Inhibition of rat gut reperfusion injury with an agent developed for the mouse. Evidence that amplification of injury by innate immunity is conserved between two animal species

Jalil Afnan, Cyrus Ahmadi-Yazdi, Eric G. Sheu, Sean M. Oakes, and Francis D. Moore, Jr.

Department of Surgery, Brigham and Women’s Hospital, Harvard Medical School, Boston, Massachusetts

Submitted 13 July 2009; accepted in final form 14 March 2010

Afnan J, Ahmadi-Yazdi C, Sheu EG, Oakes SM, Moore FD Jr. Inhibition of rat gut reperfusion injury with an agent developed for the mouse. Evidence that amplification of injury by innate immunity is conserved between two animal species. Am J Physiol Regul Integr Comp Physiol 298: R1675–R1681, 2010. First published March 24, 2010; doi:10.1152/ajpregu.00380.2009.—Murine reperfusion injury follows binding of specific IgM natural antibodies to neo-antigens exposed in ischemic tissue. Peptides that mimic the site of antibody binding in the injury prevent IgM binding when administered intravenously before reperfusion. To determine whether this pathogenic sequence is restricted to mice, we have tested the ability of the peptide to prevent reperfusion injury in a dissimilar species, the rat. Sprague-Dawley rats were subjected to 40 min of mesenteric ischemia followed by 180 min of reperfusion. The peptide mimic was administered intravenously prior to reperfusion. Gut injury was quantified using a scoring system based on the hematoxylin-and-eosin section.

Inhibition of rat gut reperfusion injury with an agent developed for the mouse. Evidence that amplification of injury by innate immunity is conserved between two animal species. Am J Physiol Regul Integr Comp Physiol 298: R1675–R1681, 2010. First published March 24, 2010; doi:10.1152/ajpregu.00380.2009.—Murine reperfusion injury follows binding of specific IgM natural antibodies to neo-antigens exposed in ischemic tissue. Peptides that mimic the site of antibody binding in the injury prevent IgM binding when administered intravenously before reperfusion. To determine whether this pathogenic sequence is restricted to mice, we have tested the ability of the peptide to prevent reperfusion injury in a dissimilar species, the rat. Sprague-Dawley rats were subjected to 40 min of mesenteric ischemia followed by 180 min of reperfusion. The peptide mimic was administered intravenously prior to reperfusion. Gut injury was quantified using a scoring system based on the hematoxylin-and-eosin section. 

End-organ damage subsequent to an ischemic insult becomes evident after reperfusion and is the sum of permanent anoxic damage and the inflammatory response to damaged tissue (5). The syndrome of gastrointestinal ischemia and reperfusion injury (I/R), in particular, is associated with high mortality (70–90%) (10) and produces inflammation so severe that it becomes systemic with distinct pulmonary secondary injury. Serum complement activation is a significant component of the injury process, as treatment of rats prior to or at reperfusion with a complement antagonist resulted in attenuation of both local and remote injury (8), identical to effects seen in myocardial infarction models (16). This observation was extended to mice, a species in which complement pathways can be dissected using knockouts. C3 knockout mice showed reduced injury as did C4 knockouts (17), MBL knockouts (7, 19), and antibody-knockouts (RAG-/-) (17). Reconstitution of antibody knockouts using murine antibody fractions showed that only IgM from normal animals restored injury, presumably “natural” IgM (15, 18). B1 peritoneal lymphocytes are a known source of these IgM antibodies, and accordingly, mice with deficiencies of these cells but normal IgM levels were also protected from reperfusion injury (6, 12). Cloning of these cells led to identification of a very limited number of antibody specificities that initiated injury (3, 20), and then to identification of the antigens to which the IgM species bind as a result of injury (4, 21).

A highly conserved region of nonmuscle myosin heavy chain type II A and C is one of the antigens to which the pathogenic natural IgM binds and has led to the creation of 12-mer peptide mimics representing only the conserved region of the molecule. When given to mice intravenously at or just before reperfusion, this peptide mimotope of the natural IgM binding site has been shown to reduce injury in murine models of skeletal (4), cardiac (M. C. Carroll, personal communication), and intestinal I/R (21).

