GABA in the paraventricular nucleus tonically suppresses baroreflex function: alterations during pregnancy

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Page MC, Cassaglia PA, Brooks VL. GABA in the paraventricular nucleus tonically suppresses baroreflex function: alterations during pregnancy. Am J Physiol Regul Integr Comp Physiol 300: R1452–R1458, 2011. First published March 23, 2011; doi:10.1152/ajpregu.00736.2010.—It is well established that GABAAergic inputs to the paraventricular nucleus of the hypothalamus (PVN) tonically suppress heart rate and the activity of several sympathetic nerves. However, whether GABA in the PVN enhances the baroreflex in anesthetized female virgin rats. In addition, because GABAAergic inhibition of PVN preautonomic neurons is decreased during pregnancy, it was also determined whether the effects of PVN bicuculline administration on baroreflex function were less in pregnant animals. In virgin rats, PVN microinjection of bicuculline increased (P < 0.05) baroreflex gain and maximum levels of heart rate (gain, from 1.6 ± 0.6 to 3.8 ± 1.3 bpm/mmHg; maximum, from 406 ± 18 to 475 ± 14 bpm) and of lumbar sympathetic nerve activity (gain from 2.6 ± 0.7 to 4.8 ± 1.6%/mmHg; maximum, 149 ± 32 to 273 ± 48%), indicating that PVN GABA normally suppresses baroreflex function. Pregnancy decreased heart rate baroreflex gain (pregnant, 0.9 ± 0.3 bpm/mmHg; virgin, 1.9 ± 0.2 bpm/mmHg; P < 0.05). Following PVN bicuculline administration in pregnant rats, smaller (P < 0.01) increments in baroreflex gain (pregnant, 0.6 ± 0.1 bpm/mmHg; virgin, 2.4 ± 0.9 bpm/mmHg) and maximum (pregnant, 33 ± 7 bpm; virgin, 75 ± 12 bpm; P < 0.05) were produced. Collectively, these data suggest that the PVN normally inhibits the baroreflex via tonic GABAAergic inputs and that this inhibition is less during pregnancy.

Of the PVN neurotransmitters and neuromodulators that effect these many functions, the tonically active inhibitory GABAAergic inputs to preautonomic neurons are dominant (10, 28). Indeed, local pharmacological blockade of PVN GABA receptors elicits profound increases in arterial pressure, heart rate (HR), and sympathetic activity and excites neurons that project to the brain stem or spinal cord (15, 18, 20, 22). Therefore, one aim of the present study was to test the hypothesis that this substantial GABAAergic input also tonically inhibits baroreflex control of HR and sympathetic activity.

A secondary goal was to test whether the GABAAergic inhibition can be altered, specifically during pregnancy. Pregnancy impairs baroreflex function by reducing baroreflex gain (BRG) and the maximum levels of sympathetic activity and HR achieved during acute severe hypotension (3). Yet, indirect evidence suggests that any PVN GABAergic suppressive influence on baroreflex function is lessened, thus counteracting the more primary inhibitory effect of pregnancy. First, pregnancy activates the renin-angiotensin system (3, 30), which has been shown to decrease PVN GABAAergic inhibition of sympathetic activity and heart rate (17, 19, 20). Second, a recent study by Kvochina et al. (16) demonstrated that the increases in renal sympathetic nerve activity, HR, and arterial pressure following blockade of PVN GABA receptors were less in anesthetized rats studied near term compared with nonpregnant animals. Therefore, the second aim of the present study was to test the hypothesis that GABAergic suppression of baroreflex function is reduced during pregnancy, by determining whether the increase in BRG following PVN bicuculline is blunted in pregnant rats.

