

# The therapeutic potential of hydrogen sulfide: separating hype from hope

**Kenneth R. Olson**

*Indiana University School of Medicine-South Bend, South Bend, Indiana*

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**Olson KR.** The therapeutic potential of hydrogen sulfide: separating hype from hope. *Am J Physiol Regul Integr Comp Physiol* 301: R297–R312, 2011. First published May 4, 2011; doi:10.1152/ajpregu.00045.2011.—Hydrogen sulfide (H<sub>2</sub>S) has become the hot new signaling molecule that seemingly affects all organ systems and biological processes in which it has been investigated. It has also been shown to have both proinflammatory and anti-inflammatory actions and proapoptotic and anti-apoptotic effects and has even been reported to induce a hypometabolic state (suspended animation) in a few vertebrates. The exuberance over potential clinical applications of natural and synthetic H<sub>2</sub>S-“donating” compounds is understandable and a number of these function-targeted drugs have been developed and show clinical promise. However, the concentration of H<sub>2</sub>S in tissues and blood, as well as the intrinsic factors that affect these levels, has not been resolved, and it is imperative to address these points to distinguish between the physiological, pharmacological, and toxicological effects of this molecule. This review will provide an overview of H<sub>2</sub>S metabolism, a summary of many of its reported “physiological” actions, and it will discuss the recent development of a number of H<sub>2</sub>S-donating drugs that show clinical potential. It will also examine some of the misconceptions of H<sub>2</sub>S chemistry that have appeared in the literature and attempt to realign the definition of “physiological” H<sub>2</sub>S concentrations upon which much of this exuberance has been established.

hydrogen sulfide-donating drugs; vasoactivity; ischemia reperfusion injury; sulfur cycle; gasotransmitter

THE INITIAL DISCOVERY by Hideo Kimura’s group that hydrogen sulfide (H<sub>2</sub>S)<sup>1</sup> was a biologically relevant signaling molecule (reviewed in Ref. 74) has heightened interest in the physiology and pharmacology of gaseous mediators. Unlike the first gaseous signaling molecule, nitric oxide (NO), whose introduction was met with initial skepticism, H<sub>2</sub>S has more or less been enthusiastically embraced by the scientific community, and there has been considerable effort to expeditiously imbue this obnoxious smelling gas into medical applications. This wave of exuberance has reheightened interest in the dietary sources of H<sub>2</sub>S, and it has spawned the development of a number of H<sub>2</sub>S-“donating” drugs, many of which are in various stages of clinical trials. However, it is becoming increasingly evident that there is still much to be learned about the basic properties of H<sub>2</sub>S measurement, metabolism, and signaling mechanisms. This review will provide an overview of the effects of H<sub>2</sub>S on physiological systems, summarize the new H<sub>2</sub>S-donating drugs that are showing clinical potential, and take a critical look at the some of the remaining uncertainties surrounding H<sub>2</sub>S chemistry and tissue concentrations.

## *Hydrogen Sulfide as a Toxic Gas*

The toxic effects of H<sub>2</sub>S have been known for centuries, and it remains second only to carbon monoxide as the most common cause of gas-related fatalities in the workplace (46, 190). H<sub>2</sub>S has even gained notoriety in a recent spate of 220 suicide cases in less than 3 mo in Japan (107). Less is known of the effects of low-level ambient H<sub>2</sub>S that are often associated with sewage plants, waste lagoons, natural gas/oil wells, and oil refineries, as well as a variety of other industrial applications. Recent studies on residents of Southeastern New Mexico exposed to these environments have shown positive correlations with H<sub>2</sub>S exposure and impaired neurobehavioral functions compared with controls (73). This suggests that even “therapeutic” use of H<sub>2</sub>S is not without potential hazards. Thresholds for the major effects of H<sub>2</sub>S exposure are shown in Table 1.

The inhibitory effects of H<sub>2</sub>S on mitochondrial cytochrome-*c* oxidase have been well characterized and this is generally assumed to be the focus of H<sub>2</sub>S toxicity (34). However, the clinical presentation of poisoning by H<sub>2</sub>S and cyanide, another well-known inhibitor of oxidative phosphorylation that also inhibits cytochrome-*c* oxidase, are so distinct as to suggest different modes of toxicity (46). Another rather unusual feature of H<sub>2</sub>S toxicity is an extremely steep dose-effect response. Early studies in dogs (47) and other mammals (38, 25), and more recent anecdotal information from human cases (46) have shown that H<sub>2</sub>S toxicity is closely correlated with H<sub>2</sub>S concentration and considerably less dependent upon the duration of exposure. This suggests that animals can

<sup>1</sup> Unless otherwise noted, H<sub>2</sub>S refers to the sum of dissolved H<sub>2</sub>S gas and HS<sup>-</sup>, often referred to as “sulfide”. At physiological pH, S<sub>2-</sub> is assumed to be negligible.

Address for reprint requests and other correspondence: K. R. Olson, Indiana Univ. School of Medicine-South Bend, 1234 Notre Dame Ave., South Bend, IN 46617 (e-mail: kolson.1@nd.edu).

Table 1. *The effects of H<sub>2</sub>S exposure*

Ambient H <sub>2</sub> S, ppm	Equivalent Total Plasma Sulfide, μM <sup>a</sup>	Effects
0.01–0.3	0.003–0.1	Threshold for detection
1–3	0.3–1	offensive odor, headaches
10	3.3	8-h occupational exposure limit in Alberta, Canada
15	4.9	15-min exposure limit in Alberta, Canada
20–50	6.5–16.2	eye and lung irritation
100	32.5	olfactory paralysis
250–500	81.1–162.3	pulmonary edema
500	162.3	sudden unconsciousness (“knockdown”), death within 4 to 8 h
1000	324.5	immediate collapse, breathing ceases within several breaths

All except “Equivalent Total Plasma Sulfide” column modified from Guidotti (46). <sup>a</sup>Equivalent plasma sulfide calculated after Whitfield et al. (186, supplemental information), assuming H<sub>2</sub>S equilibrates across the alveolar membranes (169), Henry’s Law constant for H<sub>2</sub>S at 37°C, 140 mM NaCl is 0.0649 M·atm<sup>-1</sup> (27), and 20% of total sulfide exists as H<sub>2</sub>S gas (115).

rapidly metabolize H<sub>2</sub>S up to a critical level and, as a corollary, this efficient metabolic capacity should keep free H<sub>2</sub>S at very low levels. These studies should, but have not often, raised questions regarding “physiological” concentrations of H<sub>2</sub>S in tissues and blood. This point is discussed in detail in a later section.

*Hydrogen Sulfide Biosynthesis and Metabolism*

**Biosynthesis.** Much of the metabolism of sulfides, including H<sub>2</sub>S, passes through cysteine (Cys) metabolism (Fig. 1). Cysteine can be oxidized to cysteinesulfinate (Csa), or it can be desulfurated by reducing reactions that generate either H<sub>2</sub>S or sulfane sulfur (a persulfide; 149). In the oxidative—and generally assumed catabolic—pathway for cysteine, cysteine dioxygenase (CDO) catalyzes the addition of molecular oxygen to cysteine producing Csa, which may be further oxidized to

sulfite or taurine (149). As perhaps a general indication of a broad-spectrum of sulfur-mediated effects on biological systems, both Csa and its metabolites have themselves been shown to affect a variety of physiological processes (68, 100). CDO is found in liver, adipose, intestine, pancreas, and kidney. Because activity of CDO is highly regulated by dietary cysteine, CDO is a regulator, if not the primary one, of cysteine availability in vivo. By oxidizing excess and presumably toxic cysteine, CDO provides a constant and low-level background of cysteine for H<sub>2</sub>S and sulfane sulfur biosynthesis. This may be important in preventing excessive H<sub>2</sub>S production (33).

H<sub>2</sub>S can be produced from cysteine via a variety of biochemical pathways. Early studies indicated that cystathionine β-synthase (CBS) was the predominant enzymatic pathway for H<sub>2</sub>S production in the brain, whereas cystathionine γ-lyase (CSE, also known as CGL) was responsible for H<sub>2</sub>S production in the

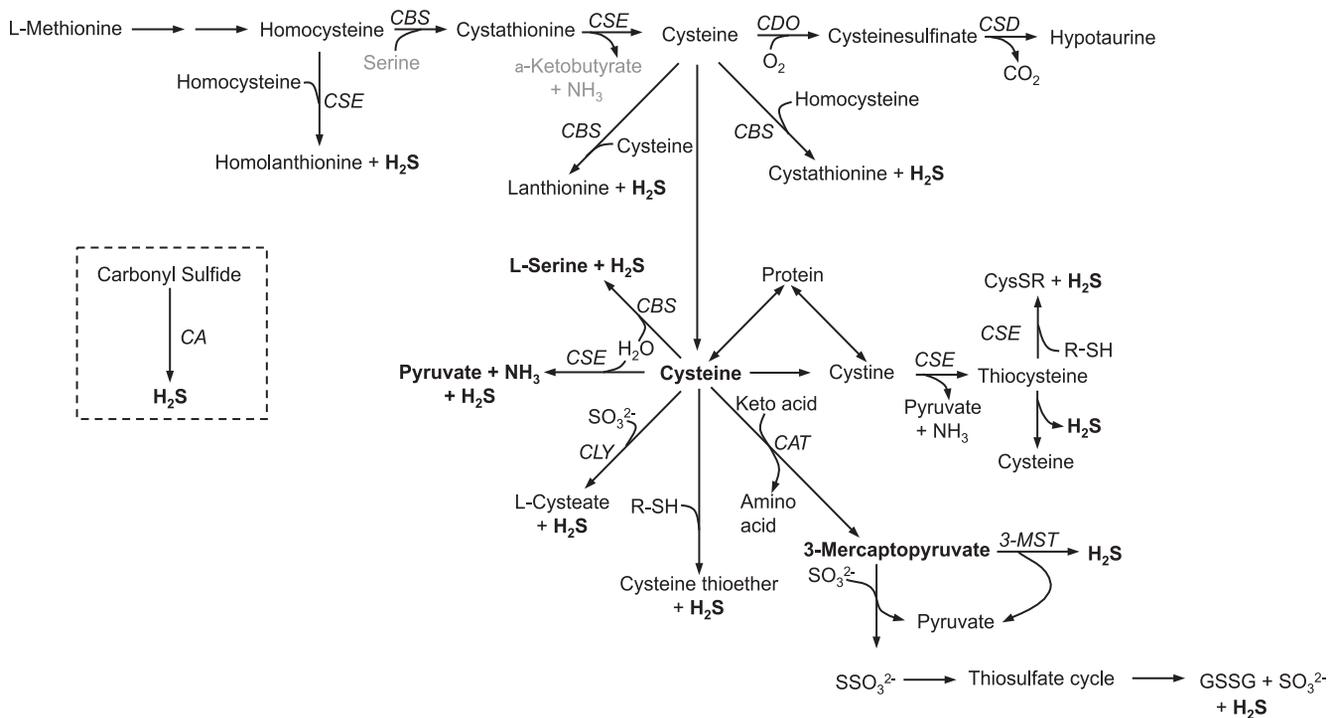


Fig. 1. Potential pathways for H<sub>2</sub>S production and metabolism. Inset shows potential H<sub>2</sub>S production from carbonyl sulfide. CA, carbonic anhydrase; CAT, cysteine aminotransferase; CBS, cystathionine β-synthase; CDO, cysteine dioxygenase; CLY, cysteine lyase; CSD, cysteine sulfinate decarboxylase; CSE, cystathionine γ-lyase; MST, 3-mercaptopyruvate sulfurtransferase; R-SH, thiol. [Modified from Julian et al. (65), Kabil et al. (66), Singh et al. (143), and Stipanuk and Ueki (149).]

vasculature (75). Recent studies have shown that CBS is present in the endothelium and two enzymes acting in tandem, cysteine aminotransferase (CAT) and 3-mercaptopyruvate sulfurtransferase (MST), are also present in vascular endothelium and brain, whereas MST, but not CAT, is found in vascular smooth muscle (75, 119). CAT transfers the amine group from cysteine to an acceptor, such as  $\alpha$ -ketoglutarate, resulting in 3-mercaptopyruvate, which is then desulfurated by MST. In addition to H<sub>2</sub>S, reduced sulfur in the form of sulfane sulfur can also be generated and, in fact, sulfane sulfur appears to be the only product of the CAT-MST pathway (66). Kimura's group found relatively high levels of CAT-MST in the brain, and they proposed that this is a major pathway for H<sub>2</sub>S production in the brain, but they also suggested that the H<sub>2</sub>S is immediately "stored" as sulfane sulfur, the latter serving as a less labile form of H<sub>2</sub>S that may be readily accessible during appropriate physiological conditions (60, 141). However, reducing conditions and an alkaline environment are necessary for cleavage of this RS-S bond to form H<sub>2</sub>S and because these conditions may not be routinely encountered intracellularly, the significance of the CAT/MST pathway in H<sub>2</sub>S synthesis remains questionable. Both CBS and CSE have recently been shown to circulate in human plasma and to generate H<sub>2</sub>S from cysteine or homocysteine plus cysteine (13). This generation of H<sub>2</sub>S has been proposed not only to reduce circulating homocysteine, but also to protect the endothelium from oxidative stress (12).

Both CBS and CSE are cytosolic, pyridoxyl-5'-phosphate-dependent, enzymes. CBS activity appears to be controlled by a number of factors. *S*-adenosylmethionine (AdoMet) is an allosteric activator of CBS and when AdoMet levels are low, CBS activity decreases to direct sulfur flow through the transmethylation pathway, thereby conserving methionine. Elevated AdoMet increases CBS activity to produce cysteine via the transsulfuration pathway (148). CBS contains a heme group that, when it binds with carbon monoxide (CO), inhibits the enzyme. CBS is also inhibited by reducing conditions, but contrary to a number of earlier reports, neither NO nor calmodulin appears to be physiological regulators of CBS activity (8).

Using physiologically relevant substrate concentrations and kinetic simulations, Banerjee's group (cf. 23, 66, 143) concluded that 1) H<sub>2</sub>S generation from cysteine is primarily catalyzed by CSE, 2) H<sub>2</sub>S production by CBS is through condensation of cysteine and homocysteine and depending on the level of AdoMet activation, this may account for 25–70% of the H<sub>2</sub>S generated under resting conditions, 3) H<sub>2</sub>S biosynthesis can occur independent of cysteine; condensation of two molecules of homocysteine, catalyzed by CSE, yields homolanthionine and H<sub>2</sub>S, and may account for as much as 30% of the total H<sub>2</sub>S biosynthesis, 4) CSE activity is substantially increased by elevated homocysteine, whereas CBS activity is unaffected. Condensation of two homocysteine molecules, along with the condensation of homocysteine and cysteine, appear to be important clearance pathways in hyperhomocysteinemia. It has been proposed that during severely elevated homocysteine (200  $\mu$ M), as seen in hyperhomocysteinemia,  $\alpha$ ,  $\gamma$ -elimination and  $\gamma$ -replacement of homocysteine, catalyzed by CSE, may produce excessive amounts of H<sub>2</sub>S and thereby contribute to the cardiovascular pathology associated with this condition (23).

Commonly used inhibitors of CSE include propargyl glycine (PPG) and  $\beta$ -cyanoalanine. Aminoxyacetate (AOA) is commonly used to inhibit CBS and hydroxylamine to inhibit both

enzymes (although a number of studies erroneously claim this is a specific inhibitor of CBS). Unfortunately, none of these inhibitors are specific for sulfur metabolism and H<sub>2</sub>S production; furthermore, they are often poorly absorbed by tissues (153).