The uncovering of this level of molecular specificity in the response to ischemia and reperfusion suggests that this mechanism of inflammatory triggering may not be confined to the mouse. In addition, the binding site of the pathogenic IgM in the hinge region of nonmuscle myosin heavy chain has a sequence that is highly conserved between microbes and mammals. We hypothesize that the mechanism is exactly conserved in additional species. Our direct testing in rats is presented below and appears to confirm our hypothesis.

MATERIALS AND METHODS

Animals

Male Sprague-Dawley rats (300–350 g) were purchased (Charles River Laboratories, Wilmington, MA). The protocol for this study was approved by the Harvard Medical School Institutional Animal Care and Use Committee. Animals in this study were maintained in accordance with the guidelines of the Committee on Animals of Harvard Medical School and those prepared by the Committee on the Care and Use of Laboratory Animals of the Institute of Laboratory Animal Resources, National Research Council [Department of Health, Education and Human Services, publication no. 85-23 (National Institutes of Health), revised 1985].

Intestinal I/R

The animals were anesthetized with pentobarbital sodium (50 mg/kg ip), and body temperature was maintained with warming pads and the closure of the abdomen between manipulations. A midline laparotomy was performed, and a microvascular clip (Roboz, Gaithersburg, MD) was applied to the origin of the superior mesenteric artery, taking care to exclude the superior mesenteric vein. Thirty-five minutes after occlusion, animals received either peptide (300 μg/300
μl saline/animal) or normal saline (300 μl/animal) via tail vein injection. This dose was a weight-based multiple of the effective dose in mice. At 40 min, the microvascular clip was removed, and reperfusion was confirmed by the presence of pulsations observed in the mesenteric vessels and a return of color to the bowel from its ischemic pallor. All animals received 1.0 ml intravenous normal saline fluid resuscitation. Three hours after clip removal, the animals were euthanized with an overdose of pentobarbital sodium (90 mg/kg), and blood was obtained by cardiac puncture. Ischemic segments of distal small bowel were harvested from all animals and irrigated with intraluminal normal saline solution. Sham animals underwent an identical procedure without microvascular clip application.

Peptides

\( N2 \) 12-amino acid synthetic peptide (New England Peptide, Gardner, MA) with the amino acid sequence of the hinge region of nonmuscle myosin heavy chain II (LMKNMDPLNDNV).

\( SKMM \) 12-amino acid synthetic peptide (New England Peptide, Gardner, MA) with the amino acid sequence of the corresponding site from skeletal muscle myosin heavy chain (LDKNKDPLNETV).

Permeability Index

Gut. Animals for permeability measurements received 1 μCi of \(^{125}\)I-labeled albumin (ICN, Irvine, Ca) in 0.3 ml of normal saline by tail vein injection 10 min before reperfusion. The permeability index (PI) was calculated from counting and weighing intestine and blood: (cpm/g of jejunal loop)/(cpm/g of blood). The intestinal segments were dried in a convection oven (Precision Scientific Group, Chicago, IL) at 90°C for 72 h and then reweighed to correct for blood cpm in the intestine.

Lung. The lungs were lavaged at death with 10 ml of normal saline for a total bronchoalveolar lavage (BAL) return of 6 ml. This was weighed and counted. The PI was calculated as (cpm/g of BAL)/(cpm/g of blood).

Western Blot Analysis

Gut samples from each of the three groups were homogenized in equal volumes of 2% SDS buffer with enzyme inhibitors, chilled, and then centrifuged at 16,000 g for 10 min to collect the supernatant as tissue extract. Equal amounts of protein were then reduced and separated on 4–20% graduated polyacrylamide minigels and transferred to nitrocellulose membranes. The membranes were blocked in buffer containing 5% nonfat milk and 0.1% Tween-20 overnight and then incubated in HRP-conjugated goat anti-Rat IgM (Southern Biotech, Birmingham, AL), at a dilution of 1:10,000. Membranes were developed with an enhanced chemiluminescent Western blot analysis kit (Amersham Biosciences, Piscataway, NJ).