METHODS

Animals. Experiments were performed using female virgin (240–280 g) or pregnant (390–465 g) Sprague-Dawley rats (Charles River, Wilmington, MA). After arrival, all rats were housed in a room with a 12:12-h light-dark cycle and had free access to food (LabDiet 5001, Richmond, IN) and water; at least 5 days were allowed before any experimentation. Some rats were placed in a male rat’s home cage, and vaginal cytology was examined daily. The presence of sperm was designated pregnancy day 0 (term is 21–22 days), and the female rat was returned to its home cage. All procedures were conducted in accordance with the National Institutes of Health’s Guide for the Health and Use of Laboratory Animals and were approved by the Institutional (Oregon Health & Science University) Animal Care and Use Committee.

Surgery. Throughout the surgery and experiment, body temperature was maintained at 37 ± 1°C using a rectal thermistor, heat lamp, and heating pad. Anesthesia was induced with 5% isoflurane in 100% oxygen. A tracheal tube was first inserted, so that the animals could be artificially ventilated, and a surgical plane of anesthesia was maintained with 2% isoflurane in 100% oxygen. Femoral arterial and venous catheters were implanted for arterial pressure measurements and infusions, respectively. In some rats, after
a midline abdominal incision, a bipolar stainless-steel electrode was placed around a lumbar nerve and secured using lightweight silicone material (Kwik-Sil, World Precision Instruments, Sarasota, FL). Rats were then positioned in the stereotaxic device (David Kopf, Tujunga, CA), with the skull flattened between bregma and lambda. A midline incision was made on the top of the skull, and an opening was burred through the bone on the midline caudal to bregma, to prepare for PVN microinjections or microinjections lateral or rostral to PVN. After completion of all surgical manipulations, an intravenous infusion of urethane (\( \sim 1.1 \text{ g/kg in 1 ml saline} \)) was then administered over \( \sim 30 \text{ min} \); beginning 10 min after initiating the urethane infusion, the gas anesthetic was slowly withdrawn, but artificial ventilation with 100% oxygen was maintained throughout the experiment. After completion of surgery and the urethane infusion, the rats were allowed to stabilize for \( \sim 60 \text{ min} \) before experimentation. The depth of anesthesia was periodically assessed by confirming the lack of response to tail or paw pinch. Additional urethane (0.2 g/kg) was occasionally administered intravenously when needed.

**PVN microinjections.** Single-barreled glass micropipettes (20- to 50-\( \mu \text{m} \) tip diameter) containing bicuculline were directed toward the PVN using the following coordinates (using bregma and the dorsal surface of the dura as zero): 1.8 mm caudal, 0.5 mm lateral, and 7.2–7.6 mm ventral. In addition, to document the anatomical specificity of the PVN injections, additional experiments were performed in which bicuculline was microinjected rostral or lateral to PVN. Coordinates lateral to the PVN were 1.8 mm caudal, 2.0 mm lateral, and 7.2 mm ventral, and coordinates rostral to the PVN were between 0.3 and 0.5 mm caudal, 0.5 mm lateral, and 7.2 mm ventral. Microinjections of bicuculline [60–70 nl of 1 mM in artificial cerebrospinal fluid (aCSF) containing (in mM): 128 NaCl, 2.6 KCl, 1.3 CaCl\(_2\), 0.9 MgCl\(_2\), 20 NaHCO\(_3\), and 1.3 Na\(_2\)HPO\(_4\), pH 7.4] or the aCSF vehicle were made unilaterally over \( \sim 3–7 \text{ s} \) using a PicoPump (World Precision Instruments); the successful microinjection of drugs was verified by watching, through a microscope reticule, the movement of the fluid meniscus a distance calibrated to be \( \sim 60 \text{ nl} \). At the conclusion of the experiment, \( \sim 50 \text{ nl} \) of 2.5% Alcian blue in 0.5 M sodium acetate was injected into the PVN using the same pipette and coordinates as for injections. The brain was removed and placed in 4% paraformaldehyde in PBS for at least 48 h. The hypothalamus was subsequently cut into 25-\( \mu \text{m} \) sections using a cryostat; sections were mounted on glass microscope slides and counter-stained with neutral red. Correct placement into PVN was indicated by dye centered just dorsally, ventrally, or within the nucleus, \( \sim 1.8 \) to 2.2 mm caudal to bregma (Fig. 1).