#### Other Potential Biosynthetic Pathways

There are numerous other potential metabolic pathways for H<sub>2</sub>S generation that have been described in invertebrates (Fig. 1; Ref. 65), but these have not been systematically evaluated in mammalian tissues. The resurgent interest in H<sub>2</sub>S will undoubtedly lead to reevaluation of these, heretofore, overlooked biosynthetic pathways and identification of novel ones. Indeed, the literature is replete with studies that show that many of the biological effects produced by H<sub>2</sub>S can also be affected by a variety of other sulfur-donating molecules. One potentially novel pathway that needs to be investigated is H<sub>2</sub>S production from carbonyl sulfide (COS; chemical structure: O=C=S). Like H<sub>2</sub>S, COS is a gas that has both natural (volcanoes, hot springs, oils and trees) and man-made (biomass and fossil fuel consumption, wastewater treatment, etc.) origins and it is the most prevalent sulfur gas in the atmosphere (152). COS is the only volatile sulfur that is increased in exhaled air of patients with cystic fibrosis (69) or of lung transplant patients during the acute rejection phase (150). COS is also exhaled by patients with chronic liver disease (135). COS has been demonstrated to be produced by porcine coronary arteries *in vitro*, and the rate of COS production is enhanced by stimulating the vessels with ACh or the calcium ionophore, A23187 (7). In solution, COS slowly decomposes to H<sub>2</sub>S, but this reaction is greatly accelerated by the enzyme carbonic anhydrase. In fact, CO<sub>2</sub> and COS may be the primary substrates of this enzyme (134). Whether or not the biosynthesis of COS is related to H<sub>2</sub>S production and subsequent signaling events remains to be determined.

#### Metabolism (Inactivation)

Oxidation of H<sub>2</sub>S occurs in the mitochondria (53). As shown in Fig. 2, two membrane-bound sulfide:quinone oxidoreductases (SQR) oxidize sulfide to the level of elemental sulfur, simultaneously reducing a cysteine disulfide, and resulting in

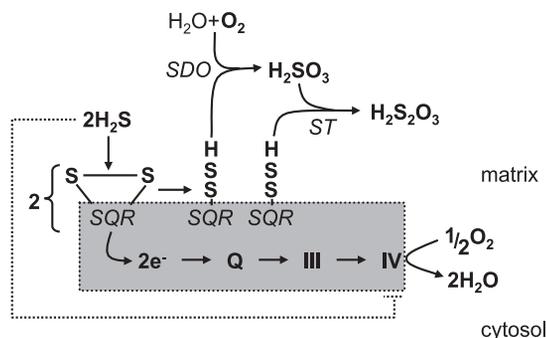


Fig. 2. Mitochondrial oxidation of H<sub>2</sub>S. Two sulfide:quinone oxidoreductase (SQR) in the mitochondrial membrane (stippled box) oxidize sulfide to the level of elemental sulfur, simultaneously reducing a cysteine disulfide, and resulting in the formation of a persulfide group at one of the SQR cysteines (SQR-SSH). Sulfur dioxygenase (SDO) then oxidizes one persulfide to sulfite (H<sub>2</sub>SO<sub>3</sub>), consuming molecular oxygen and water in the process. The second persulfide is transferred from the SQR to sulfite by sulfur transferase (ST) producing thiosulfate (H<sub>2</sub>S<sub>2</sub>O<sub>3</sub>). Electrons from H<sub>2</sub>S are fed into the respiratory chain via the quinone pool (Q), and ultimately transferred to oxygen by cytochrome-c oxidase (complex IV).

formation of persulfide groups at one of the SQR cysteines. Sulfur dioxygenase (SDO) then oxidizes one of the persulfides to sulfite (H<sub>2</sub>SO<sub>3</sub>), consuming molecular oxygen and water in the process. Sulfur from the second persulfide is transferred from the SQR to sulfite by sulfur transferase producing thiosulfate (H<sub>2</sub>S<sub>2</sub>O<sub>3</sub>). Most thiosulfate is further metabolized to sulfate by thiosulfate reductase and sulfite oxidase. Electrons from H<sub>2</sub>S are fed into the respiratory chain via the quinone pool (Q), and finally transferred to oxygen at complex IV. Oxygen consumption is obligatory during H<sub>2</sub>S metabolism, and 1 mol of oxygen is consumed for every mol of H<sub>2</sub>S oxidized along the electron transport chain (53). Oxidation of H<sub>2</sub>S to thiosulfate requires additional oxygen at the level of SDO, resulting in a net utilization of 1.5 mol of oxygen per mol of H<sub>2</sub>S (or 0.75 mol of O<sub>2</sub> per mol H<sub>2</sub>S; Ref. 82). Metabolism of H<sub>2</sub>S through SQR appears ubiquitous in tissues, a notable exception being brain (82). It is important to note that sulfide oxidation in the mitochondria appears to take priority over oxidation of other carbon-based substrates, ensuring its efficient removal (24). This plus the fact that the capacity of cells to oxidize sulfide appears to be considerably greater than the estimated rate of sulfide production (24) ensures that intracellular H<sub>2</sub>S concentrations are very low. Interestingly, the statin, atorvastatin, increases H<sub>2</sub>S production in perivascular adipose tissue by producing coenzyme Q<sub>9</sub> deficiency and thereby inhibiting mitochondrial oxidation (189).

The relationship between H<sub>2</sub>S and O<sub>2</sub> consumption is classical hormesis; at low concentrations, H<sub>2</sub>S stimulates oxygen consumption (and may even result in net ATP production), whereas it is inhibited by elevated H<sub>2</sub>S. This was originally shown in invertebrates and lower vertebrates and more recently demonstrated in the mammalian colon (45). At higher concentrations, H<sub>2</sub>S inhibits the respiratory chain by directly inhibiting cytochrome-*c* oxidase (24). The exact H<sub>2</sub>S concentration at which this occurs is unclear; purified cytochrome-*c* oxidase is inhibited by <1 μM H<sub>2</sub>S, whereas progressively greater (2 or 3 orders of magnitude) higher H<sub>2</sub>S concentrations are needed to inhibit the enzyme in intact mitochondria and then whole cells. Cytochrome-*c* oxidase is half maximally inhibited by ~20 μM H<sub>2</sub>S and may not be fully inhibited until H<sub>2</sub>S concentrations reach 40–50 μM (6, 24). This may reflect diffusion limitation as the enzyme becomes further removed from the exogenously administered H<sub>2</sub>S. It also should provide a cautionary note in interpreting studies that routinely employ 100 μM–1 mM H<sub>2</sub>S to demonstrate a “physiological” effect. The converse, i.e., the effect of O<sub>2</sub> on H<sub>2</sub>S consumption, is discussed in *H<sub>2</sub>S and oxygen sensing*.

### *H<sub>2</sub>S Biology*

Interest in H<sub>2</sub>S biology has spawned nearly as many reviews (at latest count, 32 in 2010 alone) as original articles. Reviews have even appeared where, at the time, the effects of H<sub>2</sub>S on a particular system were unknown (87, 196). The following sections provide a brief overview of H<sub>2</sub>S biology. For further details, the reader is referred to a few of the most recent reviews following each section.

*H<sub>2</sub>S and the nervous system.* Potentiation of the *N*-methyl-D-aspartate (NMDA) receptor with the resultant alteration of long-term potentiation (LTP) in the hippocampus was the first biological effect ascribed to H<sub>2</sub>S (1). Not long thereafter, it was

noted that patients with Down syndrome had elevated concentrations of H<sub>2</sub>S in cerebral spinal fluid. This would be predicted from the fact that chromosome 21 encodes CBS (which may be the major H<sub>2</sub>S-producing enzyme in the brain) and is overexpressed in these patients (70). It has also been suggested that deficiencies in H<sub>2</sub>S biosynthesis are associated with Alzheimer's disease (see Ref. 37, reviewed in Ref. 130) and that exogenous H<sub>2</sub>S may have therapeutic potential by reducing amyloid beta protein plaques (201). H<sub>2</sub>S has been proposed to modulate nociception (40, 144), induce μ opioid receptor-dependent analgesia (30), prevent neurodegeneration and movement disorders in mouse models of Parkinson's disease (55, 72), and may reduce the stress response of the hypothalamic-pituitary-adrenal axis (102). It has also been proposed to antagonize homocysteine-induced neurotoxicity (162).

The protective effects of H<sub>2</sub>S have been demonstrated in a number of neurological systems. H<sub>2</sub>S has been shown to protect neurons against hypoxic injury (165), inhibit hypochlorous acid-mediated oxidative damage (183), and increase glutathione production and suppress oxidative stress in mitochondria (76). Conversely, H<sub>2</sub>S has been shown to mediate cerebral ischemic damage (129) and produce vanilloid receptor 1-mediated neurogenic inflammation in airways (170).

H<sub>2</sub>S increases cAMP production in neurons and subsequent activation of PKA may account portion of the LTP. Other functions of H<sub>2</sub>S include upregulation of GABA B receptor and neuronal hyperpolarization via K<sub>ATP</sub> channel activation and induction of calcium waves in astrocytes (130), regulation of intracellular pH in glial cells (98), and the above-mentioned increase in glutathione production. For recent reviews, see Refs. 56, 130, 160 and 144.

*H<sub>2</sub>S and the gastrointestinal system.* The initial interest in H<sub>2</sub>S in the gastrointestinal (GI) system stemmed from the well-known production of H<sub>2</sub>S by sulfate-reducing bacteria in the colon and the presumed need to protect tissues from this toxic molecule (133). Today, more is known about the effects of H<sub>2</sub>S in the colon than any other segment of the GI tract; however, anti-inflammatory actions of H<sub>2</sub>S in the stomach appear to be of important therapeutic value and other areas have received increased attention as well.

H<sub>2</sub>S is synthesized in the stomach, jejunum, ileum, and colon. CSE immunoreactivity is diffusely distributed throughout the gastrointestinal tract most likely due to its association with the vasculature, whereas CBS staining is predominantly in muscularis mucosa, cell mucosa, and lamina propria but not associated with goblet, crypt, and epithelial cells (105).

H<sub>2</sub>S relaxes smooth muscle in the stomach (28) intestine (113), and colon (29). The mechanisms of H<sub>2</sub>S on GI motility have not been fully resolved, and in most instances, we are merely left with a list of factors that do not affect motility. In the stomach H<sub>2</sub>S acts partly via activation of myosin light-chain phosphatase (28); in the colon, the effects of H<sub>2</sub>S are independent of intracellular calcium and not mediated through known K<sup>+</sup> channels, myosin light-chain phosphatase, or Rho kinase (29), and in the ileum, H<sub>2</sub>S relaxation is independent of extrinsic or enteric nerves, NO, K<sub>ATP</sub>, and KCa<sup>+</sup> channels (113). H<sub>2</sub>S inhibits pacemaker activity of mouse small intestine interstitial cells of Cajal by modulating intracellular calcium through mechanisms independent of K<sup>+</sup> channels (122). Proliferation of these interstitial cells is also stimulated by H<sub>2</sub>S, which acts via phosphorylation of AKT protein kinase (57).

H<sub>2</sub>S stimulates chloride secretion in the intestine by targeting vanilloid receptors (transient receptor potential vanilloid 1) on afferent nerves, which, in turn, activate cholinergic secretomotor neurons via release of substance P (79).

H<sub>2</sub>S has both anti-inflammatory and inflammatory effects in the GI tract; however, the former is perhaps better characterized and appears to be of therapeutic value. In the colon, H<sub>2</sub>S is anti-inflammatory and enhances ulcer healing, independent of nitric oxide synthase and K<sub>ATP</sub> channel involvement (176). H<sub>2</sub>S production is increased in experimental models of colitis and H<sub>2</sub>S protects against and promotes resolution of this colitis (177). However, H<sub>2</sub>S modulates the expression of genes involved in cell-cycle progression and may trigger both inflammatory and DNA repair processes, which may contribute to colorectal cancer (5).

In the pancreas, H<sub>2</sub>S is a mediator of inflammatory caerulein-induced pancreatitis (17, 158, 159). H<sub>2</sub>S acts through ICAM-1 expression and stimulates neutrophil adhesion through the NF- $\kappa$ B and Src-family kinases (157). However, H<sub>2</sub>S has also been shown to protect pancreatic  $\beta$  cells from oxidative stress (164).

Inhibition of CSE, which is found in both hepatocytes and the bile duct, stimulates biliary bicarbonate secretion, whereas exogenous H<sub>2</sub>S inhibits it (39). Bile acids increase liver CSE expression via activation of the farnesoid X receptor, the resultant H<sub>2</sub>S production is proposed to maintain vasodilation and minimize the chance for portal hypertension (131). For recent reviews, see Refs. 64, 71, 96, 106, 133, and 175.

*H<sub>2</sub>S and the cardiovascular system.* Collectively, the involvement of H<sub>2</sub>S on heart and blood vessel physiology has received more attention than any other organ system, even though the therapeutic applications of H<sub>2</sub>S are less evident.

The vasodilatory effects of H<sub>2</sub>S on systemic blood vessels were the first cardiovascular effects of this transmitter described (54). This has been confirmed repeatedly and even observed in pulmonary arteries of diving mammals (119). H<sub>2</sub>S-induced relaxation appears to depend on extracellular Ca<sup>2+</sup> (203), and although K<sub>ATP</sub> channels, are frequently assumed to mediate the H<sub>2</sub>S relaxation (63, 86, 203, 204), this mechanism typically accounts for no more than half of the relaxation in most vessels. In some animals, such as the mouse, K<sub>ATP</sub> channels are not involved at all in the response. H<sub>2</sub>S may also signal via other pathways, such as activation of adenylate cyclase, which, in turn, inhibits superoxide formation, NADPH oxidase, and Rac<sub>1</sub> activity (112); it may produce intracellular acidosis and alter intracellular redox status, stimulate an anion exchanger (97), or operate through Ca<sup>2+</sup>-dependent K<sup>+</sup> (K<sub>Ca</sub>) channels (77, 161, 206). Relaxation of rat aorta by exogenous H<sub>2</sub>S does not depend on vascular prostaglandin synthesis, PKC, or cAMP, nor does it involve superoxide or H<sub>2</sub>O<sub>2</sub> production (77, 78, 204). Observations that H<sub>2</sub>S sulfhydrates and may regulate biological activity of numerous proteins, including actin (109), suggests that additional key steps in H<sub>2</sub>S-mediated vascular signaling are soon to be unraveled. However, even this mechanism has been questioned on the basis of the seemingly nonselectivity and promiscuity of this process (96), and the suggestion that for this to occur, the cysteine residues must be in the oxidized state, and these are rare in the reducing intracellular environment (66). H<sub>2</sub>S may also indirectly relax blood vessels *in vivo* through its ability to

inhibit angiotensin-converting enzyme and thus prevent formation of the vasoconstrictor ANG II.

Recent evidence has turned to H<sub>2</sub>S as the elusive endothelium-derived hyperpolarization factor, the third endothelium-derived relaxing factor that, along with NO and prostacyclin, signals vasodilation (180). Crosstalk between H<sub>2</sub>S, NO, and CO has been suggested to contribute to vasoactivity and, although CO inhibits CBS (8), interactions between H<sub>2</sub>S and NO are far from resolved. NO production has been shown to be directly inhibited by H<sub>2</sub>S (81), or indirectly stimulated by it through activating NF- $\kappa$ B, which activates the ERK1/2, which, in turn, activates inducible nitric oxide synthase (iNOS) (62). H<sub>2</sub>S relaxations have been reported to be independent of NO synthesis or cGMP activation (77, 78, 203). As described above, NO does not appear to directly affect H<sub>2</sub>S production (8). There is also evidence that H<sub>2</sub>S and NO may form a simple S-nitrosothiol with vasoactive properties of its own (184).

Reports of H<sub>2</sub>S-mediated vasoconstrictory responses in mammalian systemic vessels are less common, and many of these show an endothelium-dependent effect that has been attributed to H<sub>2</sub>S inactivation of NO. Low concentrations of H<sub>2</sub>S (<200  $\mu$ M) produce endothelium-dependent contraction of human internal mammary arteries and rat and mouse aortas (2, 81, 181), and low-dose H<sub>2</sub>S infusion increases blood pressure in the rat (2). These contractions have been proposed to result from H<sub>2</sub>S inactivation of endothelial NO via production of an inactive nitrosothiol (2, 181), whereas Kubu et al. (81) showed that H<sub>2</sub>S directly inhibited NO production. Other studies suggest that H<sub>2</sub>S may have direct, albeit modest, constrictory effects on systemic vascular smooth muscle. Lim et al. (95) observed 1  $\mu$ M H<sub>2</sub>S contractions of rat aortas that were partially independent of both the endothelium and K<sub>ATP</sub> channels and due, in part, to down-regulation of cAMP. Direct H<sub>2</sub>S-mediated vasoconstriction has been demonstrated in systemic vessels of nonmammalian vertebrates, and H<sub>2</sub>S contracts pulmonary vessels in terrestrial mammals in response to hypoxia (32, 117, 118).