Histopathology

Paraffin sections of formaldehyde-fixed intestinal tissue were stained with hematoxylin and eosin and examined by light microscopy for injury. A modified injury scoring system was used on the basis of that of Williams et al. (17). Scores of 0, 1, or 2 were given to villi that were uninjured, partially injured, or severely injured, respectively. Partial injury was defined as any areas of denuded epithelium or partial shortening (<25%) of the villus. Severe injury was defined as more extensive areas of denuded epithelium, extreme shortening (>25%), or evidence of villus sloughing. Scores of 0, 1, or 2 were, therefore, assigned to each villi viewed under a high-power field. Six random high-power fields were viewed per animal, and the average was taken. A score of 0–100 was, therefore, obtained for each animal, ranging from completely normal intact villi (0) to maximally injured villi (100). (Fig. 1).

Immunohistochemistry Analysis

IgM and C3c staining was performed on paraformaldehyde-fixed serial sections of distal jejunum. All slides were baked at 60°C for 1 h before deparaffinization and rehydration. For IgM staining (1:500) HRP-labeled goat anti-rat IgM (Southern Biotech, Birmingham, AL) was used. For C3, sections were stained with (1:100) Rabbit anti-human C3c (Dako, Carpinteria, CA) followed by (1:500) biotinylated.
goat anti-rabbit IgG (Vector Laboratories, Burlingame, CA) then (1:1,000) avidin-HRP (eBioscience, San Diego, CA). The slides were developed with 3,3’-diaminobenzidine (Sigma, St. Louis, MO), according to the standard protocol.

Statistical Analysis

Data are presented as means ± SE. Comparisons between groups were performed by one-way ANOVA, and when significant difference was found, Student’s t-test with the Bonferroni correction for multiple comparisons was applied.

RESULTS

The macroscopic appearance of the bowel provided a striking example of the protective effect of peptide administration (Fig. 2). Similar regions of terminal small bowel were used for macroscopic and microscopic injury evaluation. Gut from animals that did not receive N2 was edematous, with regions of patchy discoloration and evidence of intraluminal hemorrhage. In contrast, administration of intravenous N2 just prior to reperfusion revealed a more normal looking appearance to the bowel, generally more comparable to the sham animal.

On microscopy, using the gut injury scoring system to grade six random fields, we observed that there was significant protection against mucosal injury in the N2 peptide-treated animal group, compared with I/R injured, untreated animals (Fig. 3). Rat gut injury score was 65.2 ± 11.3 (n = 6) in I/R injured animals. For N2-treated animals, the injury score was 36.2 ± 8.4 (n = 6) (P < 0.01 compared with injured, untreated animals). Animals with sham injury had an injury score 2.1 ± 1.1 (n = 6). The animals that received control peptide (SkMM) scored 54.3 ± 10.6 (n = 6) (P < 0.05), a decrease in injury with respect to the saline group, but not as significant as in the N2 treatment group. SkMM was less protective than N2 (P < 0.01).

Immunohistochemistry was used to demonstrate the contribution of complement and the classical pathway to the pathogenesis of rat I/R injury. Representative slides identify IgM and C3c as deposited in the injured villi (Fig. 4). C3c is a stable cleavage product of C3b, covalently bound to target surfaces as a result of complement activation, only. Notably, villi that were extremely injured or sloughed off were so disintegrated that the locus of IgM or C3c staining could not be seen, so areas of lesser injury were selected, instead of the random sections used for the intestinal scoring system above. Both IgM- and C3-dependent staining were seen at the tips of injured villi. The control slide used sham-injured tissue, and IgM staining was only seen within the serum of the normal vasculature. C3c cannot be demonstrated, as this protein is generated only by complement activation. Use of this technique to compare the degree of deposition of C3c and IgM between I/R injured animals vs. N2-treated I/R injured animals proved impossible as injured villi in injured, untreated animals were not visible for quantitation. Instead, whole tissue homogenates were made for Western blot analysis to compare relative amounts of tissue IgM content. This demonstrated a marked increase in intestinal IgM content in I/R injured animals compared with N2-treated I/R injured animals or sham injured animals. (Fig. 5).

