Response of cardiac sympathetic nerve activity to intravenous irbesartan in heart failure

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Submitted 19 November 2009; accepted in final form 4 February 2010

Heart failure (HF) is associated with intense neurohumoral activation, in particular, increased sympathetic nerve activity (SNA) and activation of the renin-angiotensin system (RAS). Although activation of these systems is initially beneficial, in the long term these changes are detrimental (2, 24). Drugs that decrease the deleterious effects of increased circulating ANG II in the brain (12, 26), where it has been shown to act to increase CSNA (35).

To address these questions, we have examined the effect of intravenous infusion of irbesartan on directly recorded CSNA in HF patients. As with other ARBs, there is evidence that irbesartan can cross the blood-brain barrier, although its ability to do so is less than that of losartan (4, 26). Thus we examined the effect of short-term infusion of irbesartan to minimize any central effects due to its crossing of the blood-brain barrier. Furthermore, there is evidence that chronic administration of ARBs in HF patients improved baroreflex control of heart rate (HR) (33), and short-term angiotensin receptor blockade in an experimental animal model of HF sensitized the baroreflex control of HR (22). The effects of RAS inhibition on CSNA have been assessed using 123I-meta-iodobenzylguanidine scintigraphy or cardiac norepinephrine spillover (2, 17), but these indirect techniques do not allow beat-to-beat measurement of CSNA and construction of baroreflex curves. Therefore, we also examined the effects of irbesartan treatment on the baroreflex control of CSNA and HR in HF.

Methods

Adult merino ewes (35–49 kg body wt) were housed in individual metabolism cages with other sheep. Experiments were started when sheep were accustomed to laboratory conditions and human contact. Sheep were fed a diet of oaten chaff (800 g/day) and had ad libitum access to water. Experimental procedures were approved by the Animal Experimental Ethics Committee of the Howard Florey Institute in accordance with the Prevention of Cruelty to Animals Act 1986, under the guidelines of the National Health and Medical Research Council of Australia’s Code of Practice for the Care and Use of Animals for Experimental Purposes, which conforms with the Guide for the Care and Use of Laboratory Animals (NIH Publication No. 85-23, revised 1996).

Surgical procedures. Before the studies, sheep underwent two aseptic surgical procedures, each separated by ≥2 wk. Anesthesia was induced with thiopental sodium (15 mg/kg iv) and, after intubation, was maintained with 1.5–2.0% isoflurane-O2. The first surgical procedure involved preparation of a carotid arterial loop in all sheep and, in those to be paced into HF, percutaneous insertion of a pacemaking lead (Medtronic, Minneapolis, MN) via the right jugular vein into the right ventricle. HF was induced by rapid ventricular pacing for 8–10 wk. In a further operation, intrafascicular electrodes were implanted into the left cardiothoracic nerves (34, 35). In all operations, animals were treated with antibiotics (900 mg of procaine penicillin; Ilum Propen, Troy Laboratories, NSW, Australia) at the start of surgery and then for 2 days postoperatively. Postsurgical analgesia was maintained.
with flunixin meglumine (1 mg/kg im; Troy Laboratories) at the start of surgery and 4 and 16 h after surgery. Experiments were conducted on standing, conscious sheep; to minimize any effect of surgical stress, experiments were not started until 4 days after implantation of the electrodes. On the day before implantation of recording electrodes, cannulas were inserted into the carotid artery and jugular vein for measurement of arterial pressure and infusion, respectively (34, 35).

**Experimental protocols.** The development of HF was assessed by measurement of ejection fraction using short-axis M-wave echocardiography on conscious sheep lying on their right side. After placement of ventricular pacing leads, a basal measurement was made before the start of ventricular pacing at 200–220 beats/min. Echocardiography was then performed weekly with the pacing switched off immediately before the measurement. Sheep were considered to be in HF when ejection fraction had fallen to <40%.

CSNA was recorded differentially between the pair of electrodes with the best signal-to-noise ratio. The signal was amplified ($\times$100,000) and filtered (band pass 300–1,000 Hz), displayed on an oscilloscope, and passed through an audio amplifier and loudspeaker. CSNA (5,000 Hz) and arterial blood pressure (100 Hz) were recorded on computer using a CED micro 1401 interface and Spike 2 software (Cambridge Electronic Design, Cambridge, UK).