H<sub>2</sub>S has a variety of other effects on the vasculature that are not directly vasoactive. At times, the findings are contradictory, but nevertheless, many are suggestive of therapeutic potential. H<sub>2</sub>S has been shown to be both proinflammatory and anti-inflammatory, to reduce leukocyte adhesion, to inhibit platelet aggregation, and although it is proangiogenic, to reduce deleterious vascular remodeling that often accompanies vascular damage (35, 89, 155). H<sub>2</sub>S is not only a mild antioxidant, but it also stimulates cysteine uptake and synthesis of glutathione. H<sub>2</sub>S has been implicated in hypotension associated with septic and hypovolemic shock, and inappropriate H<sub>2</sub>S regulation of insulin secretion in type II diabetes may contribute to macrovascular and microvascular pathologies (85). Inhibition of plasma renin activity by H<sub>2</sub>S is antihypertensive in renin-dependent hypertensive rats (99) and can potentially augment the depressor effect of H<sub>2</sub>S vasodilation.

While H<sub>2</sub>S has been shown to have negative inotropic and chronotropic effects on the heart (207), most interest has centered around its cardioprotective abilities. Numerous studies have shown that transient application of H<sub>2</sub>S or H<sub>2</sub>S donors can mimic hypoxic preconditioning and postconditioning and that increased endogenous H<sub>2</sub>S biosynthesis can also protect the heart from ischemia/reperfusion injury (reviewed by Refs. 35, 83, 156). Furthermore, the potential for H<sub>2</sub>S-mediated

protection from ischemia/reperfusion injury has been demonstrated in a number of extracardiac organs, including the kidney (171), which presumably offsets the reduction in endogenous H<sub>2</sub>S production (192), liver and small intestine (52, 198), skeletal muscle (49, 51), and cellular components of cutaneous tissue (50). As in the vasculature, H<sub>2</sub>S has been proposed to combine with NO to produce a nitrosothiol with inotropic properties (195, 194). For recent reviews, see Refs. 11, 15, 36, 83, 114, 155, 156, 172, 180, and 207.

**H<sub>2</sub>S and the respiratory system.** Much of the focus of H<sub>2</sub>S activity in the lung has focused on pulmonary blood flow and pulmonary vascular resistance. Increasing pulmonary blood flow in rats via a aortocaval shunt decreases CSE mRNA and CSE-mediated H<sub>2</sub>S production (140). During chronic hypoxia and the associated pulmonary hypertension, plasma and lung tissue production of H<sub>2</sub>S is decreased (182, 199, 202) and CSE activity is suppressed (199). Hypoxic pulmonary hypertension is further increased after CSE inhibition with PPG (199, 202), whereas exogenous H<sub>2</sub>S reduces pulmonary arterial pressure (182, 199) but (surprisingly), this does not affect aortic pressure (199). Exogenous H<sub>2</sub>S also decreases tissue GSSG and increases total antioxidant capacity (182). H<sub>2</sub>S paradoxically constricts isolated resistance pulmonary arterioles in terrestrial mammals but dilates those of diving mammals (sea lions), which is consistent with the response of these vessels to hypoxia and the specific needs of the animal (119). H<sub>2</sub>S also relaxes precontracted mouse bronchial smooth muscle via a mechanism that is independent of K<sub>ATP</sub> channels, soluble guanylyl cyclase, cyclooxygenases 1 and 2, and tachykinins (80).

Plasma H<sub>2</sub>S is reported to decrease in rats with oelic acid-induced lung injury; exogenous H<sub>2</sub>S increases arterial PaO<sub>2</sub>, decreases pulmonary edema and infiltration of polymorphonuclear cells, decreases IL-6 and IL-8, but increases IL-10, suggesting that endogenous H<sub>2</sub>S production is decreased in this model of lung injury (93). CSE expression in airway and vascular smooth muscle decreases in ovalbumin-induced lung asthma, whereas exogenous H<sub>2</sub>S alleviates inflammation, restores expiratory flow, and attenuates iNOS activation (22). Ventilator-induced lung injury also enhances the inflammatory response, which is reversed by exogenous H<sub>2</sub>S (3). These studies suggest H<sub>2</sub>S is anti-inflammatory and anti-remodeling in a variety of lung pathologies in addition to hypoxia. For a recent review, see Ref. 120.

**H<sub>2</sub>S and the kidney.** H<sub>2</sub>S affects both the renal tubule and vasculature. H<sub>2</sub>S is produced in the kidney by combined actions of CBS and CSE (191). Simultaneous administration of AOA and PPG (but neither independently) decreases glomerular filtration rate (GFR) and sodium and potassium excretion, whereas these are increased by infusion of either H<sub>2</sub>S or Cys (191). In the two-kidney, one-clip rat model of renal vascular hypertension, exogenous H<sub>2</sub>S decreases blood pressure, decreases plasma renin activity and ANG II concentration (but it does not affect plasma angiotensin-converting enzyme activity), and it inhibits upregulation of renin mRNA (99). In a genetic model of hyperhomocysteinemia, H<sub>2</sub>S production is down-regulated, GFR decreases, glomerular inflammation increases, and these effects that can be reversed by exogenous H<sub>2</sub>S (138). In a mouse model of diabetic nephropathy induced by streptozotocin, plasma and renal cortex H<sub>2</sub>S decrease, TGF- $\beta$ 1 and collagen IV increase, and these changes are prevented by exogenous H<sub>2</sub>S (197). Upregulation of TGF- $\beta$ 1

and collagen IV and reduced CSE expression produced by high glucose in culture mesangial cells are also prevented by exogenous H<sub>2</sub>S (197). In human patients, hemodialysis appears to lower plasma H<sub>2</sub>S (125). For a recent review, see Ref. 14.

**H<sub>2</sub>S and reproduction.** Although CBS has been identified in Leydig, Sertoli, and germ cells, and CSE has been found in Sertoli cells and germ cells in the rat testis (151), most of the attention has been focused on the vasodilatory properties of H<sub>2</sub>S in the corpus cavernosum and the potential for H<sub>2</sub>S therapy in erectile dysfunction (26, 146). Human and rat vas deferens smooth muscle contains both CBS and CSE and is relaxed by H<sub>2</sub>S (88). H<sub>2</sub>S has also been shown to be synthesized by the rat uterus, fetal membranes, and placenta, as well as human placenta. CBS and CSE were identified in all rat intrauterine tissues, as well as in human placenta myometrium, amnion, and chorion (123). H<sub>2</sub>S also produces dilation in vaginal and clitoral cavernosal muscle strips in the rabbit (147). However, the role of H<sub>2</sub>S in reproduction per se is unknown.

**H<sub>2</sub>S interactions with heme proteins.** Because of the known interactions of NO and CO with iron centers in a variety of heme proteins, the ability of H<sub>2</sub>S to reduce methemoglobin, the estimation that acid-labile H<sub>2</sub>S could be released from a variety of cytochromes, and the inhibitory effect of H<sub>2</sub>S on cytochrome-*c* oxidase, it was only natural to assume that H<sub>2</sub>S would serve some physiological function in heme proteins. Although it has been observed that a modified hemoglobin, hemoglobin I (HbI) does indeed serve a physiological function in sulfide transport in the clam (*Lucina pectinata*), there does not seem to be an analogous activity in mammals. Relatively high H<sub>2</sub>S concentrations (3:1, H<sub>2</sub>S: hemoglobin) favor formation of sulfhemoglobin and sulfmyoglobin, which may lower the oxygen affinity by 135- and 2,500-fold, respectively, but this does not appear to be a physiological process because it is doubtful that H<sub>2</sub>S concentrations even approach these levels in vivo. While H<sub>2</sub>S concentrations may increase during toxic exposure, the adverse effect of sulfhemoglobin formation will be somewhat offset by a concomitant H<sub>2</sub>S-induced right shift in the oxyhemoglobin curve. In most cases, sulfhemoglobinemia toxicity is well tolerated and resolved by red blood cell replacement. Sulfheme formation requires a histidine residue in the heme environment. Such a histidine is lacking in cytochrome *c*, which, therefore, does not form a sulfheme when exposed to H<sub>2</sub>S (reviewed in Ref. 127). H<sub>2</sub>S has recently been shown to form sulfheme with human neuroglobin, which also appears to depend on a histidine residue (18). However, this reaction was produced by first forming ferric (Fe<sup>3+</sup>) neuroglobin, and the physiological significance of this oxidized form is unknown. For a recent review, see Ref. 127.

**H<sub>2</sub>S and oxygen sensing.** We initially proposed that H<sub>2</sub>S metabolism serves as an intrinsic oxygen sensor in the vasculature (117), and these observations have been extended to include systemic and pulmonary vessels from a variety of vertebrates (117, 119), in fish gill chemoreceptive cells (118), urinary bladder (31), and the mammalian carotid body (124, 166, 167). Key in this hypothesis is the ability of tissues to rapidly consume H<sub>2</sub>S in the presence of oxygen (Fig. 3A) and observations that the rate of H<sub>2</sub>S metabolism is coupled to tissue or mitochondrial oxygen at physiologically relevant P<sub>O</sub><sub>2</sub>s (Fig. 3B). The relationship between oxygen consumption and H<sub>2</sub>S production are considered in more detail below. Mechanisms of H<sub>2</sub>S-mediated vasodilation were described previously

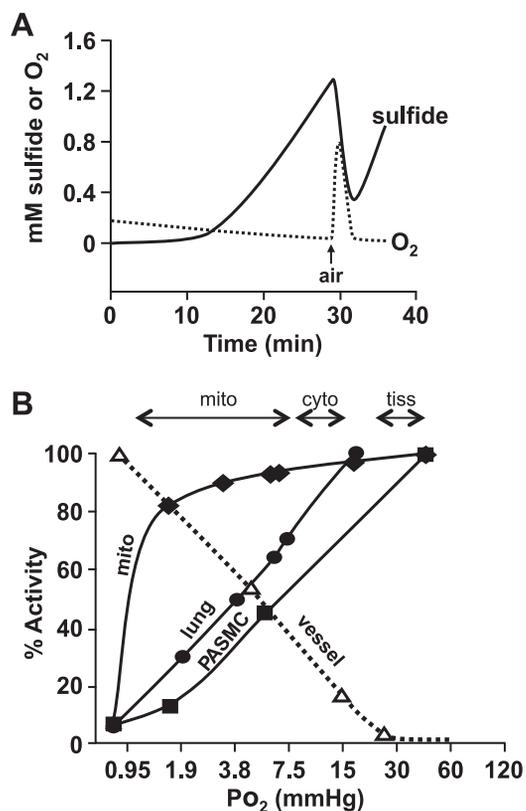


Fig. 3. Inverse relationship between H<sub>2</sub>S and O<sub>2</sub> in tissues. **A:** H<sub>2</sub>S production by homogenized rat lung is converted to net H<sub>2</sub>S consumption in the presence of O<sub>2</sub>. Tissue was deoxygenated by gassing with 100% nitrogen and primed with cysteine (1 mM) and  $\alpha$ -ketoglutarate (1 mM) and placed in a sealed container. H<sub>2</sub>S and O<sub>2</sub> were continuously recorded with amperometric electrodes. H<sub>2</sub>S is expressed as total sulfide (H<sub>2</sub>S plus HS<sup>-</sup>) calculated from tissue pH. As O<sub>2</sub> falls, H<sub>2</sub>S concentration increases; injection of a small air bubble (arrow) immediately decreases H<sub>2</sub>S concentration, which then resumes after the O<sub>2</sub> has been consumed. **B:** effect of O<sub>2</sub> on H<sub>2</sub>S consumption by pulmonary arterial smooth muscle cells (PASMC), homogenized bovine lung (lung), and purified mitochondria (mito) compared with O<sub>2</sub> dependence of hypoxic pulmonary vasoconstriction of isolated bovine pulmonary arteries (vessel). Percent activity refers to the degree of H<sub>2</sub>S consumption (100% = all H<sub>2</sub>S consumed) or the percentage of hypoxic contraction (100% = maximum vessel contraction). Half-maximal H<sub>2</sub>S consumption and vessel contraction occurs at approximately the same PO<sub>2</sub>. [A: modified from Olson and Whitfield (118); B: modified from Olson et al. (119).]

in H<sub>2</sub>S and the cardiovascular system. The mechanism of H<sub>2</sub>S-mediated hypoxic vasoconstriction remains to be identified, although it may be similar to activation of type 1 glomus cells in the mammalian carotid body, where H<sub>2</sub>S inhibits large-conductance calcium-sensitive potassium (BK<sub>Ca</sub>) channels (166, 167). Although never directly addressed in the literature, it seems likely that the hypoxia-induced increase in tissue H<sub>2</sub>S also is the initial stimulus in preconditioning and postconditioning effects associated with reperfusion injury and may also contribute to the pathology of reperfusion injury in unconditioned tissues. For recent reviews, see 14, 116, and 118.

**H<sub>2</sub>S and metabolism.** Recent studies showing that inhaled H<sub>2</sub>S can induce a “suspended animation-like state” in small mammals have heightened expectations of using H<sub>2</sub>S treatment clinically (4). Potential applications of this hypometabolic state, in addition to protection from ischemia/reperfusion injury described above, include organ preservation prior to trans-

plantation, protective metabolic depression during bypass surgery or following severe trauma associated with shock, sepsis, and acute lung injury (4). However, attempts to induce similar metabolic depression and protection in large mammals have produced conflicting results, and clearly, much needs to be done in this field prior to clinical applicability. Even in small rodents, the metabolic effects of H<sub>2</sub>S remain unclear. Baumgart et al. (12) showed in small rodents that while H<sub>2</sub>S inhalation during hypothermia did not alter the hemodynamic and cardiac effects of hypothermia itself, it did improve mitochondrial respiration, and they proposed that this may be the benefit of exogenous H<sub>2</sub>S during hypothermia. Interestingly, the authors also found that the H<sub>2</sub>S exposure increased aerobic glucose utilization. This observation is difficult to reconcile with the metabolic studies of Bouillaud and Blachier (24), who showed a mitochondrial preference for sulfide oxidation over other carbon-based substrates; see *Metabolism (Inactivation)*. Clearly, more needs to be learned regarding the transition from mitochondrial oxidation of H<sub>2</sub>S as an energy source to the inhibitory effects of H<sub>2</sub>S on oxidative phosphorylation. For a recent review, see Ref. 4; for other general reviews, see Refs. 42, 48, 58, 67, 74, 75, 89, 90, 101, 110, 121, 136, 185, and 200.

#### Disorders of H<sub>2</sub>S Metabolism

Relatively few clinical conditions are currently attributable to H<sub>2</sub>S metabolism. Ethylmalonic encephalopathy is an autosomal recessive disorder characterized by early-onset encephalopathy, microangiopathy, chronic diarrhea, and defective cytochrome-*c* oxidase (168). The gene *ETHE1* encodes ETHE1, a mitochondrial dioxygenase, the absence of which appears to adversely affect mitochondrial H<sub>2</sub>S oxidation, resulting in elevated tissue H<sub>2</sub>S and associated toxicity. Paradoxically, however, H<sub>2</sub>S<sub>3</sub>O<sub>2</sub> levels are also increased. Deficiencies in CBS activity result in hyperhomocysteinemia and the associated cardiovascular, ocular, neural, and skeletal problems (103, 187). CSE deficiency produces cystathioninuria and is secondarily associated with a wide range of diseases, including diabetes insipidus, Down syndrome, neuroblastoma, hepatoblastoma, and celiac disease; however, it is not associated with any overt clinical abnormalities (66). Mercaptolactate-cysteine disulfiduria is associated with mental retardation (66). For reviews, see Refs. 66, 103, 137, and 187.