**Baroreflex function.** Baroreflex function curves were produced and recorded as previously described (26). Briefly, mean arterial pressure (MAP) was first rapidly decreased to \( \sim 50 \text{ mmHg} \) by intravenous infusion of nitroprusside (1 mg/ml; 20 \( \mu \text{l} /\text{min} \)). Then, MAP was slowly and smoothly raised to \( >175 \text{ mmHg} \) over \( \sim 3–5 \text{ min} \) by both decreasing the rate of nitroprusside infusion and infusing phenylephrine at increasing rates (1 mg/ml; 1–35 \( \mu \text{l} /\text{min} \)). Curves were constructed from data obtained during the MAP uprising from 50 to 175 mmHg. MAP, HR, and lumbar sympathetic nerve activity (LSNA; band pass filtered between 100 and 3,000 Hz) were sampled at 2,000 Hz using a Biopac MP100 data acquisition and analysis system. Raw LSNA was rectified and integrated, and the LSNA, MAP, and HR data were grouped into 1-s bins, from which mean values were obtained. LSNA background noise was taken as the post mortem value and was subtracted from experimental LSNA values. LSNA was normalized to control LSNA before the experimental infusions were initiated (percentage of control). The sigmoidal baroreflex relationships between MAP and HR or LSNA generated in each experiment were fitted and compared using the Boltzmann equation: HR = HR\(_{\text{max}}\) = \((P1 - P2)/\left[1 + \exp (MAP - P3)/P4\right] + P2\). P1 is the maximum HR or LSNA, P2 is the minimum HR or LSNA, P3 is the MAP associated with the HR or LSNA value midway between the maximal and minimal HR or LSNA (BP\(_{50}\); denotes position of the curve on the x-axis), and P4 is the width (or mmHg over which the baroreflex operates), the coefficient used to calculate maximum gain, \( -\left(P1 - P2\right)/\left(P4 \times 4\right) \), which is an index of the slope of the most linear part of the sigmoidal baroreflex curve.

**Experimental protocols.** Baseline data were taken as the average of data collected 30 s prior to each baroreflex function assessment. Two control baroreflex curves were first determined, \( \sim 30 \text{ min} \) apart, to establish experimental stability. In most cases, results were within 10%, and the second curve was used for data analysis. In the first set of nonpregnant rats (\( n = 6 \)) instrumented with recording electrodes for LSNA, bicuculline or aCSF was microinjected into PVN, and beginning \( \sim 5 \text{ min} \) after completing the injection, baroreflex function curves for control of LSNA and HR were regenerated. After at least 1 h, the other solution was microinjected, and a second curve was produced. In a second set of nonpregnant animals, following construction of control HR baroreflex curves, bicuculline was microinjected rostral (\( n = 5 \)) or lateral (\( n = 5 \)) to the PVN, and another curve was generated. Finally, in a third set of virgin (\( n = 12 \)) and pregnant (\( n = 8 \); studied on gestational day 20) rats, HR baroreflex function curves were produced before and \( \sim 5 \text{ min} \) after completing a bicuculline microinjection. In four virgin and two pregnant rats in this set, aCSF was also microinjected into PVN (either before or after bicuculline administration), and baroreflex curves were produced as for the first set.
Data analysis. The effects of PVN microinjection of bicuculline and aCSF in virgin rats on hemodynamics and HR/LSNA sigmoidal baroreflex parameters were determined using one-way ANOVA for repeated measures and the post hoc Newman-Keuls test. A paired t-test was used to assess differences between preinjection and postinjection responses in control virgin rats that received bicuculline lateral or rostral to PVN. Differences between and within the second set of virgin and pregnant rats in MAP, HR, and sigmoidal baroreflex curve parameters were determined using two-way ANOVA for repeated measures and the post hoc Newman-Keuls test; in the subset of virgin rats in which the aCSF vehicle was microinjected, differences were assessed using a paired t-test. \( P < 0.05 \) was considered statistically significant.