Another manifestation of the local effects of I/R injury is increased tissue edema. There was a statistically significant decrease in the gut PI of N2 peptide-treated animals from I/R injured saline-treated animals: N2 treated rat gut PI = 2.240 ± 0.355 (n = 6), compared with PI = 4.211 ± 0.408 (n = 6) for I/R injured saline-treated animals (P < 0.01). Treatment with control peptide SkMM resulted in PI = 3.474 ± 0.387 (n = 6), statistically different from the N2 group (P < 0.05) but not statistically significant when compared with the saline group. Sham-injured rat gut PI = 1.448 ± 0.159 (n = 6). (Fig. 6).

Gut I/R injury typically produces a pronounced secondary lung injury. Bronchoalveolar lavage was performed to assess lung extravasation of radio-labeled albumin, which is a reflection of pulmonary microvascular integrity. This technique showed a statistically significant decrease in pulmonary permeability index (PI) in the I/R injured, N2 peptide-treated
group PI = 1.197 ± 0.194 (n = 6) compared with the I/R injured, untreated rat lung PI = 2.398 ± 0.286 (n = 6), P < 0.05. The control peptide SkMM was found to have a PI = 1.751 ± 0.136 (n = 6), significantly different from N2 (P < 0.05), but not statistically different from the saline group. The PI of sham-injured rats was 0.894 ± 0.11 (n = 6). (Fig. 7).

DISCUSSION

In this model of rat gut I/R injury, we provide evidence for the attenuation of rat I/R injury following administration of N2, a 12-mer peptide that is the highly conserved binding site on the injury neo-antigen expressed during ischemia in mice and to which pathogenic IgM binds. This drug was given after ischemia and just prior to reperfusion, similar to clinically encountered circumstances.

When similar regions of rat ileum were compared following I/R, a dramatic macroscopic difference was noted (Fig. 2). Untreated I/R rat gut appeared edematous and necrotic with areas of intraluminal hemorrhage, an appearance shared only to a small degree by I/R-injured N2-treated intestine and not at all by sham injured intestine. These findings were an indication of 1) the gross effects of I/R injury with increased vascular permeability, vascular disruption, and necrosis and 2) the ability of N2 peptide to prevent this.

For N2 peptide to have this effect in the rat, there must be both pathogenic antibody specificity directed to N2, the same neo-antigen (nonmuscle myosin heavy chain) and the same expression of this antigen occurring after ischemia. We confirmed this gross finding in several ways. First, conventional histological quantification of injury at the level of the intestinal villi showed that significant protection from I/R-induced mucosal injury was conferred by N2 peptide treatment (Fig. 3). Second, we attempted to use immunohistochemistry and demonstrated deposition of both IgM and C3c on the injured villi (Fig. 4), implicating activation of the same antibody and serum complement components that had been seen in previous models of I/R injury (15, 18). The use of C3c in this analysis allows identification of products that can result only from complement activation and not as aggregations of necrotic tissue or blood. But, the most injured villi simply slough and are unavailable to make a comparison of treated to untreated animals. Therefore, Western blots of intestinal tissue homogenates were compared for the presence of IgM. IgM was dramatically increased in I/R injured tissue and, in turn, reduced in N2-treated I/R injured...
tissue, even with the N2 lane containing relatively more tissue homogenate (Fig. 5). This result indicates not only lessened inflammatory activation in the N2-treated animals but also that N2 peptide was acting by its anticipated mechanism: interference with natural IgM binding to ischemic tissue (4, 21).