#### H<sub>2</sub>S Donating Drugs

An appreciation, if not understanding, of the health benefits of sulfur springs, garlic, and cruciferous vegetables has been known since ancient times. Recent evidence suggests that the common denominator in these folk remedies may be their ability to produce H<sub>2</sub>S. Considerable effort is now under way to study and promote dietary intake of these sulfur-containing foods and in the synthesis of novel orally active compounds. The latter approach has proven especially effective when the H<sub>2</sub>S-donating compound is attached to another drug. A few of the more common H<sub>2</sub>S-donating drugs of this type are shown in Fig. 4. Additional drugs are described in recent reviews (20, 104), and the progress of these drugs in clinical trials can be accessed from the website “www.clinicaltrials.gov”. A list of patent applications on H<sub>2</sub>S-releasing molecules and dosages can be found in Bannenberg and Vieira (9).

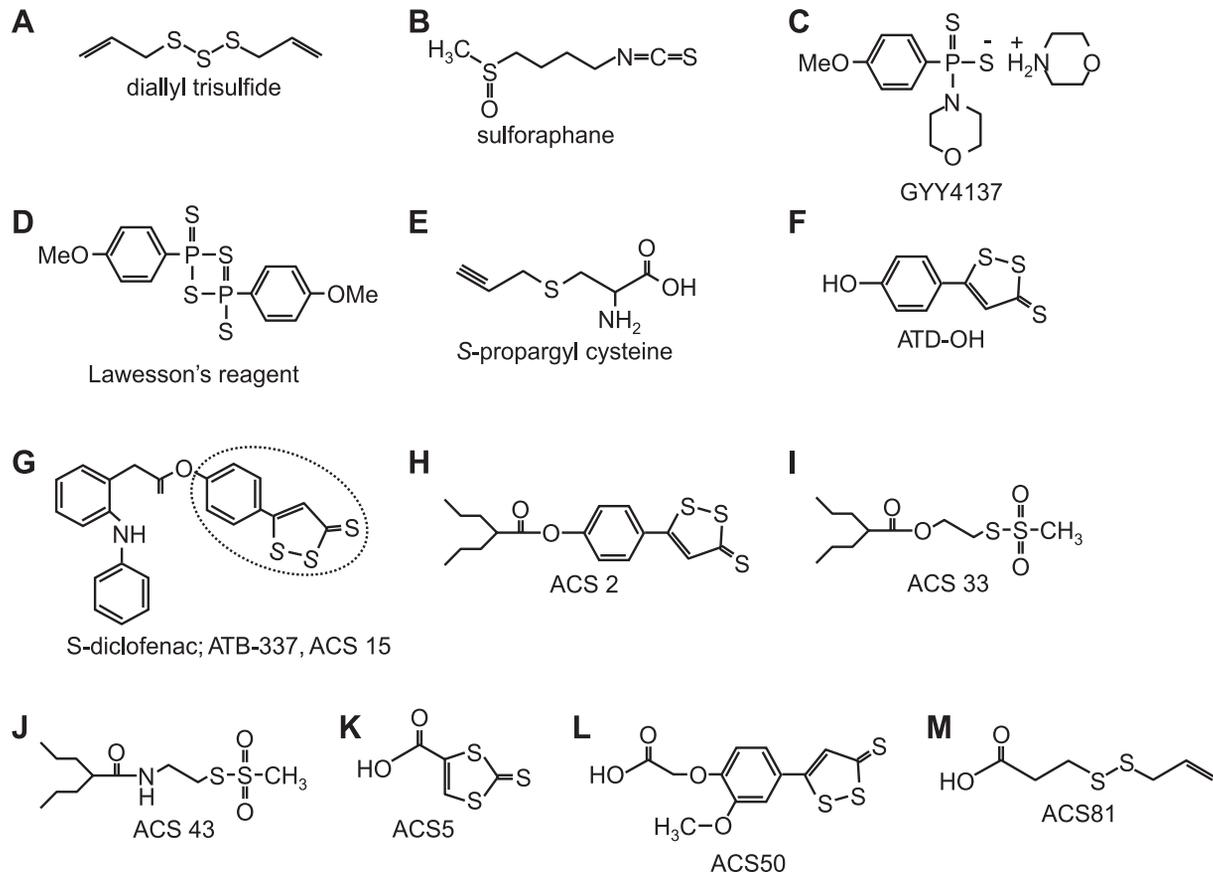


Fig. 4. H<sub>2</sub>S-donating compounds. *A*: diallyl trisulfide, one of two active components of garlic from which H<sub>2</sub>S is synthesized. *B*: sulforaphane, the sulfur-containing moiety in broccoli. *C*: H<sub>2</sub>S-releasing compound morpholin-4-ium 4-methoxyphenyl(morpholino) phosphinodithioate (GYY4137). *D*: Lawesson's reagent. *E*: S-propargyl cysteine, a cysteine analog. *F*: anethole trithione hydroxide (ADT-OH), the dithiolethione sulfur donor, is frequently added to a variety of compounds such as diclofenac (*G*) or valproate (*H*). *I*, *J*: methanethiosulfonate derivatives of valproate. *K*–*M*: other H<sub>2</sub>S donors that have been combined with L-DOPA. ATB, Antibe Threapeutics (Hamilton, Canada); ACS, CTG-Pharma, (Milan, Italy).

**Naturally occurring H<sub>2</sub>S-donating drugs.** The best characterized, naturally occurring H<sub>2</sub>S-donating compound from garlic (*Allium sativum*) is allicin (diallyl thiosulfinate), which decomposes in water to a number of compounds. Two of these, diallyl disulfide and diallyl trisulfide (or DATS; Fig. 4A) are the most efficacious H<sub>2</sub>S donors and readily vasodilate rat aortas (16). It should be noted garlic-mediated H<sub>2</sub>S production by red blood cells and in buffer was determined under anoxic conditions, and H<sub>2</sub>S production by rat aorta was measured at P<sub>O<sub>2</sub></sub> < 36 mmHg. How these reactions proceed under more physiological conditions (e.g., P<sub>O<sub>2</sub></sub>) remains to be determined.

Sulforaphane (Fig. 4B), the isothiocyanate compound from broccoli (*Brassica oleracea*), protects vascular smooth muscle cells and endothelial cells from oxidative and inflammatory stress and suppresses angiogenesis (61, 139, 208). It also protects hearts from ischemia reperfusion-induced injury (108). Sulforaphane also has neuroprotective and anti-inflammatory actions mediated, in part, through activation of heme oxygenase-1 (HO-1) and provides some protection against ischemia-reperfusion injury, hemorrhage, and serotonin-induced toxicity (19). Sulforaphane is rapidly absorbed by humans, reaching peak concentrations at 1 h and declining thereafter with a half-life of 1.8 h (193). A related isothiocyanate compound, erucin, is found in high levels in rocket salad species (*Eruca sativa*) and produces a concentration-dependent

induction of a number of cellular antioxidants and enzymes. It has not yet been determined whether either sulforaphane or erucin are metabolized to H<sub>2</sub>S, or if erucin has any beneficial effects on the cardiovascular system.

**Synthetic H<sub>2</sub>S-donating drugs.** A number of H<sub>2</sub>S compounds have been synthesized with the intent of slowing the rate of H<sub>2</sub>S release and thereby avoiding the transient H<sub>2</sub>S surge characteristic of the sulfide salts. GYY4137 [morpholin-4-ium 4-methoxyphenyl(morpholino) phosphinodithioate (Cayman Chemical, Ann Arbor, MI); Fig. 4C] (92) is a water-soluble molecule that is reported to slowly (over 90 min) release H<sub>2</sub>S in acidic phosphate buffer. When GYY4137 was injected intraperitoneally or intravenously into rats, plasma H<sub>2</sub>S increased from control 33 μmol/l to ~80 μmol/l in 30 min and was still elevated (50 μmol/l) 3 h later. By comparison, NaHS rapidly (within seconds) generated H<sub>2</sub>S in buffer and did not affect plasma H<sub>2</sub>S concentration when injected intravenously. GYY4137 produced a K<sub>ATP</sub> channel-mediated relaxation of rat aortas and dilated the perfused kidney. In vivo, GYY4137 exhibits antihypertensive activity. Curiously, H<sub>2</sub>S generation from either GYY4137 or NaHS in phosphate buffer was measured in real time with sensitive amperometric electrodes, whereas following injection of these compounds in vivo, plasma H<sub>2</sub>S was measured with an indirect and questionable methylene blue method (see *Separating Hype from Hope*).

Lawesson's reagent (Fig. 4D), another H<sub>2</sub>S donor has been used with some success as an anti-inflammatory drug in the stomach (176).

Stimulation of H<sub>2</sub>S production and the augmentation of H<sub>2</sub>S-like effects by exogenous cysteine are well known. Several cysteine analogs have been synthesized to mimic these effects such as *S*-propyl cysteine, *S*-allyl cysteine (in garlic), and *S*-propargyl cysteine, and they exert the expected cardioprotective effects (178). *S*-propargyl cysteine has also shown promise in preventing cognitive impairment in a rat model of Alzheimer's disease (44), and its structure is shown in Fig. 4E. In a patent application (<http://www.faqs.org/patents/app/20090036534>) *S*-propargyl cysteine and *S*-allyl cysteine (50 mg·kg<sup>-1</sup>·day<sup>-1</sup> ip for 7 days) was reported to increase plasma H<sub>2</sub>S in rats with myocardial injury from 34.7 to 91.6 and 61.1 μM, respectively.

Additional benefits have been realized by complexing H<sub>2</sub>S donors with other clinically efficacious drugs. The bulk of this work has successfully focused on blending the protective effects of H<sub>2</sub>S with NSAIDs that of themselves often have adverse side effects in the gastrointestinal tract (174). Anethole trithione (ADT-OH; Fig. 4F), a dithiolethione, is one of the most commonly used H<sub>2</sub>S donors. This molecule has been combined with numerous NSAIDs, including aspirin (ACS14; CTG-Pharma, Groton, CT), diclofenac (ATB-337; Antibe Therapeutics, Calgary, AL, Canada; ACS 15, *S*-diclofenac; Fig. 4G), indomethacin (ATB-343), mesalamine (ATB-429) and sulindac (*S*-sulindac). Dithioline derivatives of sildenafil (ACS6), valproate (ACS 2, *S*-valproate; Fig. 4H) and the anti-glaucoma drug latanoprost (ACS-67) have been developed, as have other methanethiosulfonate derivatives of valproate (ACS 33, Fig. 4I and ACS 43, Fig. 4J) (20, 126). A number of H<sub>2</sub>S-releasing molecules with potential antioxidant and anti-inflammatory properties (Fig. 4, *K–M*) have been coupled to levodopa (84). A patent application for ADT-OH conjugated with the angiotensin AT<sub>1</sub> receptor inhibitor losartan (H<sub>2</sub>S-EXP 3714) has been reported (104), but efficacy studies have not been published.

The effects of H<sub>2</sub>S-donating drugs are beginning to be examined. In cultured rat aortic smooth muscle cells, *S*-diclofenac, but not diclofenac, dose-dependently inhibits cell proliferation and survival (10). *S*-diclofenac (47.2 μmol/kg ip) has no effect on blood pressure or heart rate over 180 min but down-regulates expression of genes encoding enzymes synthesizing nitric oxide, prostanoids, and H<sub>2</sub>S. *S*-diclofenac also reduces plasma IL-1β/TNF-α, elevates plasma IL-10, and increases plasma H<sub>2</sub>S concentration from 25 to 37 μmol/l at 45 min and to 33 μmol/l 6 h post injection (91). *S*-diclofenac perfusion (10 and 30 μM) protects against ischemia-reperfusion injury in the isolated rat heart, which is mediated partly by opening K<sub>ATP</sub> channels, and possibly by increasing cysteine uptake and GSH synthesis. In this model, *S*-diclofenac also reduces creatine kinase and lactate dehydrogenase release and decreases the inhibitory effect of diclofenac on protective PGI<sub>2</sub> production (132). Intravenous injection of H<sub>2</sub>S-releasing aspirin (ACS14; 0.1 mmol/kg) produces a slight transient rise in plasma H<sub>2</sub>S [0.55 to 1.4 μmol/l that remains slightly elevated (0.62 μmol/l) at 150 min (145)]. Oral administration for 7 days of equimolar doses (~0.12 mmol/kg) of ACS14, ACS21 (a metabolite of ACS 14), or ADT-OH has no effect on systemic blood pressure and heart rate, but increases plasma GSH and cardiac and aortic GSH. ACS14 also produces a concentration-

dependent increase in HO-1 promoter activity in NIH3T3-HO-1-*luc* cells. ACS14 does not adversely affect aspirin's ability to inhibit thromboxane synthesis (145). Intraperitoneal injection of ACS14 or ADT-OH (both 0.1 mmol/kg) increases plasma H<sub>2</sub>S from 0.4 to ~0.65 μM in 15 min with a 2nd peak (0.5 μM) at 12–24 h, which is attributed to elevated plasma cysteine and subsequent metabolism to H<sub>2</sub>S. Both ACS14 and ADT-OH decrease plasma homocysteine and malonyldialdehyde (an indicator of oxidative stress) and increase total (reduced plus disulfide) cysteine and GSH. These drugs may have additional cardiovascular benefit by lowering plasma homocysteine (43).

H<sub>2</sub>S-donating sildenafil (ACS6) combines the inhibitory action of sildenafil on PKG with an H<sub>2</sub>S inhibition of PKA in porcine pulmonary arterial endothelial cells. Collectively, these inhibit superoxide formation and gp91<sup>phox</sup> expression, suggesting ACS6 may be effective in treating adult respiratory distress syndrome. H<sub>2</sub>S release from 10 μM NaHS peaks in 30 min, whereas H<sub>2</sub>S release from 10 μM ACS6 peaks in 120 min. Furthermore, more H<sub>2</sub>S is released from ACS6 than NaHS and only 25% mol/mol H<sub>2</sub>S is released from either NaHS or ACS6. ACS6 release of H<sub>2</sub>S is ~4 times greater when incubated with endothelial cells than in buffer (111). In rabbit corpus cavernosum, ACS6 protects against oxidative stress by stimulating both PKA (H<sub>2</sub>S effect) and PKG (sildenafil effect). There is no specific vasodilatory benefit from H<sub>2</sub>S released by ACS6 (142), which seems to be more or less typical for all synthetic H<sub>2</sub>S-donating drugs.

Sodium sulfide (NaHS) and sodium sulfide (Na<sub>2</sub>S) have long been used to generate H<sub>2</sub>S. While these are frequently called "H<sub>2</sub>S donors", and have even been reported to slowly release H<sub>2</sub>S (111), they are sulfide salts, and when placed in water, their dissociation and subsequent H<sub>2</sub>S formation are nearly instantaneous (84). Caution should be exercised with Na<sub>2</sub>S, as this is a strong alkali. Na<sub>2</sub>S in sterile, buffered solution is produced by Ikaria as IK-1001 and is currently in clinical trials for reduction of reperfusion/injury (NCT00858936). Calcium sulfide (CaS) has recently been shown to have similar actions (94) but does not appear to convey any distinct advantage over the other salts. For recent reviews, see Refs. 9, 20, 104, 128, 153, 154, 173, and 205.

### Separating Hype from Hope

There is little doubt that exogenous H<sub>2</sub>S affects a myriad of physiological systems, and many studies have been corroborated to some extent by compounds that can theoretically alter endogenous H<sub>2</sub>S production. Moreover, the beneficial results obtained with the H<sub>2</sub>S-"donating" compounds is encouraging and bespeaks of wide-ranging potential. However, as is often the case with a novel and exciting field, critical details can be missed or overlooked in the accompanying exuberance. The following sections take a careful look at a number of areas that require a thoughtful and more thorough analysis.