RESULTS

Effects of microinjection of bicuculline or aCSF into the PVN on baroreflex control of LSNA and HR in nonpregnant rats. PVN bicuculline increased MAP, HR and LSNA (Table 1). In addition, gain of baroreflex control of HR and LSNA increased, as well as HR and LSNA baroreflex maximums, HR baroreflex minimum, and HR and LSNA baroreflex ranges (Fig. 2, Table 1). Neither HR and LSNA width nor BP50 were altered significantly (Table 1). Baroreflex control of HR and LSNA were also not changed following microinjection of the aCSF vehicle into the PVN (Fig. 2, Table 1).

Effects of microinjection of bicuculline rostral or lateral to PVN. In nonpregnant rats, microinjection of bicuculline in regions outside of PVN did not significantly alter HR baroreflex function (Table 2).

Effects of pregnancy. Pregnancy decreased MAP and increased HR (Fig. 3). In addition, baroreflex function was impaired as reflected by decreases in baroreflex gain (Figs. 3 and 4) and increases in width (Table 3); no other baroreflex parameters were significantly altered.

Comparison of effects of microinjection of bicuculline into PVN between nonpregnant and pregnant rats. As in the first experimental series, microinjection of bicuculline into the PVN in nonpregnant rats increased MAP, HR (Fig. 4) and baroreflex gain (Fig. 3). PVN bicuculline also increased maximum and minimum baroreflex HR and HR range, without affecting width or BP50 (Table 3).

In pregnant rats, microinjection of bicuculline into the PVN increased MAP and baroreflex gain, without significantly increasing HR (Figs. 3 and 4). Nevertheless, the increment in gain in pregnant rats \((0.6 \pm 0.1 \text{ bpm/mmHg})\) was less \((P < 0.01)\) than in virgin rats \((2.4 \pm 0.9 \text{ bpm/mmHg})\). Bicuculline also increased maximum baroreflex HR and decreased width.

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### Table 1. Effect of PVN bicuculline and aCSF on basal values and baroreflex parameters

<table>
<thead>
<tr>
<th>Basal Values</th>
<th>MAP, mmHg</th>
<th>HR, bpm</th>
<th>LSNA, %</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>103 ± 5</td>
<td>367 ± 13</td>
<td>100 ± 0</td>
</tr>
<tr>
<td>Bicuculline</td>
<td>115 ± 5*</td>
<td>430 ± 29*</td>
<td>222 ± 16*</td>
</tr>
<tr>
<td>aCSF</td>
<td>99 ± 4</td>
<td>367 ± 19</td>
<td>89 ± 12</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>HR curves</th>
<th>HR Max, bpm</th>
<th>HR Min, bpm</th>
<th>HR Range, bpm</th>
<th>Width, mmHg</th>
<th>BP50, mmHg</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>406 ± 18</td>
<td>330 ± 19</td>
<td>76 ± 22</td>
<td>13.4 ± 2.2</td>
<td>123 ± 6</td>
</tr>
<tr>
<td>Bicuculline</td>
<td>475 ± 14*</td>
<td>344 ± 19*</td>
<td>132 ± 22*</td>
<td>11.0 ± 1.9</td>
<td>126 ± 3</td>
</tr>
<tr>
<td>aCSF</td>
<td>415 ± 4</td>
<td>326 ± 19</td>
<td>89 ± 19</td>
<td>14.8 ± 1.8</td>
<td>121 ± 4</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>LSNA curves</th>
<th>LSNA Max, %</th>
<th>LSNA Min, %</th>
<th>LSNA Range, %</th>
<th>Width, mmHg</th>
<th>BP50, mmHg</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>149 ± 32</td>
<td>1 ± 14</td>
<td>148 ± 38</td>
<td>12.6 ± 2.6</td>
<td>122 ± 7</td>
</tr>
<tr>
<td>Bicuculline</td>
<td>273 ± 48*</td>
<td>18 ± 17</td>
<td>254 ± 44*</td>
<td>12.7 ± 2.5</td>
<td>110 ± 7</td>
</tr>
<tr>
<td>aCSF</td>
<td>148 ± 35</td>
<td>10 ± 20</td>
<td>138 ± 38</td>
<td>12.4 ± 3.0</td>
<td>118 ± 6</td>
</tr>
</tbody>
</table>

