in the literature. Low doses of FSH delivered by osmotic minipump to estrogen-maintained hypophysectomized rats increased FSH receptor mRNA, whereas an additional high dose given as a bolus injection caused a rapid decrease (27). Administration of FSH alone to hypophysectomized rats increased FSH receptor expression (determined by ligand binding), whereas estradiol alone had no effect (39). In the same study, a more rapid increase was observed when both hormones were given together, however, longer exposures to estradiol ultimately resulted in reduced FSH receptor expression. Several growth factors, including members of the transforming growth factor-β family and epidermal growth factor, also influence FSH receptor expression on rat granulosa cells (13). In middle-aged women, ovarian FSH receptor expression was greatest during the late follicular phase of the menstrual cycle (30). In perimenopausal women, high serum FSH concentrations were associated with diminished ovarian FSH receptor expression (48). Conflicting data are reported for FSH receptor expression by nonovarian cells: FSH inhibited FSH receptor promoter activation in osteoclast precursor cells (49), but FSH receptor expression was increased in the bile duct epithelial cells of rats treated with FSH (28). In the present study, the expression of FSH receptors was not related to the expression of any of the cytokine receptors.

The biphasic dose response observed for FSH in vitro (Fig. 3) was similar to the results of previous reports of FSH actions on leukocytes: 10 mIU/ml FSH augmented IL-2-induced proliferation of lymphocytes, but the response was diminished at 100 mIU/ml (3). FSH also enhanced endotoxin-induced IL-1β secretion and phytohemagglutinin-induced IL-2 secretion in a biphasic manner, with maximal effects achieved in the range of 5–25 mIU/ml (25).

**Cytokines and Bone Density**

The inverse relationship between IL-1Ra and BMD (Fig. 7) is consistent with its role as a natural antagonist. IL-1Ra is approximately the same size as IL-1β but has limited sequence homology; it binds to both IL-1 receptor types but does not induce signal transduction (10). Pacifici et al. (33) reported that hormone replacement therapy ameliorated bone loss in postmenopausal women, in part, by reducing the ratio of IL-1β/
IL-1Ra secreted by monocytes. Abrahamsen et al. (1) also reported that bone loss in perimenopausal women was inversely related to serum IL-1Ra.

The relative expression and shedding of IL-1 receptors was related to physical activity levels (Figs. 8 and 9). The expression of signal-transducing IL-1RII was diminished relative to the expression of IL-1RII on monocytes isolated from active women. IL-1RII is characterized as a “decoy” receptor because it binds IL-1β but it does not transduce a signal across the membrane of the target cell, resulting in reduced responsiveness to IL-1β (10). This differential receptor expression may be significant to bone homeostasis since a subpopulation of circulating monocytes are precursors for osteoclasts (4). Thus receptor expression on circulating monocytes might serve as an estimate of osteoclast responsiveness to IL-1β. Soluble receptors result from enzymatic cleavage of membrane-bound receptors, as well as secretion of receptors coded by alternatively spliced mRNA transcripts (12). Soluble IL-1 receptors (both type I and II) intercept IL-1β in the extracellular fluid and render it unable to bind on a target cell. Receptor shedding also results in reduced receptor availability on the membranes of target cells themselves.

Body Fat, Leptin, and Bone Density

It is well-recognized that increased body mass promotes and preserves BMD. Whereas lean mass influences BMD through mechanical loading, fat mass may also be influential via metabolic or endocrine mediators, such as leptin. Significant, positive associations between circulating leptin concentrations and BMD have been reported for women (5, 16, 46), but less so for men (46). Such associations might be explained mechanistically by in vitro data indicating that leptin enhances human bone marrow cell differentiation into osteoblasts (47) and inhibits human blood mononuclear cell differentiation into osteoclasts (19). Current data (Fig. 10) support the concept that leptin may also indirectly affect bone homeostasis by increasing IL-1Ra, thereby inhibiting the osteoresorptive activity of IL-1β. The observed correlation between leptin and IL-1Ra is consistent with in vitro evidence that leptin stimulates monocytes to secrete IL-1Ra (15). IL-1Ra, in turn, inhibits the bone-resorbing activity of IL-1β (2).

Perspectives and Significance

The present study indicates that BMD is inversely related to circulating FSH concentrations in women throughout their reproductive years. This relationship may be due in part to direct effects of FSH on osteoclasts, but the present data indicate that FSH also stimulates monocytes to secrete IL-1β, which can activate osteoclasts and promote bone resorption. Moreover, in addition to benefits provided by loading the skeleton, fat mass and physical activity may also preserve bone mass by inhibiting the osteoresorptive activity of IL-1β through induction of IL-1Ra and modification of IL-1 receptor expression. The associative observations of the current study provide a rationale for conducting weight (fat) loss and exercise training studies to determine if such interventions actually cause changes in the expression or secretion of factors that regulate IL-1β.

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GRANTS

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