*H<sub>2</sub>S chemistry—what can H<sub>2</sub>S do?* In their review, Sen et al. (137) state that "H<sub>2</sub>S is a strong oxidant," and although H<sub>2</sub>S is more commonly thought of as a strong reducing agent, Kabil and Banerjee (66) show it is a relatively weak reducing agent, especially compared with other intracellular thiols, such as glutathione. H<sub>2</sub>S has also been proposed to serve as a potent antioxidant; however, its very low concentration in tissues argues against this as well, and the protective effects cannot be completely accounted for by direct reactions with oxidants (21). Furthermore, because H<sub>2</sub>S inhibits oxidative phosphory-

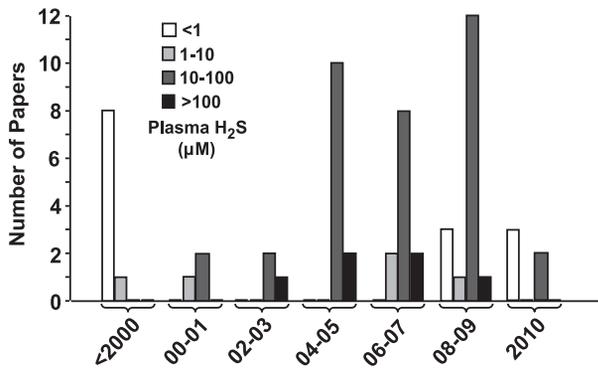


Fig. 5. Frequency distribution of papers reporting plasma H<sub>2</sub>S at various concentrations. Prior to 2000, nearly all studies reported H<sub>2</sub>S as undetectable (<math><1</math>). Subsequently, the number of studies reporting H<sub>2</sub>S between 10 and 100 μM has progressively increased (most between 20 and 40 μM), with a few studies reporting values in excess of 100 μM (most between 200 and 300 μM). [Modified from Olson (116).]

lation at ~20–40 μM (6, 24) and these concentrations were greatly exceeded in experimental conditions involving either exogenous H<sub>2</sub>S or H<sub>2</sub>S donors, it can be difficult to separate specific H<sub>2</sub>S effects from general metabolic depression or even more general reducing and/or antioxidant actions.

**Blood and tissue H<sub>2</sub>S concentrations—what is real and what is an artifact?** The overwhelming majority of studies and reviews on the biology of H<sub>2</sub>S refer to “physiological” concentrations of 20–40 μM H<sub>2</sub>S in blood (with some reports of plasma H<sub>2</sub>S approaching 300 μM). In turn, 20–300 μM H<sub>2</sub>S has been used to validate many “physiological” experiments. However, reports of plasma H<sub>2</sub>S > 1 μM have been relatively recent; prior to the year 2000, most reported values were <math><1</math> μM (Fig. 5), and these early studies were largely ignored by those that followed. That plasma H<sub>2</sub>S levels > 10 μM are unrealistic has been argued on the basis of both practical and methodological considerations (116). In addition, the use of newer methods, especially a polarographic (amperometric) sensor that directly measures H<sub>2</sub>S gas, fails to find plasma H<sub>2</sub>S approaching even 1 μM (186). There are also a number of other practical considerations, namely 1) there is no odor of H<sub>2</sub>S in plasma, which there should be even at 1 μM, 2) H<sub>2</sub>S rapidly equilibrates across the lung and would be readily exhaled if it existed in plasma (59, 169), 3) many of the reported plasma values would exceed toxic levels based on air quality standards (Table 1), 4) relative to point 2, it has been calculated that there is insufficient sulfur in the body to sustain H<sub>2</sub>S production at this level (41), 5) H<sub>2</sub>S is rapidly consumed by tissues in the presence of oxygen (Fig. 3) (82, 118, 138), and 6) H<sub>2</sub>S inhibits mitochondrial respiration at 20–40 μM. Furthermore, because H<sub>2</sub>S is metabolized by the mitochondria as fast as it is generated under normoxic conditions, H<sub>2</sub>S will increase only when tissue P<sub>O</sub><sub>2</sub> falls.

In a recent study, Wintner et al. (188) examined kinetics of plasma H<sub>2</sub>S using a newly developed monobromodimane-based assay (MBA) and compared this to the polarographic sensor. Similar to the findings of Whitfield et al. (186), they observed that exogenous H<sub>2</sub>S rapidly disappears from whole blood but disappears slowly when added to plasma or buffer. However, when measured with the MBA, total sulfide increased and decayed slowly in blood, similar to that observed in plasma and buffer. Bolus injections or continuous infusion

of H<sub>2</sub>S exhibited similar discrepancies between the MBA and polarographic measurements, i.e., more sulfide is detected by the former. The authors propose that the MBA is measuring sulfide in a “reversible sulfide sink,” which is most likely a persulfide. This, they propose can be readily mobilized under physiological conditions and represents the true “biologically available sulfide in vivo.” There are several problems with this assumption. First, this persulfide cannot be in the plasma as the MBA and polarographic responses are the same, and therefore, it must reside in red blood cells. Second, the amount of sulfide injected or infused greatly exceeds the amount measured by the MBA; assuming plasma volume is 4% of body weight, bolus injection 4 mg/kg of sodium sulfide would theoretically increase plasma sulfide to 312 μM, yet the MBA only measured 4.5 μM sulfide, less than 2% of the injected dose. Infusion of sulfide produces similar results, after 1 h of infusion of 20 mg·kg<sup>-1</sup>·h<sup>-1</sup>, blood sulfide measured with the MBA was 4.0 μM, whereas if the sulfide was confined to the plasma, its theoretical concentration would be over 6,300 μM. Even if distributed throughout the entire body water, sulfide would still be in excess of 420 μM, again less than 1% of the infused dose. Third, the authors provide no evidence that this extra sulfide can, in fact, be readily mobilized and is, therefore, biologically available. Fourth, a simple experiment can test the hypothesis that exogenous sulfide can be released from a persulfide pool. In this experiment, H<sub>2</sub>S is measured with polarographic sensor, and a strong reducing agent, DTT, known to liberate sulfane sulfur, is added to blood before and after exogenous H<sub>2</sub>S. As shown in Fig. 6, raising the concentration of DTT in buffer to 10 mM increased current flow equivalent to approximately one-fifth of the response produced by increasing H<sub>2</sub>S concentration to 50 μM (Fig. 6A). A similar 10-mM increase in DTT

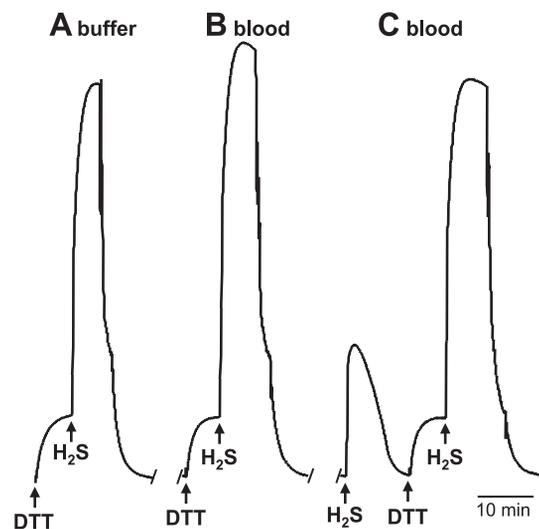


Fig. 6. Lack of evidence of sulfane sulfur in trout blood. A: DTT added to Cortland buffer produces a characteristic current in the polarographic H<sub>2</sub>S sensor. The addition of H<sub>2</sub>S increases current flow five-fold. B: similar additions of DTT and H<sub>2</sub>S to heparinized trout blood produce identical responses. C: H<sub>2</sub>S added to blood without prior DTT treatment rapidly disappears. The addition of DTT to H<sub>2</sub>S-treated blood produces the same response as DTT addition to Cortland (A) or untreated blood (B) and subsequent addition of H<sub>2</sub>S in the presence of DTT also produces the same response as H<sub>2</sub>S (A and B). Each addition of DTT will produce a final DTT concentration of 10 mM, and each addition of H<sub>2</sub>S will increase the final concentration by 50 μM (N. L. Whitfield, E. L. Kreimier, and K. R. Olson, unpublished data).

in whole blood produced an identical increase in current flow, and the presence of DTT did not affect the response to a subsequent addition of H<sub>2</sub>S (Fig. 6B). When H<sub>2</sub>S was added to untreated blood (Fig. 6C), the H<sub>2</sub>S was rapidly consumed. The addition of DTT to the H<sub>2</sub>S-spiked blood produced the same response as DTT addition to buffer or untreated blood, i.e., there was no evidence of liberation of H<sub>2</sub>S from sulfane sulfur over the 10-min treatment period. A second addition of H<sub>2</sub>S to the H<sub>2</sub>S-DTT-treated blood produced a response similar to that produced by the addition of H<sub>2</sub>S to the buffer or untreated blood. The consumption of H<sub>2</sub>S by untreated blood and the inability of DTT-treated blood to consume H<sub>2</sub>S suggest that H<sub>2</sub>S is consumed (presumably oxidized) in blood during reduction of another blood constituent and that this is prevented by the high concentrations of another strong reductant, DTT.

Thus, one must question whether a reported fall in plasma H<sub>2</sub>S concentration from 41 to 18 μmol/l in CSE gene-deleted mice is really key evidence for H<sub>2</sub>S as a “physiological” gasotransmitter of comparable importance to nitric oxide and carbon monoxide (42). Clearly, one must also question the numerous reports showing elevated (>10 μM) plasma H<sub>2</sub>S concentrations achieved with the H<sub>2</sub>S-donating drugs. Also worrisome are clinical trials that use these inaccurate methods to evaluate plasma hydrogen sulfide as a prognostic indicator of shock-related mortality (NTC 01088490).

Similar arguments can be made against excessive tissue H<sub>2</sub>S concentrations and tissue production. Recent studies have shown that H<sub>2</sub>S is consumed by tissues in the presence of oxygen, and H<sub>2</sub>S production is only observed under hypoxic or anoxic conditions (41, 118). In fact, even exogenous H<sub>2</sub>S is quickly and efficiently consumed by tissues at oxygen partial pressure (P<sub>O<sub>2</sub></sub>) greater than 10 mmHg (118). By comparison, water or tissue samples in equilibrium with room air typically have a P<sub>O<sub>2</sub></sub> greater than 140 mmHg. In addition, unphysiologically high cysteine (usually 10 mM, compared with normal <1 mM) is used in tissue production studies, and serine and homocysteine are typically absent. This can artificially increase the rate of H<sub>2</sub>S production and shuttle sulfur metabolism through normally minor metabolic pathways (149). It is also important to stress that, to date, no study has identified any stimulus for H<sub>2</sub>S production in cells in real time and under physiological conditions, other than showing an inverse relationship between H<sub>2</sub>S production and P<sub>O<sub>2</sub></sub>. For recent reviews, see Refs. 24, 96, 116, and 163.

*Does H<sub>2</sub>S fulfill the criteria of a “gasotransmitter”?* In a recent review, Linden et al. (96) critically evaluated the criteria for a “gasotransmitter” as originally proposed by Wang (179) and concluded that there are still sufficient questions remaining before H<sub>2</sub>S can be accepted as a biologically relevant signaling molecule. Foremost among these is the discrepancy between tissue and plasma concentrations and the dose of exogenous H<sub>2</sub>S needed to produce physiological responses. In addition, there is relatively little evidence regarding how H<sub>2</sub>S production or tissue concentrations are regulated. Presumably, these issues will be resolved when the methodology improves, as historically been the case with numerous other signaling molecules. There are other questions that must be resolved. Is H<sub>2</sub>S only transiently present in blood and tissue, and does it serve to initiate downstream effects that can be sustained for lengthy periods? Is H<sub>2</sub>S merely a by-product or intermediate of the “true” signaling molecule? Are there other more relevant

biochemical processes that are also inhibited by the promiscuity of “classical” inhibitors of H<sub>2</sub>S biosynthesis that have led us astray? If H<sub>2</sub>S is the principal signaling moiety, can H<sub>2</sub>S-donating drugs be targeted for tissue-specific H<sub>2</sub>S release? It would seem that these are pressing questions that, if answered, may save considerable time and effort as we attempt to target H<sub>2</sub>S pathways to treat the variety of diseases that have been implicated in H<sub>2</sub>S pathophysiology. For a recent review, see Ref. 96.

### *Perspectives and Significance*

H<sub>2</sub>S has been associated with life from the onset, at times supporting it as a useful substrate and at times destroying it. It is not surprising that through evolution, animals have learned to live in and around H<sub>2</sub>S and have ultimately incorporated this versatile molecule into their biochemistry. Recent awareness of this fact has greatly expanded the field of “gasotransmitters”. However, unlike its predecessor, nitric oxide, whose discovery was met with initial skepticism, H<sub>2</sub>S has been readily embraced by the scientific community and quickly targeted for its therapeutic potential. This exuberance has at times let enthusiasm reign over skepticism. As this field begins to mature, the need to recalibrate this balance is becoming increasingly evident.

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### DISCLOSURES

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### REFERENCES

1. Abe K, Kimura H. The possible role of hydrogen sulfide as an endogenous neuromodulator. *J Neurosci* 16: 1066–1071, 1996.
2. Ali MY, Ping CY, Mok YY, Ling L, Whiteman M, Bhatia M, Moore PK. Regulation of vascular nitric oxide in vitro and in vivo; a new role for endogenous hydrogen sulfide? *Br J Pharmacol* 149: 625–634, 2006.
3. Aslami H, Heinen A, Roelofs JJ, Zuurbier CJ, Schultz MJ, Juffermans NP. Suspended animation inducer hydrogen sulfide is protective in an in vivo model of ventilator-induced lung injury. *Intensive Care Med* 36: 1946–1952, 2010.
4. Aslami H, Schultz MJ, Juffermans NP. Potential applications of hydrogen sulfide-induced suspended animation. *Curr Med Chem* 16: 1295–1303, 2009.
5. Attene-Ramos MS, Nava GM, Muellner MG, Wagner ED, Plewa MJ, Gaskins HR. DNA damage and toxicogenomic analyses of hydrogen sulfide in human intestinal epithelial FHs 74 Int cells. *Environ Mol Mutagen* 51: 304–314, 2010.
6. Bagarinao T. Sulfide as an environmental factor and toxicant: tolerance and adaptations in aquatic organisms. *Aquat Toxicol (Amst)* 24: 21–62, 1992.
7. Balazy M, bu-Yousef IA, Harpp DN, Park J. Identification of carbonyl sulfide and sulfur dioxide in porcine coronary artery by gas chromatography/mass spectrometry, possible relevance to EDHF. *Biochem Biophys Res Commun* 311: 728–734, 2003.
8. Banerjee R, Zou CG. Redox regulation and reaction mechanism of human cystathionine-beta-synthase: a PLP-dependent hemesensor protein. *Arch Biochem Biophys* 433: 144–156, 2005.
9. Bannenberg GL, Vieira HL. Therapeutic applications of the gaseous mediators carbon monoxide and hydrogen sulfide. *Expert Opin Ther Pat* 19: 663–682, 2009.
10. Baskar R, Sparatore A, Del SP, Moore PK. Effect of S-diclofenac, a novel hydrogen sulfide releasing derivative inhibit rat vascular smooth muscle cell proliferation. *Eur J Pharmacol* 594: 1–8, 2008.
11. Baumgart K, Radermacher P, Wagner F. Applying gases for micro-circulatory and cellular oxygenation in sepsis: effects of nitric oxide,