MAP, mean arterial blood pressure; HR, heart rate; LSNA, lumbar sympathetic nerve activity; aCSF, artificial cerebrospinal fluid. Max, baroreflex maximum HR or LSNA; Min, baroreflex minimum HR or LSNA; BP50, see text. *\( P < 0.05 \), control vs. bicuculline; \( n = 6 \) virgin rats.
without significantly altering minimum baroreflex HR, HR range, or BPso (Table 3). However, MAP, baroreflex gain, width, and maximum HR remained reduced compared with virgin rats (Figs. 3 and 4 and Table 3).

In the subset of pregnant and nonpregnant animals tested, as in the first experiment, microinjection of the aCSF vehicle into PVN did not appear to change basal values or baroreflex function (e.g., baroreflex gain in bpm/mmHg: virgin rats, 1.4 ± 0.2 to 1.3 ± 0.3, P > 0.50, n = 4; pregnant rats, 0.9 ± 0.4 to 1.1 ± 0.6, n = 2).

**DISCUSSION**

The purpose of the present study was to test the hypothesis that PVN GABAergic inputs tonically suppress baroreflex function and to determine whether this suppression is dimin-

**Table 2. Effect of microinjections of bicuculline lateral or rostral to PVN on mean arterial pressure, heart rate and baroreflex parameters**

<table>
<thead>
<tr>
<th></th>
<th>MAP, mmHg</th>
<th>HR, bpm</th>
<th>Gain, bpm/mmHg</th>
<th>Max, bpm</th>
<th>Min, bpm</th>
<th>Width, mmHg</th>
<th>BPso, mmHg</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control Lateral</td>
<td>101 ± 3</td>
<td>364 ± 24</td>
<td>1.6 ± 0.2</td>
<td>366 ± 21</td>
<td>308 ± 31</td>
<td>8.2 ± 1.1</td>
<td>120 ± 3</td>
</tr>
<tr>
<td>Bicuculline Lateral</td>
<td>102 ± 4</td>
<td>371 ± 23</td>
<td>1.6 ± 0.3</td>
<td>389 ± 20</td>
<td>303 ± 33</td>
<td>13.3 ± 1.0</td>
<td>130 ± 3</td>
</tr>
<tr>
<td>Control Rostral</td>
<td>107 ± 4</td>
<td>359 ± 8</td>
<td>1.8 ± 0.4</td>
<td>379 ± 19</td>
<td>309 ± 12</td>
<td>11.0 ± 1.3</td>
<td>123 ± 4</td>
</tr>
<tr>
<td>Bicuculline Rostral</td>
<td>106 ± 4</td>
<td>373 ± 12</td>
<td>1.7 ± 0.6</td>
<td>389 ± 24</td>
<td>323 ± 13</td>
<td>10.8 ± 1.8</td>
<td>122 ± 3</td>
</tr>
</tbody>
</table>

Max, Baroreflex maximum HR; Min, Baroreflex minimum HR.