Recordings of CSNA were started 4 days after surgical implantation of cardiac sympathetic nerve electrodes, and experiments were conducted between 10:00 and 15:00. The pacing was switched off 30 min before the start of experiments and restarted at the end of each day’s experiment. A 5-min recording of resting CSNA and arterial pressure was made in conscious sheep from the normal and HF groups. Then baroreflex curves were generated by measurement of the CSNA and HR responses to increases and decreases in arterial pressure induced by intravenous administration of phenylephrine hydrochloride (33, 67, 133, and 330 mg/min; NeoSynephrine, Abbot Australasia, Kurnell, NSW, Australia) and sodium nitroprusside (42, 83, 167, and 417 mg/min; David Bull Laboratories, Mulgrave, Vic, Australia) for 1–2 min at each dose, with treatments given in random order (27, 34). After construction of baroreflex curves, irbesartan (Bristol-Myers Squibb, Australia) was infused (12 mg·kg$^{-1}$·h$^{-1}$ iv). After 90 min of infusion, the baroreflex was retested during the irbesartan infusion. The dose of irbesartan infused (12 mg·kg$^{-1}$·h$^{-1}$ for 90 min) was demonstrated to abolish the pressor response ($+19 \pm 6$ mmHg) to an intravenous bolus of ANG II (0.2 µg, n = 4).

The ability of systemically infused irbesartan to cross the blood–brain barrier and block AT$_1$ receptors within the blood–brain barrier was assessed in a separate group of animals. In five normal sheep, guide tubes were implanted above the lateral ventricles, as previously described (35). The mean arterial pressure and HR responses to intracerebroventricular infusion of ANG II (10 nmol·ml$^{-1}$·h$^{-1}$ for 30 min) were recorded before and after infusion of irbesartan (12 mg·kg$^{-1}$·h$^{-1}$ iv for 90 min).

**Data analysis.** All data were analyzed on a beat-to-beat basis using custom-written routines in the Spike 2 program (Cambridge Electronic Design) (34, 35). For each heartbeat, the program determined diastolic blood pressure, systolic blood pressure, mean arterial blood pressure, heart period, and the number of discriminated spikes above threshold between the following diastolic pressures as a measure of burst size. The background noise was taken as the spikes per second during the highest dose of phenylephrine, when CSNA was abolished, and this value was subtracted from the data collected on that day. We demonstrated previously that counting discriminated spikes per burst as a measure of sympathetic activity gives results similar to integration and rectification of the sympathetic signal (28). Burst frequency was calculated as the percentage of heartbeats that included spikes of activity above background. The accuracy of burst determination was checked by eye over the 5-min control data for each sheep.

Baroreceptor relations were constructed from data collected during infusion of phenylephrine and nitroprusside (34). CSNA was plotted against diastolic blood pressure, because it has a closer correlation to sympathetic activity than either systolic or mean arterial pressure (29). A four-parameter sigmoidal logistic equation was used to fit the data (SigmPlot version 8.0, SPSS). Variables from the equations of all the graphs were grouped for the normal and HF animals, and average baroreflex relations were plotted.

Values are means ± SE. Data were analyzed using ANOVA (SigmaStat version 2.03, Access Softek). If significant, post hoc hypotheses were tested for significance, as planned beforehand, with the Dunn-Sidak method used for protection against multiple comparisons. P < 0.05 was considered statistically significant.

**RESULTS**

**Comparison of normal and HF groups.** Left ventricular ejection fraction and fractional shortening, measured in conscious sheep by echocardiography, gradually decreased over the 8–10 wk of rapid ventricular pacing at 200–220 beats/min. In the HF animals, at 1–2 days before implantation of recording electrodes, ejection fraction (36±3%) and fractional shortening (15±2%) were significantly reduced compared with the prepaced values (82±3% and 52±2%, respectively, both P < 0.001). Pacing-induced left ventricular dilatation was demonstrated by the increases in end-systolic diameter (from 1.6 ± 0.2 to 3.5 ± 0.3 cm, P < 0.01) and end-diastolic diameter (from 3.2 ± 0.3 to 4.1 ± 0.4 cm, P < 0.05).