- carbon monoxide, and hydrogen sulfide. *Curr Opin Anaesthesiol* 22: 168–176, 2009.
12. Baumgart K, Wagner F, Groger M, Weber S, Barth E, Vogt JA, Wachter U, Huber-Lang M, Knoferl MW, Albuszies G, Georgieff M, Asfar P, Szabo C, Calzia E, Radermacher P, Simkova V. Cardiac and metabolic effects of hypothermia and inhaled hydrogen sulfide in anesthetized and ventilated mice. *Crit Care Med* 38: 588–595, 2010.
  13. Bearden SE, Beard RS Jr, Pfau JC. Extracellular transsulfuration generates hydrogen sulfide from homocysteine and protects endothelium from redox stress. *Am J Physiol Heart Circ Physiol* 299: H1568–H1576, 2010.
  14. Beltowski J. Hypoxia in the renal medulla: Implications for hydrogen sulfide signaling. *J Pharmacol Exp Ther* 334: 358–363, 2010.
  15. Beltowski J, Jamroz-Wisniewska A, Tokarzewska D. Hydrogen sulfide and its modulation in arterial hypertension and atherosclerosis. *Cardiovasc Hematol Agents Med Chem* 8: 173–186, 2010.
  16. Benavides GA, Squadrito GL, Mills RW, Patel HD, Isbell TS, Patel RP, Darley-USmar VM, Doeller JE, Kraus DW. Hydrogen sulfide mediates the vasoactivity of garlic. *Proc Natl Acad Sci USA* 104: 17977–17982, 2007.
  17. Bhatia M, Wong FL, Fu D, Lau HY, Moomhala SM, Moore PK. Role of hydrogen sulfide in acute pancreatitis and associated lung injury. *FASEB J* 19: 623–625, 2005.
  18. Brittain T, Yosaatmadja Y, Henty K. The interaction of human neuroglobin with hydrogen sulphide. *IUBMB Life* 60: 135–138, 2008.
  19. Calabrese V, Cornelius C, nкова-Kostova AT, Calabrese EJ, Mattson MP. Cellular stress responses, the hormesis paradigm, and vitagenes: novel targets for therapeutic intervention in neurodegenerative disorders. *Antioxid Redox Signal* 13: 1763–1811 2010.
  20. Caliendo G, Cirino G, Santagada V, Wallace JL. Synthesis and biological effects of hydrogen sulfide (H<sub>2</sub>S): development of H<sub>2</sub>S-releasing drugs as pharmaceuticals. *J Med Chem* 53: 6275–6286, 2010.
  21. Carballal S, Trujillo M, Cuevasanta E, Bartsaghi S, Moller MN, Folkes LK, Garcia-Bereguain MA, Gutierrez-Merino C, Wardman P, Denicola A, Radi R, Alvarez B. Reactivity of hydrogen sulfide with peroxynitrite and other oxidants of biological interest. *Free Radic Biol Med* 50: 196–205, 2010.
  22. Chen YH, Wu R, Geng B, Qi YF, Wang PP, Yao WZ, Tang CS. Endogenous hydrogen sulfide reduces airway inflammation and remodeling in a rat model of asthma. *Cytokine* 45: 117–123, 2009.
  23. Chiku T, Padovani D, Zhu W, Singh S, Vitvitsky V, Banerjee R. H<sub>2</sub>S biogenesis by human cystathionine gamma-lyase leads to the novel sulfur metabolites lanthionine and homolanthionine and is responsive to the grade of hyperhomocysteinemia. *J Biol Chem* 284: 11601–11612, 2009.
  24. Collman JP, Ghosh S, Dey A, Decreau RA. Using a functional enzyme model to understand the chemistry behind hydrogen sulfide-induced hibernation. *Proc Natl Acad Sci USA* 106: 22090–22095, 2009.
  25. Curtis CG, Bartholomew TC, Rose FA, Dodgson KS. Detoxication of sodium 35 S-sulphide in the rat. *Biochem Pharmacol* 21: 2313–2321, 1972.
  26. d'Emmanuele di Villa Bianca R, Sorrentino R, Maffia P, Mirone V, Imbimbo C, Fusco F, De Palma R, Ignarro LJ, Cirino G. Hydrogen sulfide as a mediator of human corpus cavernosum smooth-muscle relaxation. *Proc Natl Acad Sci USA* 106: 4513–4518, 2009.
  27. DeBruyn WJ, Swartz E, Hu JH, Shorter JA, Davidovits P, Worsnop DR, Zahniser MS, Kolb CE. Henry's law solubilities and Setchenow coefficients for biogenic reduced sulfur species obtained from gas-liquid uptake measurements. *J Geophys Res* 100: 7245–7251, 1995.
  28. Dhaese I, Lefebvre RA. Myosin light chain phosphatase activation is involved in the hydrogen sulfide-induced relaxation in mouse gastric fundus. *Eur J Pharmacol* 606: 180–186, 2009.
  29. Dhaese I, van Colen, I, Lefebvre RA. Mechanisms of action of hydrogen sulfide in relaxation of mouse distal colonic smooth muscle. *Eur J Pharmacol* 628: 179–186, 2010.
  30. Distrutti E, Cipriani S, Renga B, Mencarelli A, Migliorati M, Cianetti S, Fiorucci S. Hydrogen sulphide induces micro opioid receptor-dependent analgesia in a rodent model of visceral pain [Online]. *Mol Pain* 6: 36, 2010.
  31. Dombkowski RA, Doellman MM, Head SK, Olson KR. Hydrogen sulfide mediates hypoxia-induced relaxation of trout urinary bladder smooth muscle. *J Exp Biol* 209: 3234–3240, 2006.
  32. Dombkowski RA, Russell MJ, Schulman AA, Doellman MM, Olson KR. Vertebrate phylogeny of hydrogen sulfide vasoactivity. *Am J Physiol Regul Integr Comp Physiol* 288: R243–R252, 2005.
  33. Dominy JE, Stipanuk MH. New roles for cysteine and transsulfuration enzymes: production of H<sub>2</sub>S, a neuromodulator and smooth muscle relaxant. *Nutr Rev* 62: 348–353, 2004.
  34. Dorman DC, Moulin FJ, McManus BE, Mahle KC, James RA, Struve MF. Cytochrome oxidase inhibition induced by acute hydrogen sulfide inhalation: correlation with tissue sulfide concentrations in the rat brain, liver, lung, and nasal epithelium. *Toxicol Sci* 65: 18–25, 2002.
  35. Elsey DJ, Fowkes RC, Baxter GF. L-cysteine stimulates hydrogen sulfide synthesis in myocardium associated with attenuation of ischemia-reperfusion injury. *J Cardiovasc Pharmacol Ther* 15: 53–59, 2010.
  36. Elsey DJ, Fowkes RC, Baxter GF. Regulation of cardiovascular cell function by hydrogen sulfide (H<sub>2</sub>S). *Cell Biochem Funct* 28: 95–106, 2010.
  37. Eto K, Asada T, Arima K, Makifuchi T, Kimura H. Brain hydrogen sulfide is severely decreased in Alzheimer's disease. *Biochem Biophys Res Commun* 293: 1485–1488, 2002.
  38. Evans CL. The toxicity of hydrogen sulphide and other sulphides. *Q J Exp Physiol* 52: 231–248, 1967.
  39. Fujii K, Sakuragawa T, Kashiba M, Sugiura Y, Kondo M, Maruyama K, Goda N, Nimura Y, Suematsu M. Hydrogen sulfide as an endogenous modulator of biliary bicarbonate excretion in the rat liver. *Antioxid Redox Signal* 7: 788–794, 2005.
  40. Fukushima O, Nishimura S, Matsunami M, Aoki Y, Nishikawa H, Ishikura H, Kawabata A. Phosphorylation of ERK in the spinal dorsal horn following pancreatic pronociceptive stimuli with proteinase-activated receptor-2 agonists and hydrogen sulfide in rats: Evidence for involvement of distinct mechanisms. *J Neurosci Res* 88: 3198–3205, 2010.
  41. Furne J, Saeed A, Levitt MD. Whole tissue hydrogen sulfide concentrations are orders of magnitude lower than presently accepted values. *Am J Physiol Regul Integr Comp Physiol* 295: R1479–R1485, 2008.
  42. Gadalla MM, Snyder SH. Hydrogen sulfide as a gasotransmitter. *J Neurochem* 113: 14–26, 2010.
  43. Giustarini D, Del SP, Sparatore A, Rossi R. Modulation of thiol homeostasis induced by H<sub>2</sub>S-releasing aspirin. *Free Radic Biol Med* 48: 1263–1272, 2010.
  44. Gong QH, Pan LL, Liu XH, Wang Q, Huang H, Zhu YZ. S-propargyl-cysteine (ZYZ-802), a sulphur-containing amino acid, attenuates β-amyloid-induced cognitive deficits and pro-inflammatory response: involvement of ERK1/2 and NF-κB pathway in rats. *Amino Acids* 40: 601–610, 2011.
  45. Gubern M, Andriamihaja M, Nubel T, Blachier F, Bouillaud F. Sulfide, the first inorganic substrate for human cells. *FASEB J* 21: 1699–1706, 2007.
  46. Guidotti TL. Hydrogen sulfide: advances in understanding human toxicity. *Int J Toxicol* 29: 569–581, 2010.
  47. Haggard HW. The fate of sulfites in the blood. *J Biol Chem* 49: 519–529, 1921.
  48. Hart JL. Role of sulfur-containing gaseous substances in the cardiovascular system. *Front Biosci* 3: 736–749, 2011.
  49. Henderson PW, Jimenez N, Ruffino J, Sohn AM, Weinstein AL, Krijgh DD, Reiffel AJ, Spector JA. Therapeutic delivery of hydrogen sulfide for salvage of ischemic skeletal muscle after the onset of critical ischemia. *J Vasc Surg* 53: 785–791, 2011.
  50. Henderson PW, Singh SP, Belkin D, Nagineni V, Weinstein AL, Weissich J, Spector JA. Hydrogen sulfide protects against ischemia-reperfusion injury in an in vitro model of cutaneous tissue transplantation. *J Surg Res* 159: 451–455, 2010.
  51. Henderson PW, Singh SP, Weinstein AL, Nagineni V, Rafii DC, Kadouch D, Krijgh DD, Spector JA. Therapeutic metabolic inhibition: hydrogen sulfide significantly mitigates skeletal muscle ischemia reperfusion injury in vitro and in vivo. *Plast Reconstr Surg* 126: 1890–1898, 2010.
  52. Henderson PW, Weinstein AL, Sohn AM, Jimenez N, Krijgh DD, Spector JA. Hydrogen sulfide attenuates intestinal ischemia-reperfusion injury when delivered in the post-ischemic period. *J Gastroenterol Hepatol* 25: 1642–1647, 2010.
  53. Hildebrandt TM, Grieshaber MK. Three enzymatic activities catalyze the oxidation of sulfide to thiosulfate in mammalian and invertebrate mitochondria. *FEBS J* 275: 3352–3361, 2008.
  54. Hosoki R, Matsuki N, Kimura H. The possible role of hydrogen sulfide as an endogenous smooth muscle relaxant in synergy with nitric oxide. *Biochem Biophys Res Commun* 237: 527–531, 1997.