![Fig. 3. Effect of PVN BIC microinjection on mean arterial pressure (MAP), HR, and BRG. *P < 0.05, BIC compared with CON. †P < 0.05, pregnant compared with nonpregnant.](http://ajpregu.physiology.org/)

**Fig. 3.** Effect of PVN BIC microinjection on mean arterial pressure (MAP), HR, and BRG. *P < 0.05, BIC compared with CON. †P < 0.05, pregnant compared with nonpregnant.
ished during pregnancy. The major new findings are 1) blockade of PVN GABA<sub>A</sub> receptors via PVN microinjection of bicuculline enhances baroreflex control of HR and LSNA by increasing BRG and the reflex-induced maximal levels of HR and LSNA, and 2) the effects of bicuculline to improve baroreflex control of HR are smaller in late-pregnant rats. From these data, we conclude that the PVN normally inhibits the baroreflex via tonic GABAergic inputs and that this inhibition is less during pregnancy. Moreover, it would appear that the mechanisms by which pregnancy impairs baroreflex function do not involve greater GABAergic inhibition in the PVN.

While PVN GABA has been clearly established to tonically inhibit the basal activity of multiple sympathetic nerves (15, 27), a major goal of the present series of experiments was to test whether it also suppresses baroreflex function. Previous studies have suggested that the PVN may be involved in the regulation of the arterial baroreceptor reflex. Electrophysiological studies demonstrated that PVN neurons that project to the spinal cord are sensitive to changes in arterial pressure (23). Moreover, using c-Fos immunocytochemistry as an indirect index of neuronal activation, several previous reports suggest that PVN preautonomic neurons, albeit a small percentage, are stimulated by decreases in arterial pressure (2, 8). Patel and Schmid (25) further reported that PVN injection of lidocaine enhances lumbar sympathoinhibition in response to pressor stimuli (25). Conversely, in other studies, electrical stimulation of the PVN inhibited baroreflex responses (6, 11). However, the use of lidocaine or electrical stimulation may also have inhibited or stimulated fibers in passage, rather than influencing PVN neuronal activity directly. Whether the substantial GABAergic restraint of PVN preautonomic neurons also contributes to PVN regulation of the baroreflex has not been previously tested.

In the present study, we found that PVN microinjection of the GABA<sub>A</sub> antagonist, bicuculline, markedly increased gain of baroreflex control of HR and LSNA. In addition, the maximum levels of LSNA and HR achieved during severe hypotension were significantly increased by local blockade of PVN GABAergic influences. Importantly, neither PVN microinjection of the aCSF vehicle nor microinjection of bicuculline outside of the PVN significantly altered the baroreflex, indicating that the preparation was stable and that the effect of bicuculline was site specific. Therefore, we conclude that GABAergic inputs to the PVN tonically suppress baroreflex function and that release of this inhibition can enhance baroreflex gain and maximal levels of LSNA and HR.

Another major goal of this study was to determine whether PVN GABAergic inhibition of the baroreflex can be altered, specifically during pregnancy. We found that local blockade of PVN GABA<sub>A</sub> receptors only slightly enhanced baroreflex function in pregnant animals, and baroreflex function remained depressed in pregnant rats compared with virgin animals receiving PVN bicuculline. These results suggest that pregnancy reduces the inhibition of the baroreflex by endogenous GABA in PVN, which is consistent with a recent report that, during pregnancy in rats, PVN GABAergic inhibition of basal LSNA is also reduced (16). An alternative interpretation is that tonic excitatory drive of PVN preautonomic neurons, which would be revealed following blockade of PVN GABA<sub>A</sub> receptors (4, 5), is lessened. Arguing against this possibility, considerable

Table 3. Comparison of effects of PVN bicuculline on HR baroreflex parameters in pregnant and nonpregnant rats