The resting level of CSNA was significantly higher in the HF group than in the normal group when assessed as burst incidence (87 ± 3 vs. 44 ± 3 bursts/100 heartbeats) or as total nerve activity (22 ± 4 vs. 10 ± 3 spikes/s, n = 7 per group; Figs. 1 and 2). There were no significant differences in the resting levels of systolic pressure, diastolic pressure, or HR between the normal and HF groups (Fig. 2). In HF, the baroreflex control of HR was impaired, with reductions in the maximum gain and top plateau compared with normal animals, but the baroreflex control of CSNA was not significantly altered (Fig. 3, Table 1).

**Effects of intravenous irbesartan in the normal group.** Irbesartan infusion significantly decreased mean arterial pressure (MAP) and heart period (HR). The responses of CSNA and cardiac nerve (CN) spikes in conscious animals to increases in arterial pressure are shown in Fig. 1.
Effects of intravenous irbesartan in the HF group. In the HF group, irbesartan significantly decreased mean arterial pressure (from $78 \pm 4$ to $73 \pm 2$ mmHg, $P < 0.05$) and diastolic pressure (from $71 \pm 3$ to $66 \pm 2$ mmHg, $P < 0.05$), but the fall in systolic pressure was not significant (from $93 \pm 4$ to $90 \pm 2$ mmHg). These changes were not accompanied by a reflex increase in HR or CSNA (Fig. 2). As in the normal group, in the HF group, irbesartan caused a leftward shift in the CSNA and HR baroreflex curves, indicated by the reductions in the BP_50 for each curve (Table 1, Fig. 3). In contrast to the normal animals, in the HF group, irbesartan decreased the maximum gain of the CSNA baroreflex curve (Table 1, Fig. 3).
Response to central administration of ANG II during intravenous infusion of irbesartan. During the control period, intracerebroventricular infusion of ANG II (10 nmol·ml⁻¹·h⁻¹) increased mean arterial pressure by 15 ± 1 mmHg. After intravenous infusion of irbesartan (12 mg·kg⁻¹·h⁻¹ for 90 min), the pressor response to intracerebroventricular ANG II was slightly, but significantly, reduced (12 ± 1 mmHg, P < 0.05).

DISCUSSION

This is the first study to examine the effects of systemic administration of an ARB on directly recorded CSNA in conscious animals in the normal healthy state and in HF. The major finding was that, in the normal state and in HF, short-term intravenous infusion of irbesartan caused a leftward shift in the CSNA and HR arterial baroreflex curves, so that the decreases in diastolic and mean arterial pressures caused by treatment did not induce reflex increases in CSNA or HR. In HF, irbesartan also increased the maximum gain of the CSNA baroreflex curve but did not change the baroreflex control of HR. In addition, the results confirm our previous reports of a large increase in CSNA in HF, which occurred in the presence of normal arterial baroreflex control of CSNA but desensitized baroreflex control of HR (27, 34).

Effects of irbesartan on resting CSNA. The finding that irbesartan decreased mean arterial pressure without inducing a reflex increase in CSNA in HF has important clinical implications in light of the extensive use of ARBs in HF and the well-established detrimental effect of increased sympathetic drive to the heart in HF (19). In the HF group, irbesartan reduced diastolic pressure by 5 ± 1 mmHg, but this baroreceptor unloading did not cause the expected reflex increase in CSNA. Calculated from the individual arterial baroreceptor curves from untreated HF animals, this decrease in diastolic pressure would normally increase CSNA by 22 ± 7% of the maximum level seen when arterial pressure was lowered by sodium nitroprusside (Fig. 3). Irbesartan had a similar effect in normal animals, which is consistent with the finding that, in normal subjects, intravenous enalapril reduced mean arterial pressure after 30–40 min but did not change cardiac norepinephrine spillover, although renal norepinephrine spillover increased (15). In two further studies of the acute effects of ACE inhibition, it was found that, in the presence of a decrease in blood pressure, muscle SNA was unchanged in normal subjects but reduced in HF patients (6, 23). In patients with HF, a number of studies used ¹²³I-meta-iodobenzylguanidine scintigraphy as a measure of CSNA to examine the effect of 6–9 mo of treatment with ARBs or ACE inhibitors (16–18, 30, 32). In all cases, these studies demonstrated a reduction in this index of CSNA, but it is important to note that the treatments were associated with improved cardiac function, which would have acted independently to reduce CSNA.