55. **Hu LF, Lu M, Tiong CX, Dawe GS, Hu G, Bian JS.** Neuroprotective effects of hydrogen sulfide on Parkinson's disease rat models. *Aging Cell* 9: 135–146, 2010.
56. **Hu LF, Lu M, Wong PT, Bian JS.** Hydrogen sulfide: neurophysiology and neuropathology. *Antioxid Redox Signal*. In press.
57. **Huang Y, Li F, Tong W, Zhang A, He Y, Fu T, Liu B.** Hydrogen sulfide, a gaseous transmitter, stimulates proliferation of interstitial cells of Cajal via phosphorylation of AKT protein kinase. *Tohoku J Exp Med* 221: 125–132, 2010.
58. **Hughes MN, Centelles MN, Moore KP.** Making and working with hydrogen sulfide: The chemistry and generation of hydrogen sulfide in vitro and its measurement in vivo: a review. *Free Radic Biol Med* 47: 1346–1353, 2009.
59. **Insko MA, Deckwerth TL, Hill P, Toombs CF, Szabó C.** Detection of exhaled hydrogen sulphide gas in rats exposed to intravenous sodium sulphide. *Br J Pharmacol* 157: 944–951, 2009.
60. **Ishigami M, Hiraki K, Umemura K, Ogasawara Y, Ishii K, Kimura H.** A source of hydrogen sulfide and a mechanism of its release in the brain. *Antioxid Redox Signal* 11: 205–214, 2009.
61. **Jackson SJ, Singletary KW, Venema RC.** Sulforaphane suppresses angiogenesis and disrupts endothelial mitotic progression and microtubule polymerization. *Vascul Pharmacol* 46: 77–84, 2007.
62. **Jeong SO, Pae HO, Oh GS, Jeong GS, Lee BS, Lee S, Kim dY, Rhew HY, Lee KM, Chung HT.** Hydrogen sulfide potentiates interleukin-1 $\beta$ -induced nitric oxide production via enhancement of extracellular signal-regulated kinase activation in rat vascular smooth muscle cells. *Biochem Biophys Res Commun* 345: 938–944, 2006.
63. **Jiang B, Tang G, Cao K, Wu L, Wang R.** Molecular mechanism for H<sub>2</sub>S-induced activation of K(ATP) channels. *Antioxid Redox Signal* 12: 1167–1178, 2010.
64. **Jimenez M.** Hydrogen sulfide as a signaling molecule in the enteric nervous system. *Neurogastroenterol Motil* 22: 1149–1153, 2010.
65. **Julian D, Statile JL, Wohlgemuth SE, Arp AJ.** Enzymatic hydrogen sulfide production in marine invertebrate tissues. *Comp Biochem Physiol [A] Mol Integr Physiol* 133: 105–115, 2002.
66. **Kabil O, Banerjee R.** Redox biochemistry of hydrogen sulfide. *J Biol Chem* 285: 21903–21907, 2010.
67. **Kajimura M, Fukuda R, Bateman RM, Yamamoto T, Suematsu M.** Interactions of multiple gas-transducing systems: hallmarks and uncertainties of CO, NO, and H<sub>2</sub>S gas biology. *Antioxid Redox Signal* 13: 157–192, 2010.
68. **Kalir A, Kalir HK.** Biological activity of sulfinic acid derivatives. In: *The Chemistry of Sulfinic acids, Esters and Their Derivatives*, edited by Patai S. New York, NY: John Wiley and Sons, 1990, p. 665–676.
69. **Kamboures MA, Blake DR, Cooper DM, Newcomb RL, Barker M, Larson JK, Meinardi S, Nussbaum E, Rowland FS.** Breath sulfides and pulmonary function in cystic fibrosis. *Proc Natl Acad Sci USA* 102: 15762–15767, 2005.
70. **Kamoun P, Belardinelli MC, Chabli A, Lallouchi K, Chadeaux-Vekemans B.** Endogenous hydrogen sulfide overproduction in Down syndrome. *Am J Med Genet A* 116A: 310–311, 2003.
71. **Kasperek MS, Linden DR, Kreis ME, Sarr MG.** Gasotransmitters in the gastrointestinal tract. *Surgery* 143: 455–459, 2008.
72. **Kida K, Yamada M, Tokuda K, Marutani E, Kakinohana M, Kaneki M, Ichinose F.** Inhaled hydrogen sulfide prevents neurodegeneration and movement disorder in a mouse model of Parkinson's disease. *Antioxid Redox Signal*. In press.
73. **Kilburn KH, Thrasher JD, Gray MR.** Low-level hydrogen sulfide and central nervous system dysfunction. *Toxicol Ind Health* 26: 387–405, 2010.
74. **Kimura H.** Hydrogen sulfide: from brain to gut. *Antioxid Redox Signal* 12: 1111–1123, 2010.
75. **Kimura H.** Hydrogen sulfide: its production, release and functions. *Amino Acids*. In press.
76. **Kimura Y, Goto Y, Kimura H.** Hydrogen sulfide increases glutathione production and suppresses oxidative stress in mitochondria. *Antioxid Redox Signal* 12: 1–13, 2010.
77. **Kiss L, Deitch EA, Szabó C.** Hydrogen sulfide decreases adenosine triphosphate levels in aortic rings and leads to vasorelaxation via metabolic inhibition. *Life Sci* 83: 589–594, 2008.
78. **Koenitzer JR, Isbell TS, Patel HD, Benavides GA, Dickinson DA, Patel RP, Darley-Usmar VM, Lancaster JR Jr, Doeller JE, Kraus DW.** Hydrogen sulfide mediates vasoactivity in an O<sub>2</sub>-dependent manner. *Am J Physiol Heart Circ Physiol* 292: H1953–H1960, 2007.
79. **Krueger D, Foerster M, Mueller K, Zeller F, Slotta-Huspenina J, Donovan J, Grundy D, Schemann M.** Signaling mechanisms involved in the intestinal pro-secretory actions of hydrogen sulfide. *Neurogastroenterol Motil* 22: 224–231, 2010.
80. **Kubo S, Doe I, Kurokawa Y, Kawabata A.** Hydrogen sulfide causes relaxation in mouse bronchial smooth muscle. *J Pharm Sci* 104: 392–396, 2007.
81. **Kubo S, Doe I, Kurokawa Y, Nishikawa H, Kawabata A.** Direct inhibition of endothelial nitric oxide synthase by hydrogen sulfide: contribution to dual modulation of vascular tension. *Toxicology* 232: 138–146, 2007.
82. **Lagoutte E, Mimoun S, Andriamihaja M, Chaumontet C, Blachier F, Bouillaud F.** Oxidation of hydrogen sulfide remains a priority in mammalian cells and causes reverse electron transfer in colonocytes. *Biochim Biophys Acta* 1797: 1500–1511, 2010.
83. **Lavu M, Bhushan S, Lefter DJ.** Hydrogen sulfide-mediated cardioprotection: mechanisms and therapeutic potential. *Clin Sci* 120: 219–229, 2010.
84. **Lee M, Tazzari V, Giustarini D, Rossi R, Sparatore A, Del SP, McGeer E, McGeer PL.** Effects of hydrogen sulfide-releasing L-DOPA derivatives on glial activation: potential for treating Parkinson disease. *J Biol Chem* 285: 17318–17328, 2010.
85. **Lefter DJ.** Potential importance of alterations in hydrogen sulphide (H<sub>2</sub>S) bioavailability in diabetes. *Br J Pharmacol* 155: 617–619, 2008.
86. **Leffler CW, Parfenova H, Basuroy S, Jagger JH, Umstot ES, Fedinec AL.** Hydrogen sulfide and cerebral microvascular tone in newborn pigs. *Am J Physiol Heart Circ Physiol* 300: H440–H447, 2011.
87. **Lefter CW, Parfenova H, Jagger JH, Wang R.** Carbon monoxide and hydrogen sulfide: gaseous messengers in cerebrovascular circulation. *J Appl Physiol* 100: 1065–1076, 2006.
88. **Li J, Li Y, Du Y, Mou K, Sun H, Zang Y, Liu C.** Endogenous hydrogen sulfide as a mediator of vas deferens smooth muscle relaxation. *Fertil Steril* 95: 1833–1835, 2010.
89. **Li L, Hsu A, Moore PK.** Actions and interactions of nitric oxide, carbon monoxide and hydrogen sulphide in the cardiovascular system and in inflammation—a tale of three gases! *Pharmacol Ther* 123: 386–400, 2009.
90. **Li L, Rose P, Moore PK.** Hydrogen sulfide and cell signaling. *Annu Rev Pharmacol Toxicol* 51: 169–187, 2011.
91. **Li L, Rossoni G, Sparatore A, Lee LC, Del SP, Moore PK.** Anti-inflammatory and gastrointestinal effects of a novel diclofenac derivative. *Free Radic Biol Med* 42: 706–719, 2007.
92. **Li L, Whiteman M, Guan YY, Neo KL, Cheng y Lee SW, Zhao Y, Baskar R, Tan CH, Moore PK.** Characterization of a novel, water-soluble hydrogen sulfide-releasing molecule (GYY4137): new insights into the biology of hydrogen sulfide. *Circulation* 117: 2351–2360, 2008.
93. **Li T, Zhao B, Wang C, Wang H, Liu Z, Li W, Jin H, Tang C, Du J.** Regulatory effects of hydrogen sulfide on IL-6, IL-8 and IL-10 levels in the plasma and pulmonary tissue of rats with acute lung injury. *Exp Biol Med (Maywood)* 233: 1081–1087, 2008.
94. **Li YF, Xiao CS, Hui RT.** Calcium sulfide (CaS), a donor of hydrogen sulfide (H<sub>2</sub>S): a new antihypertensive drug? *Med Hypotheses* 73: 445–447, 2009.
95. **Lim JJ, Liu YH, Khin ES, Bian JS.** Vasoconstrictive effect of hydrogen sulfide involves downregulation of cAMP in vascular smooth muscle cells. *Am J Physiol Cell Physiol* 295: C1261–C1270, 2008.
96. **Linden DR, Levitt MD, Farrugia G, Szurszewski JH.** Endogenous production of H<sub>2</sub>S in the gastrointestinal tract: still in search of a physiologic function. *Antioxid Redox Signal* 12: 1135–1146, 2010.
97. **Liu YH, Bian JS.** Bicarbonate-dependent effect of hydrogen sulfide on vascular contractility in rat aortic rings. *Am J Physiol Cell Physiol* 299: C866–C872, 2010.
98. **Lu M, Choo CH, Hu LF, Tan BH, Hu G, Bian JS.** Hydrogen sulfide regulates intracellular pH in rat primary cultured glia cells. *Neurosci Res* 66: 92–98, 2010.
99. **Lu M, Liu YH, Goh HS, Wang JJ, Yong QC, Wang R, Bian JS.** Hydrogen sulfide inhibits plasma renin activity. *J Am Soc Nephrol* 21: 993–1002, 2010.
100. **Maione S, Leyva J, Palazzo E, Stella L, Rossi F.** L-Cysteinesulfinic acid modulates cardiovascular function in the periaqueductal gray area of rat. *J Cardiovasc Pharmacol* 32: 650–653, 1998.
101. **Mancardi D, Penna C, Merlino A, Del SP, Wink DA, Pagliaro P.** Physiological and pharmacological features of the novel gasotransmitter: Hydrogen sulfide. *Biochim Biophys Acta* 1787: 864–872, 2009.

102. Mancuso C, Navarra P, Preziosi P. Roles of nitric oxide, carbon monoxide, and hydrogen sulfide in the regulation of the hypothalamic-pituitary-adrenal axis. *J Neurochem* 113: 563–575, 2010.
103. Maron BA, Loscalzo J. The treatment of hyperhomocysteinemia. *Annu Rev Med* 60: 39–54, 2009.
104. Martelli A, Testai L, Breschi MC, Blandizzi C, Virdis A, Taddei S, Calderone V. Hydrogen sulphide: novel opportunity for drug discovery. *Med Res Rev* In press.
105. Martin GR, McKnight GW, Dickey MS, Coffin CS, Ferraz JG, Wallace JL. Hydrogen sulphide synthesis in the rat and mouse gastrointestinal tract. *Dig Liver Dis* 42: 103–109, 2010.
106. Medani M, Collins D, Docherty NG, Baird AW, O'Connell PR, Winter DC. Emerging role of hydrogen sulfide in colonic physiology and pathophysiology. *Inflamm Bowel Dis*. In press.
107. Morii D, Miyagatani Y, Nakamae N, Murao M, Taniyama K. Japanese experience of hydrogen sulfide: the suicide craze in 2008. *J Occup Med Toxicol* 5: 28, 2010.
108. Mukherjee S, Lekli I, Ray D, Gangopadhyay H, Raychaudhuri U, Das DK. Comparison of the protective effects of steamed and cooked broccolis on ischaemia-reperfusion-induced cardiac injury. *Br J Nutr* 103: 815–823, 2010.
109. Mustafa AK, Gadalla MM, Sen N, Kim S, Mu W, Gazi SK, Barrow RK, Yang G, Wang R, Snyder SH. H<sub>2</sub>S signals through protein S-sulphydration. *Sci Signal* 2: ra72, 2009.
110. Mustafa AK, Gadalla MM, Snyder SH. Signaling by gasotransmitters. *Sci Signal* 2: re2, 2009.
111. Muzaffar S, Jeremy JY, Sparatore A, Del SP, Angelini GD, Shukla N. H<sub>2</sub>S-donating sildenafil (ACS6) inhibits superoxide formation and gp91phox expression in arterial endothelial cells: role of protein kinases A and G. *Br J Pharmacol* 155: 984–994, 2008.
112. Muzaffar S, Shukla N, Bond M, Newby AC, Angelini GD, Sparatore A, Del SP, Jeremy JY. Exogenous hydrogen sulfide inhibits superoxide formation, NOX-1 expression and Rac1 activity in human vascular smooth muscle cells. *J Vasc Res* 45: 521–528, 2008.
113. Nagao M, Linden DR, Duenes JA, Sarr MG. Mechanisms of action of the gasotransmitter hydrogen sulfide in modulating contractile activity of longitudinal muscle of rat ileum. *J Gastrointest Surg* 15: 12–22, 2011.
114. Nicholson CK, Calvert JW. Hydrogen sulfide and ischemia-reperfusion injury. *Pharmacol Res* 62: 289–297, 2010.
115. Olson KR. Vascular actions of hydrogen sulfide in non-mammalian vertebrates. *Antioxid Redox Signal* 7: 804–812, 2005.
116. Olson KR. Is hydrogen sulfide a circulating “gasotransmitter” in vertebrate blood? *Biochim Biophys Acta* 1787: 856–863, 2009.
117. Olson KR, Dombkowski RA, Russell MJ, Doellman MM, Head SK, Whitfield NL, Madden JA. Hydrogen sulfide as an oxygen sensor/transducer in vertebrate hypoxic vasoconstriction and hypoxic vasodilation. *J Exp Biol* 209: 4011–4023, 2006.
118. Olson KR, Whitfield NL. Hydrogen sulfide and oxygen sensing in the cardiovascular system. *Antioxid Redox Signal* 12: 1219–1234, 2010.
119. Olson KR, Whitfield NL, Bearden SE, St LJ, Nilson E, Gao Y, Madden JA. Hypoxic pulmonary vasodilation: A paradigm shift with a hydrogen sulfide mechanism. *Am J Physiol Regul Integr Comp Physiol* 298: R51–R60, 2010.
120. Otulakowski G, Kavanagh BP. Hydrogen sulfide in lung injury: therapeutic hope from a toxic gas? *Anesthesiology* 113: 4–6, 2010.
121. Pae HO, Lee YC, Jo EK, Chung HT. Subtle interplay of endogenous bioactive gases (NO, CO, and H<sub>2</sub>S) in inflammation. *Arch Pharm Res* 32: 1155–1162, 2009.
122. Parajuli SP, Choi S, Lee J, Kim YD, Park CG, Kim MY, Kim HI, Yeum CH, Jun JY. The inhibitory effects of hydrogen sulfide on pacemaker activity of interstitial cells of Cajal from mouse small intestine. *Korean J Physiol Pharmacol* 14: 83–89, 2010.
123. Patel P, Vathish M, Heptinstall J, Wang R, Carson RJ. The endogenous production of hydrogen sulphide in intrauterine tissues [Online]. *Reprod Biol Endocrinol* 7: 10, 2009.
124. Peng YJ, Nanduri J, Raghuraman G, Souvannakitti D, Gadalla MM, Kumar GK, Snyder SH, Prabhakar NR. H<sub>2</sub>S mediates O<sub>2</sub> sensing in the carotid body. *Proc Natl Acad Sci USA* 107: 10719–10724, 2010.
125. Perna AF, Luciano MG, Ingrassio D, Pulzella P, Sepe I, Lanza D, Violetti E, Capasso R, Lombardi C, De Santo NG. Hydrogen sulphide-generating pathways in haemodialysis patients: a study on relevant metabolites and transcriptional regulation of genes encoding for key enzymes. *Nephrol Dial Transplant* 24: 3756–3763, 2009.
126. Perrino E, Cappelletti G, Tazzari V, Giavini E, Del SP, Sparatore A. New sulfated derivatives of valproic acid with enhanced histone deacetylase inhibitory activity. *Bioorg Med Chem Lett* 18: 1893–1897, 2008.
127. Pietri R, Roman-Morales E, Lopez-Garriga J. Hydrogen sulfide and hemeproteins: knowledge and mysteries. *Antioxid Redox Signal*. In press.
128. Predmore BL, Lefler DJ. Development of hydrogen sulfide-based therapeutics for cardiovascular disease. *J Cardiovasc Transl Res* 3: 487–498, 2010.
129. Qu K, Chen CP, Halliwell B, Moore PK, Wong PT. Hydrogen sulfide is a mediator of cerebral ischemic damage. *Stroke* 37: 889–893, 2006.
130. Qu K, Lee SW, Bian JS, Low CM, Wong PT. Hydrogen sulfide: Neurochemistry and neurobiology. *Neurochem Int* 52: 155–165, 2007.
131. Renga B, Mencarelli A, Migliorati M, Distrutti E, Fiorucci S. Bile-acid-activated farnesoid X receptor regulates hydrogen sulfide production and hepatic microcirculation. *World J Gastroenterol* 15: 2097–2108, 2009.
132. Rossoni G, Sparatore A, Tazzari V, Manfredi B, Del SP, Berti F. The hydrogen sulphide-releasing derivative of diclofenac protects against ischaemia-reperfusion injury in the isolated rabbit heart. *Br J Pharmacol* 153: 100–109, 2008.
133. Rowan FE, Docherty NG, Coffey JC, O'Connell PR. Sulphate-reducing bacteria and hydrogen sulphide in the aetiology of ulcerative colitis. *Br J Surg* 96: 151–158, 2009.
134. Schenk S, Kesselmeier J, Anders E. How does the exchange of one oxygen atom with sulfur affect the catalytic cycle of carbonic anhydrase? *Chemistry* 10: 3091–3105, 2004.
135. Sehnert SS, Jiang L, Burdick JF, Risby TH. Breath biomarkers for detection of human liver diseases: preliminary study. *Biomarkers* 7: 174–187, 2002.
136. Sen N, Snyder SH. Protein modifications involved in neurotransmitter and gasotransmitter signaling. *Trends Neurosci* 2010.
137. Sen U, Mishra PK, Tyagi N, Tyagi SC. Homocysteine to hydrogen sulfide or hypertension. *Cell Biochem Biophys* 57: 49–58, 2010.
138. Sen U, Munjal C, Qipshidze N, Abe O, Gargoum R, Tyagi SC. Hydrogen sulfide regulates homocysteine-mediated glomerulosclerosis. *Am J Nephrol* 31: 442–455, 2010.
139. Shan Y, Zhao R, Geng W, Lin N, Wang X, Du X, Wang S. Protective effect of sulforaphane on human vascular endothelial cells against lipopolysaccharide-induced inflammatory damage. *Cardiovasc Toxicol* 10: 139–145, 2010.
140. Shi L, Du J, Qi J, Wei B, Tang C, Tang X. Effects of high pulmonary blood flow on pulmonary vasculature structure and the gene expression of cystathionine-gamma-lyase. *Beijing Da Xue Xue Bao* 35: 566–570, 2003.
141. Shibuya N, Tanaka M, Yoshida M, Ogasawara Y, Togawa T, Ishii K, Kimura H. 3-Mercaptopyruvate sulfurtransferase produces hydrogen sulfide and bound sulfane sulfur in the brain. *Antioxid Redox Signal* 11: 703–714, 2009.
142. Shukla N, Rossoni G, Hotston M, Sparatore A, Del SP, Tazzari V, Persad R, Angelini GD, Jeremy JY. Effect of hydrogen sulphide-donating sildenafil (ACS6) on erectile function and oxidative stress in rabbit isolated corpus cavernosum and in hypertensive rats. *BJU Int* 103: 1522–1529, 2009.
143. Singh S, Padovani D, Leslie RA, Chiku T, Banerjee R. Relative contributions of cystathionine beta-synthase and gamma-cystathionase to H<sub>2</sub>S biogenesis via alternative trans-sulfuration reactions. *J Biol Chem* 284: 22457–22466, 2009.
144. Smith HS. Hydrogen sulfide's involvement in modulating nociception. *Pain Physician* 12: 901–910, 2009.
145. Sparatore A, Perrino E, Tazzari V, Giustarini D, Rossi R, Rossoni G, Erdman K, Schroder H, Del SP. Pharmacological profile of a novel H<sub>2</sub>S-releasing aspirin. *Free Radic Biol Med* 46: 586–592, 2009.
146. Srilatha B, Adaikan PG, Moore PK. Possible role for the novel gasotransmitter hydrogen sulphide in erectile dysfunction—a pilot study. *Eur J Pharmacol* 535: 280–282, 2006.
147. Srilatha B, Hu L, Adaikan GP, Moore PK. Initial characterization of hydrogen sulfide effects in female sexual function. *J Sex Med* 6: 1875–1884, 2009.
148. Stipanuk MH. Sulfur amino acid metabolism: pathways for production and removal of homocysteine and cysteine. *Annu Rev Nutr* 24: 539–577, 2004.