<table>
<thead>
<tr>
<th></th>
<th>Max, bpm</th>
<th>Min, bpm</th>
<th>Range, bpm</th>
<th>Width, mmHg</th>
<th>BP&lt;sub&gt;50&lt;/sub&gt;, mmHg</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>367 ± 10</td>
<td>293 ± 12</td>
<td>74 ± 8</td>
<td>10.5 ± 1.1</td>
<td>118 ± 3</td>
</tr>
<tr>
<td>Bicuculline</td>
<td>462 ± 14*</td>
<td>321 ± 14*</td>
<td>121 ± 13*</td>
<td>9.5 ± 1.5</td>
<td>125 ± 5</td>
</tr>
<tr>
<td>Pregnant</td>
<td>385 ± 14</td>
<td>323 ± 7</td>
<td>62 ± 15</td>
<td>21.0 ± 3.6†</td>
<td>124 ± 3</td>
</tr>
<tr>
<td>Bicuculline</td>
<td>418 ± 15*†</td>
<td>344 ± 10</td>
<td>75 ± 8</td>
<td>14.3 ± 1.7†*</td>
<td>121 ± 5</td>
</tr>
</tbody>
</table>

Max, baroreflex maximum HR; Min, baroreflex minimum HR. *P < 0.05, control vs. bicuculline; †P < 0.05, pregnant vs. nonpregnant.

Fig. 4. Effect of PVN microinjection of bicuculline (gray symbols and dashed lines) on baroreflex control of HR in pregnant and nonpregnant rats. Black lines and symbols are control curves.
previous work using multiple diverse approaches suggests that sympathetic tone is elevated during pregnancy (3). Moreover, in other pathophysiological states characterized by increased basal sympathetic tone, such as heart failure and hypertension, PVN GABAergic influences are also reduced (21, 22).

The mechanism by which pregnancy reduces the magnitude of PVN GABAergic inhibition was not investigated; however, previous work implicates ANG II. First, ANG II is increased during pregnancy (3, 30). Second, ANG II contributes to the increased basal levels of sympathetic activity in pregnant animals (24), indicating that angiotensin influences neural control of the circulation. Indeed, pregnancy appears to increase the relative contribution of PVN ANG II type 1 receptors to the support of arterial pressure and renal sympathetic nerve activity (16). Third, acute and chronic increases in ANG II decrease GABAergic function in PVN, possibly by presynaptic inhibition of GABA release (17, 19, 20).

In agreement with previous work in conscious and anesthetized animals (3), the current study demonstrated that pregnancy impairs gain of baroreflex control of HR. At least two mechanisms contribute to this. First, increased levels of the neurosteroid metabolite of progesterone, 3α-hydroxy-dihydroprogesterone (3α-OH-DHP), decrease the maximal level of sympathetic activity produced during hypotension by increasing GABAergic suppression of premotor neurons in the rostral ventrolateral medulla (3, 14). The present results demonstrating reduced GABA influences in PVN suggest that the effect of 3α-OH-DHP to enhance GABA actions in the PVN is minimal. Second, pregnancy decreases brain insulin, which contributes to the fall in BRG (1, 9). Insulin normally increases baroreflex gain via an action in the hypothalamus (26) and the PVN is involved (3). Insulin is an inhibitory neurotransmitter (31, 33); therefore, insulin would be expected to excite autonomic neurons by disinhibition. The present results suggest that insulin does not enhance the baroreflex by inhibition of PVN GABAergic influences, since this mechanism would predict increased PVN GABAergic inhibition during pregnancy as brain insulin levels fall. Instead, the results suggest that the mechanisms by which pregnancy impairs the baroreflex do not involve greater PVN GABAergic inhibition.

Perspectives and Significance

The present results demonstrate that GABAergic PVN inputs tonically suppress the baroreflex and that this suppression is less during pregnancy, which may counteract the dominant effect of pregnancy to inhibit baroreflex control of HR and sympathetic activity. In addition to pregnancy, other conditions like heart failure and hypertension may be associated with impaired baroreflex function (12, 13). Interestingly, in these pathophysiological states, decreased PVN GABAergic influences on basal sympathetic tone have also been observed (21, 22). Therefore, we speculate that, in these conditions, like pregnancy, the suppression of baroreflex function does not involve enhanced GABAergic inhibition of PVN preautonomic neurons.

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