Third, tissue edema and vascular permeability were used as an independent method to assess the effect of N2 peptide. The gut lumen was flushed with saline to remove any radioalbumin that accumulated as a result of mucosal necrosis or bleeding. Additionally, the superior mesenteric vein was specifically excluded from clamping to remove vasodilatation as a cause of radioalbumin retention, a feature also not identified on H&E. The PI assessment in the gut itself confirmed that animals treated with N2 peptide were relatively protected from injury (Fig. 6). The lungs, which are a remote target for I/R injury, were found to be relatively protected from injury as well (Fig. 7).

Thus, treatment of rats with N2 peptide 5 min prior to reperfusion substantially attenuated the effects of ischemia and reperfusion injury, namely, gross alterations in gut appearance, mucosal villus sloughing, intestinal deposition of IgM, and both gut and pulmonary permeability edema. We do not know whether a higher dose or an altered time of dosing would have reduced injury parameters to nil. We suspect not, as this N2 dose is similar to that seen in mice (4, 21). This indicates to us that the sum total of measurable ischemia and reperfusion injury is both the intrinsic ischemic injury to the cell and the subsequent invoked inflammatory reaction, which N2 appears to largely prevent. A homologous peptide derived from the hinge region of skeletal muscle myosin and very similar to N2 was used to test the specificity of the rat pathogenic antibody. The minor activity of SkMM on one of the three quantitative measures of injury confirms the specificity of rat pathogenic IgM for N2.

N2 peptide is a 12-amino acid peptide that contains the exact sequence of the hinge region between the major globular subunits of nonmuscle myosin heavy chain IIC, a protein that appears to be the key ischemic injury antigen (4, 21). Mice have a preformed, naturally circulating IgM that recognizes this antigen when it is exposed in the course of ischemic or thermal (14) injury and produces complement and mast cell (1) activation with eventual excessive necrosis of involved tissue. The biology of expression of nonmuscle myosin heavy chain in

Fig. 5. Western blot of SDS-PAGE (reduced) of whole intestinal homogenates, stained for peroxidase activity after reaction with HRP-conjugated goat anti-rat IgM. The 90-kDa band of the IgM heavy chain is prominent in the homogenate from I/R injured animals, which is reduced in N2-treated I/R injured animals, and absent in sham-treated animals. The 25-kDa band corresponding to the IgM light chain is poorly visualized with this antibody. This antibody also recognizes murine albumin (65 kDa), showing that there is more material loaded in the N2 treatment lane than the other lanes.

Fig. 6. Intestinal permeability as determined by radio-labeled albumin extravasation. Intestinal permeability index in N2-treated, I/R injured animals is significantly less than in injured, untreated animals (*P < 0.05). There was no significant decrease seen with the control peptide, SkMM. Intestine from N2-treated animals still demonstrated more edema than that of sham-injured animals.

Fig. 7. Pulmonary permeability index as determined by extravasation of radio-labeled albumin into bronchial-alveolar lavage. Pulmonary permeability index in N2-treated, I/R injured animals is significantly less than in injured, untreated animals (*P < 0.05). There was no significant decrease seen with the control peptide, SkMM. Permeability index in N2-treated animals was greater than in sham-injured animals.
response to injury is not understood at present. It is proposed that N2 peptide is taken up within the binding site of pathogenic natural IgM for nonmuscle myosin, thereby competitively depleting the amount of available pathogenic natural antibody to drive a reperfusion injury. Given the specific antibodies and antigens involved, it would not be surprising to discover that the same principle of an injury neo-antigen and corresponding pathogenic antibody might exist in another species, particularly given the absolute conservation of the primary structure of the hinge region of nonmuscle myosin heavy chain from yeast to humans. What is unexpected, though, is that the specificity of the binding site in pathogenic IgM has not changed, as antibodies are viewed as highly changeable since minor differences in primary structure have a profound effect on binding affinity. For N2 to have any effect in the rat, the rat has to have the identical injury antigen, the identical pathogenic IgM, and the same mechanism of amplifying a reperfusion injury. To argue that N2 peptide is active through a different mechanism ignores the three following features. First, a small peptide is instantaneously degraded in serum, unless complexed within a protective protein pocket, such as within the IgM that recognizes N2. Second, the spectrum of activity in rats is identical to what we have seen in mice. Third, we have provided evidence of reduced IgM binding after N2 administration. Our assessments of the concentration of pathogenic IgM in mice suggest that it is 1–10% of the circulating IgM. One final point concerns the structure of IgM, which is a pentamer with 10 antigen binding sites. For it to activate complement, the shape of the protein must be altered by the binding of the majority of its sites to the target. Thus, the ability of the N2 peptide to prevent complement activation by this type of antibody may be augmented, as only a few of the 10 binding sites would have to be occupied by N2 to disrupt the complement-activating conformation.