149. **Stipanuk MH, Ueki I.** Dealing with methionine/homocysteine sulfur: cysteine metabolism to taurine and inorganic sulfur. *J Inherit Metab Dis* 34: 17–32, 2011.
150. **Studer SM, Orens JB, Rosas I, Krishnan JA, Cope KA, Yang S, Conte JV, Becker PB, Risby TH.** Patterns and significance of exhaled-breath biomarkers in lung transplant recipients with acute allograft rejection. *J Heart Lung Transplant* 20: 1158–1166, 2001.
151. **Sugiura Y, Kashiba M, Maruyama K, Hoshikawa K, Sasaki R, Saito K, Kimura H, Goda N, Suematsu M.** Cadmium exposure alters metabolomics of sulfur-containing amino acids in rat testes. *Antioxid Redox Signal* 7: 781–787, 2005.
152. **Svoronos PDN, Bruno TJ.** Carbonyl sulfide: a review of its chemistry and properties. *Ind Eng Chem Res* 41: 5321–5336, 2002.
153. **Szabó C.** Hydrogen sulphide and its therapeutic potential. *Nat Rev Drug Discov* 6: 917–935, 2007.
154. **Szabo C.** Gaseotransmitters: new frontiers for translational science. *Sci Transl Med* 2: 59ps54, 2010.
155. **Szabo C, Papapetropoulos A.** Hydrogen sulfide and angiogenesis: mechanisms and applications. *Br J Pharmacol*. In press.
156. **Szabo G, Veres G, Radovits T, Gero D, Modis K, Miesel-Groschel C, Horkay F, Karck M, Szabo C.** Cardioprotective effects of hydrogen sulfide. *Nitric Oxide*. In press.
157. **Tamizhselvi R, Koh YH, Sun J, Zhang H, Bhatia M.** Hydrogen sulfide induces ICAM-1 expression and neutrophil adhesion to caerulein-treated pancreatic acinar cells through NF-kappaB and Src-family kinases pathway. *Exp Cell Res* 316: 1625–1636, 2010.
158. **Tamizhselvi R, Moore PK, Bhatia M.** Hydrogen sulfide acts as a mediator of inflammation in acute pancreatitis: in vitro studies using isolated mouse pancreatic acinar cells. *J Cell Mol Med* 11: 315–326, 2007.
159. **Tamizhselvi R, Moore PK, Bhatia M.** Inhibition of hydrogen sulfide synthesis attenuates chemokine production and protects mice against acute pancreatitis and associated lung injury. *Pancreas* 36: e24–e31, 2008.
160. **Tan BH, Wong PT, Bian JS.** Hydrogen sulfide: a novel signaling molecule in the central nervous system. *Neurochem Int* 56: 3–10, 2010.
161. **Tang G, Wu L, Wang R.** The effect of hydroxylamine on KATP channels in vascular smooth muscle and underlying mechanisms. *Mol Pharmacol* 67: 1723–1731, 2005.
162. **Tang XQ, Shen XT, Huang YE, Ren YK, Chen RQ, Hu B, He JQ, Yin WL, Xu JH, Jiang ZS.** Hydrogen sulfide antagonizes homocysteine-induced neurotoxicity in PC12 cells. *Neurosci Res* 68: 241–249, 2010.
163. **Tangerman A.** Measurement and biological significance of the volatile sulfur compounds hydrogen sulfide, methanethiol, and dimethyl sulfide in various biological matrices. *J Chromatogr B Analyt Technol Biomed Life Sci* 2009.
164. **Taniguchi S, Kang L, Kimura T, Niki I.** Hydrogen sulphide protects mouse pancreatic beta-cells from cell death induced by oxidative stress, but not by endoplasmic reticulum stress. *Br J Pharmacol* 162: 1171–1178, 2010.
165. **Tay AS, Hu LF, Lu M, Wong PT, Bian JS.** Hydrogen sulfide protects neurons against hypoxic injury via stimulation of ATP-sensitive potassium channel/protein kinase C/extracellular signal-regulated kinase/heat shock protein 90 pathway. *Neuroscience* 167: 277–286, 2010.
166. **Telezkin V, Brazier SP, Cayzac S, Muller CT, Riccardi D, Kemp PJ.** Hydrogen sulfide inhibits human BK(Ca) channels. *Adv Exp Med Biol* 648: 65–72, 2009.
167. **Telezkin V, Brazier SP, Cayzac SH, Wilkinson WJ, Riccardi D, Kemp PJ.** Mechanism of inhibition by hydrogen sulfide of native and recombinant BKCa channels. *Respir Physiol Neurobiol* 172: 169–178, 2010.
168. **Tiranti V, Viscomi C, Hildebrandt T, Di M, I, Miner R, Tiveron C, Levitt MD, Prella A, Fagiolarini G, Rimoldi M, Zeviani M.** Loss of ETHE1, a mitochondrial dioxygenase, causes fatal sulfide toxicity in ethylmalonic encephalopathy. *Nat Med* 15: 200–205, 2009.
169. **Toombs CF, Insko MA, Wintner EA, Deckwerth TL, Usansky H, Jamil K, Goldstein B, Cooreman M, Szabo C.** Detection of exhaled hydrogen sulphide gas in healthy human volunteers during intravenous administration of sodium sulphide. *Br J Clin Pharmacol* 69: 626–636, 2010.
170. **Trevisani M, Patacchini R, Nicoletti P, Gatti R, Gazzieri D, Lissi N, Zagli G, Creminon C, Geppetti P, Harrison S.** Hydrogen sulfide causes vanilloid receptor 1-mediated neurogenic inflammation in the airways. *Br J Pharmacol* 145: 1123–1131, 2005.
171. **Tripatara P, Patel NS, Brancalone V, Renshaw D, Rocha J, Sepodes B, Mota-Filipe H, Perretti M, Thiemermann C.** Characterisation of cystathionine gamma-lyase/hydrogen sulphide pathway in ischaemia/reperfusion injury of the mouse kidney: an in vivo study. *Eur J Pharmacol* 606: 205–209, 2009.
172. **Wagner CA.** Hydrogen sulfide: a new gaseous signal molecule and blood pressure regulator. *J Nephrol* 22: 173–176, 2009.
173. **Wagner F, Asfar P, Calzia E, Radermacher P, Szabo C.** Bench-to bedside review: Hydrogen sulfide—the third gaseous transmitter: applications for critical care. *Crit Care* 13: 213, 2009.
174. **Wallace JL.** Hydrogen sulfide-releasing anti-inflammatory drugs. *Trends Pharmacol Sci* 28: 501–505, 2007.
175. **Wallace JL.** Physiological and pathophysiological roles of hydrogen sulfide in the gastrointestinal tract. *Antioxid Redox Signal* 12: 1125–1133, 2010.
176. **Wallace JL, Dickey M, McKnight W, Martin GR.** Hydrogen sulfide enhances ulcer healing in rats. *FASEB J* 21: 4070–4076, 2007.
177. **Wallace JL, Wong L, McKnight W, Dickey M, Martin GR.** Endogenous and exogenous hydrogen sulfide promotes resolution of colitis in rats. *Gastroenterology* 137: 569–578, 2009.
178. **Wang Q, Wang XL, Liu HR, Rose P, Zhu YZ.** Protective effects of cysteine analogues on acute myocardial ischemia: novel modulators of endogenous H<sub>2</sub>S production. *Antioxid Redox Signal* 12: 1155–1165, 2010.
179. **Wang R.** Two's company, three's a crowd: can H<sub>2</sub>S be the third endogenous gaseous transmitter? *FASEB J* 16: 1792–1798, 2002.
180. **Wang R.** Hydrogen sulfide: a new EDRF. *Kidney Int* 76: 700–704, 2009.
181. **Webb GD, Lim LH, Oh VM, Yeo SB, Cheong YP, Ali MY, El OR, Lee CN, Wong PS, Caleb MG, Salto-Tellez M, Bhatia M, Chan ES, Taylor EA, Moore PK.** Contractile and vasorelaxant effects of hydrogen sulfide and its biosynthesis in the human internal mammary artery. *J Pharmacol Exp Ther* 324: 876–882, 2008.
182. **Wei HL, Zhang CY, Jin HF, Tang CS, Du JB.** Hydrogen sulfide regulates lung tissue-oxidized glutathione and total antioxidant capacity in hypoxic pulmonary hypertensive rats. *Acta Pharmacol Sin* 29: 670–679, 2008.
183. **Whiteman M, Cheung NS, Zhu YZ, Chu SH, Siau JL, Wong BS, Armstrong JS, Moore PK.** Hydrogen sulphide: a novel inhibitor of hypochlorous acid-mediated oxidative damage in the brain? *Biochem Biophys Res Commun* 326: 794–798, 2005.
184. **Whiteman M, Li L, Kostetski I, Chu SH, Siau JL, Bhatia M, Moore PK.** Evidence for the formation of a novel nitrosothiol from the gaseous mediators nitric oxide and hydrogen sulphide. *Biochem Biophys Res Commun* 343: 303–310, 2006.
185. **Whiteman M, Moore PK.** Hydrogen sulfide and the vasculature: a novel vasculoprotective entity and regulator of nitric oxide bioavailability? *J Cell Mol Med* 13: 488–507, 2009.
186. **Whitfield NL, Kreimler EL, Verdial FC, Skovgaard N, Olson KR.** A Reappraisal of H<sub>2</sub>S/sulfide concentration in vertebrate blood and its potential significance in ischemic preconditioning and vascular signaling. *Am J Physiol Regul Integr Comp Physiol* 294: R1930–R1937, 2008.
187. **Williams KT, Schalinke KL.** Homocysteine metabolism and its relation to health and disease. *Biofactors* 36: 19–24, 2010.
188. **Wintner EA, Deckwerth TL, Langston W, Bengtsson A, Leviten D, Hill P, Insko MA, Dumpit R, VandenEkart E, Toombs CF, Szabo C.** A monobromobimane-based assay to measure the pharmacokinetic profile of reactive sulphide species in blood. *Br J Pharmacol* 160: 941–957, 2010.
189. **Wojcicka G, Jamroz-Wisniewska A, Atanasova P, Chaldakov GN, Chylinska-Kula B, Beltowski J.** Differential effects of statins on endogenous H<sub>2</sub>S formation in perivascular adipose tissue. *Pharmacol Res* 63: 68–76, 2011.
190. **Woodall GM, Smith RL, Granville GC.** Proceedings of the Hydrogen Sulfide Health Research and Risk Assessment Symposium October 31–November 2, 2000. *Inhal Toxicol* 17: 593–639, 2005.
191. **Xia M, Chen L, Muh RW, Li PL, Li N.** Production and actions of hydrogen sulfide, a novel gaseous bioactive substance, in the kidneys. *J Pharmacol Exp Ther* 329: 1056–1062, 2009.
192. **Xu Z, Prathapasinghe G, Wu N, Hwang SY, Siow YL, OK.** Ischemia-reperfusion reduces cystathionine-β-synthase-mediated hydrogen sulfide generation in the kidney. *Am J Physiol Renal Physiol* 297: F27–F35, 2009.

193. **Ye L, Dinkova-Kostova AT, Wade KL, Zhang Y, Shapiro TA, Talalay P.** Quantitative determination of dithiocarbamates in human plasma, serum, erythrocytes and urine: pharmacokinetics of broccoli sprout isothiocyanates in humans. *Clin Chim Acta* 316: 43–53, 2002.
194. **Yong QC, Cheong JL, Hua F, Deng LW, Khoo YM, Lee HS, Perry A, Wood M, Whiteman M, Bian JS.** Regulation of heart function by endogenous gaseous mediators. Cross-talk between nitric oxide and hydrogen sulfide. *Antioxid Redox Signal*. In press.
195. **Yong QC, Hu LF, Wang S, Huang D, Bian JS.** Hydrogen sulfide interacts with nitric oxide in the heart: possible involvement of nitroxyl. *Cardiovasc Res* 88: 482–491, 2010.
196. **Yu YP, Li ZG, Wang DZ, Zhan X, Shao JH.** Hydrogen sulfide as an effective and specific novel therapy for acute carbon monoxide poisoning. *Biochem Biophys Res Commun* 404: 6–9, 2011.
197. **Yuan P, Xue H, Zhou L, Qu L, Li C, Wang Z, Ni J, Yu C, Yao T, Huang Y, Wang R, Lu L.** Rescue of mesangial cells from high glucose-induced over-proliferation and extracellular matrix secretion by hydrogen sulfide. *Nephrol Dial Transplant*. In press.
198. **Yusof M, Kamada K, Kalogeris T, Gaskin FS, Korthuis RJ.** Hydrogen sulfide triggers late-phase preconditioning in posts ischemic small intestine by an NO- and p38 MAPK-dependent mechanism. *Am J Physiol Heart Circ Physiol* 296: H868–H876, 2009.
199. **Zhang C, Du J, Bu D, Yan H, Tang X, Tang C.** The regulatory effect of hydrogen sulfide on hypoxic pulmonary hypertension in rats. *Biochem Biophys Res Commun* 302: 810–816, 2003.
200. **Zhang H, Bhatia M.** Hydrogen sulfide: a novel mediator of leukocyte activation. *Immunopharmacol Immunotoxicol* 30: 631–645, 2008.
201. **Zhang H, Gao Y, Zhao F, Dai Z, Meng T, Tu S, Yan Y.** Hydrogen sulfide reduces mRNA and protein levels of beta-site amyloid precursor protein cleaving enzyme 1 in PC12 cells. *Neurochem Int* 58: 169–175, 2011.
202. **Zhang Q, Du J, Zhou W, Yan H, Tang C, Zhang C.** Impact of hydrogen sulfide on carbon monoxide/heme oxygenase pathway in the pathogenesis of hypoxic pulmonary hypertension. *Biochem Biophys Res Commun* 317: 30–37, 2004.
203. **Zhao W, Wang R.** H<sub>2</sub>S-induced vasorelaxation and underlying cellular and molecular mechanisms. *Am J Physiol Heart Circ Physiol* 283: H474–H480, 2002.
204. **Zhao W, Zhang J, Lu Y, Wang R.** The vasorelaxant effect of H<sub>2</sub>S as a novel endogenous KATP channel opener. *EMBO J* 20: 6008–6016, 2001.
205. **Zhao Y, Wang H, Xian M.** Cysteine-activated hydrogen sulfide (H<sub>2</sub>S) donors. *J Am Chem Soc* 133: 15–17, 2011.
206. **Zhong G, Chen F, Cheng y Tang C, Du J.** The role of hydrogen sulfide generation in the pathogenesis of hypertension in rats induced by inhibition of nitric oxide synthase. *J Hypertens* 21: 1897–1885, 2003.
207. **Zhong GZ.** Hydrogen Sulfide-a potent multichannel anti-arrhythmic drug. *J Cardiovasc Dis Res* 1: 37–39, 2010.
208. **Zhu H, Jia Z, Strobl JS, Ehrlich M, Misra HP, Li Y.** Potent induction of total cellular and mitochondrial antioxidants and phase 2 enzymes by cruciferous sulforaphane in rat aortic smooth muscle cells: cytoprotection against oxidative and electrophilic stress. *Cardiovasc Toxicol* 8: 115–125, 2008.