The mechanisms by which systemically administered ARBs and ACE inhibitors reduce reflex increases in CSNA are not well defined. It is unlikely that the reduction in cardiac filling pressures that occurs with RAS inhibition (15) is responsible, since we have shown that, in the normal state, this acts to increase CSNA and, in HF, this reflex is abolished (28). Our finding that the dose of irbesartan had little effect on the pressor response to centrally administered angiotensin indicates that only a minor part of the response to irbesartan is likely to be due to its inhibition of angiotensin AT₁ receptors within the blood-brain barrier. It is possible that the effect of irbesartan depends on inhibition of an action of ANG II on the circumventricular organs, areas outside the blood-brain barrier, including the lamina terminalis and area postrema. Indeed, there is extensive evidence that ANG II acts on the area postrema to reduce the sensitivity of the arterial baroreflex (20, 31). However, in the present study, angiotensin receptor blockade decreased the sensitivity of the baroreflex control of CSNA in HF sheep, although not in normal sheep.

Effects of irbesartan on the arterial baroreflex control of CSNA and HR. The leftward shift in the CSNA and HR baroreflex curves caused by irbesartan in the normal and HF groups largely accounted for the lack of baroreflex-mediated increases in CSNA and HR in response to the decreased mean arterial pressure. Irbesartan also decreased the maximum baroreflex gain of CSNA in sheep with HF, whereas angiotensin receptor blockade in rabbits with HF increased the maximum gain of the baroreflex control of renal SNA (21). This difference may reflect the finding that HF is associated with impaired baroreflex control of renal SNA (21). The reported effects of long-term RAS blockade on the baroreflex control of HR in clinical studies are variable: both improvement and no effect have been reported (5, 14, 33). The enhancement in the baroreflex control of HR in HF patients may be related to inhibition of the direct actions of ANG II or treatment-induced improved cardiac function. In the present study, short-term angiotensin receptor blockade did not improve the sensitivity of the baroreflex control of HR, although in rabbits with HF it did partially restore the gain of the reflex (22). Long-term studies are required to determine whether chronic ARB treatment decreases resting levels of CSNA, which may result from an improvement in cardiac function and/or inhibition of central AT₁ receptors due to increased amounts of drug crossing the blood-brain barrier with prolonged treatment.

Perspectives and Significance

It is well established that SNA is increased in HF, with the degree and timing of activation varying between different organs, indicating that multiple factors act at different times to stimulate activity in individual sympathetic nerves. Although the mechanisms are not well understood, previous studies have indicated that the activated RAS in HF plays an important role in determining the increased level of renal SNA. Our studies have focused on the factors leading to the increased CSNA in HF, which is particularly harmful, inasmuch as it predisposes to arrhythmias and sudden death. The present study indicates that, in HF, inhibition of angiotensin AT₁ receptors with irbesartan reduced mean arterial pressure and shifted the CSNA baroreflex curve to the left, leading to a lack of a reflex-induced increase in CSNA. This finding may be clinically important because of the widespread use of ARBs in HF and the well-documented detrimental actions of cardiac sympathoexcitation in HF. Further experiments are required to establish the site at which irbesartan acts to modulate the baroreflex control of CSNA in HF.

ACKNOWLEDGEMENTS

The authors acknowledge the expert technical assistance of Alan McDonald and Tony Dornom.
EFFECT OF AT1 RECEPTOR BLOCKADE ON CSNA IN HF

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GRANTS
This work was supported by National Health and Medical Research Council of Australia Grants 232313 and 509204 and National Heart, Lung, and Blood Institute Grant 5-R01 HL-074932. R. Ramcharandra was the recipient of a National Heart Foundation Postdoctoral Fellowship 07M 3293, and C. N. May was supported by National Health and Medical Research Council Research Fellowships 350328 and 566819.

DISCLOSURES
No conflicts of interest are declared by the author(s).

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