It may not seem significant that a drug developed for mouse innate immunity interferes with the rat system. While earlier phylogenetic studies placed the mouse and rat in the same phylum, rodentia, their similarity was largely attributed to shared anatomical features (13). Their genetic similarity or disparity has been more recently elucidated in conjunction with shared anatomical features. First, a small peptide is instantaneously degraded in serum, unless complexed within a protective protein pocket, such as within the IgM that recognizes N2. Second, the spectrum of activity in rats is identical to what we have seen in mice. Third, we have provided evidence of reduced IgM binding after N2 administration. Our assessments of the concentration of pathogenic IgM in mice suggest that it is 1–10% of the circulating IgM. One final point concerns the structure of IgM, which is a pentamer with 10 antigen binding sites. For it to activate complement, the shape of the protein must be altered by the binding of the majority of its sites to the target. Thus, the ability of the N2 peptide to prevent complement activation by this type of antibody may be augmented, as only a few of the 10 binding sites would have to be occupied by N2 to disrupt the complement-activating conformation.

It may not seem significant that a drug developed for mouse innate immunity interferes with the rat system. While earlier phylogenetic studies placed the mouse and rat in the same phylum, rodentia, their similarity was largely attributed to shared anatomical features (13). Their genetic similarity or disparity has been more recently elucidated in conjunction with shared anatomical features. First, a small peptide is instantaneously degraded in serum, unless complexed within a protective protein pocket, such as within the IgM that recognizes N2. Second, the spectrum of activity in rats is identical to what we have seen in mice. Third, we have provided evidence of reduced IgM binding after N2 administration. Our assessments of the concentration of pathogenic IgM in mice suggest that it is 1–10% of the circulating IgM. One final point concerns the structure of IgM, which is a pentamer with 10 antigen binding sites. For it to activate complement, the shape of the protein must be altered by the binding of the majority of its sites to the target. Thus, the ability of the N2 peptide to prevent complement activation by this type of antibody may be augmented, as only a few of the 10 binding sites would have to be occupied by N2 to disrupt the complement-activating conformation.

It may not seem significant that a drug developed for mouse innate immunity interferes with the rat system. While earlier phylogenetic studies placed the mouse and rat in the same phylum, rodentia, their similarity was largely attributed to shared anatomical features (13). Their genetic similarity or disparity has been more recently elucidated in conjunction with shared anatomical features. First, a small peptide is instantaneously degraded in serum, unless complexed within a protective protein pocket, such as within the IgM that recognizes N2. Second, the spectrum of activity in rats is identical to what we have seen in mice. Third, we have provided evidence of reduced IgM binding after N2 administration. Our assessments of the concentration of pathogenic IgM in mice suggest that it is 1–10% of the circulating IgM. One final point concerns the structure of IgM, which is a pentamer with 10 antigen binding sites. For it to activate complement, the shape of the protein must be altered by the binding of the majority of its sites to the target. Thus, the ability of the N2 peptide to prevent complement activation by this type of antibody may be augmented, as only a few of the 10 binding sites would have to be occupied by N2 to disrupt the complement-activating conformation.


