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Third ventricular coinjection of subthreshold doses of NPY and AgRP stimulate food hoarding and intake and neural activation

Brett J. W. Teubner, Erin Keen-Rhinehart, and Timothy J. Bartness

Department of Biology and Obesity Reversal Center, Georgia State University, Atlanta, Georgia

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Teubner BJ, Keen-Rhinehart E, Bartness TJ. Third ventricular coinjection of subthreshold doses of NPY and AgRP stimulate food hoarding and intake and neural activation. Am J Physiol Regul Integr Comp Physiol 302: R37–R48, 2012. First published October 19, 2011; doi:10.1152/ajpregu.00475.2011.—We previously demonstrated that 3rd ventricular (3V) neuropeptide Y (NPY) or agouti-related protein (AgRP) injection potently stimulates food foraging/hoarding/intake in Siberian hamsters. Because NPY and AgRP are highly colocalized in arcuate nucleus neurons in this and other species, we tested whether subthreshold doses of NPY and AgRP coinjected into the 3V stimulates food foraging, hoarding, and intake, and/or neural activation [c-Fos immunoreactivity (c-Fos-ir)] in hamsters housed in a foraging/hoarding apparatus. In the behavioral experiment, each hamster received four 3V treatments by using subthreshold doses of NPY and AgRP coinjected for all behaviors: 1) NPY, 2) AgRP, 3) NPY+AgRP, and 4) saline with a 7-day washout period between treatments. Food foraging, intake, and hoarding were measured at 1, 2, 4, and 24 h and 2 and 3 days postinjection. Only when NPY and AgRP were coinjected was food intake and hoarding increased. After identical treatment in separate animals, c-Fos-ir was assessed at 90 min and 14 h postinjection, times when food intake (0–1 h) and hoarding (4–24 h) were uniquely stimulated. c-Fos-ir was increased in several hypothalamic nuclei previously shown to be involved in ingestive behaviors and the central nucleus of the amygdala (CeA), but only in NPY+AgRP-treated animals (90 min and 14 h: magnocellular and parvocellular regions of the hypothalamic paraventricular nucleus and perifornical area; 14 h: CeA and sub zona incerta). These results suggest that NPY and AgRP interact to stimulate food hoarding and intake at distinct times, perhaps released as a cocktail naturally with food deprivation to stimulate these behaviors.

c-fos, Siberian hamster; hypothalamic paraventricular nucleus; perifornical area; central nucleus of the amygdala; sub zona incerta

IN 1918, WALLACE CRAIG (12) broadly dichotomized behaviors into responses that bring animals in contact with a goal object (appetitive behaviors), here food, that include foraging and hoarding or the culmination of that acquisition (consummatory behaviors), here eating. Most animals forage and hoard food, including humans (for review see Ref. 50), who, in industrialized nations, seek food at stores and store food in refrigerators, freezers, pantries, and cupboards (hoard). Food hoarding may have evolved as one way to help assure energy availability under conditions where the presence of food is variable (e.g., seasonal changes, increased predation pressures) in addition to internal energy stored as body fat.

Although much has been discovered about the mechanisms underlying consummatory ingestive behaviors from studies of laboratory rats and mice, most studies typically are not designed to measure the appetitive phase of ingestive behavior (for review see Ref. 7). Therefore, we know comparatively little about the mechanisms underlying appetitive ingestive behaviors in rodents compared with consummatory ingestive behavior. Likely contributing to the dearth of information on appetitive behavior mechanisms is the lack of food hoarding by the wild counterparts to laboratory rats and mice (for review see Refs. 7 and 33); those animals eat food at the source or carry some in their mouth to a nearby safe locale, where they eat it, but not to their burrows. Thus, laboratory rats and mice perhaps are best suited for studies of consummatory ingestive behaviors and not appetitive ingestive behaviors, at least in terms of food hoarding.

By contrast with laboratory rats and mice, Siberian hamsters (Phodopus sungorus) naturally hoard food in nature (53) and in the laboratory (for review see Refs. 5 and 7). Siberian hamsters have large cheek pouches that allow them to transport considerable amounts of food from its source to their burrows, somewhat analogous to humans carrying food using grocery bags and car trunks; thus, it is not surprising that food hoarding is an important component of their ingestive behavior repertoire (for review see Ref. 7). Therefore, combining the study of Siberian hamsters with our foraging/hoarding apparatus (see Ref. 15 for complete description and one in brief below), we are in a unique position to more inclusively study both appetitive and consummatory ingestive behaviors. The most reliable stimulator of foraging and/or food hoarding across species is food deprivation (for a review see Ref. 7). In Siberian hamsters, food deprivation-refeeding results in prolonged increases in food foraging and hoarding, but not food intake, a disassociation similar to humans (21, 27, 38, 39), thereby permitting the unraveling of the mechanisms underlying appetitive and consummatory ingestive behaviors that often are intricately intertwined (for review see Refs. 7 and 33). We have successfully tested the role of several endogenous signals thought to be involved in food deprivation-induced increases in food intake seen by laboratory rats and mice by using Siberian hamsters housed in our foraging/hoarding apparatus. These studies resulted in marked increases in food hoarding with much lesser increases in food intake (i.e., ghrelin
(30–32), neuropeptide Y (NPY; 13, 18, 32) and agouti-related protein (AgRP; 17).

The arcuate nucleus (ARC) has received considerable attention as a major component of the distributed system that controls food intake. One neurochemical phenotype of these neurons is a high colocalization of NPY and AgRP (25), a colocalization that also occurs in Siberian hamsters (40). In Siberian hamsters, as in laboratory rats and mice [e.g., (47)], food deprivation increases NPY and AgRP gene expression (45). Potential release of both orexigenic peptides in vivo is suggested by the co-release of NPY and AgRP in hypothalamic explants after exposure to cocaine and amphetamine-related transcript (20), as well the presence of neurons within the hypothalamic paraventricular nucleus (PVH) that receive ARC neural projections consisting of fibers coexpressing NPY- and AgRP-immunoreactivity (11). In addition, electrophysiological measures of individual PVH neurons demonstrate a common site of action for NPY and AgRP (11), the latter primarily or exclusively being an antagonist of the melanocortin 3/4 receptor [MC 3/4R; (10)]. Collectively, these data point to a possible co-release and interaction of NPY and AgRP under conditions such as food deprivation.

Both AgRP and NPY appear to be involved with appetitive ingestive behaviors as well as their well-established effects on consummatory ingestive behavior. That is, central [3rd ventricular (3V)] injection of NPY (18) or AgRP (17) and parenchymal injection of NPY into the PVH or perifornical area (pFA; 13) increase foraging and especially food hoarding by Siberian hamsters along with smaller increases in food intake, with the hoarding levels similar in magnitude to that occurring with food deprivation-refeeding in this species [e.g., (4, 14, 16)]. Interestingly, NPY and AgRP differ in their temporal pattern of increasing food hoarding with NPY, causing early increases (1–2 day, especially the first 24 h) and AgRP causing a more prolonged increase [3–7 days; (13, 17, 18)].

We are aware of only one study in laboratory rodents where a possible interaction of NPY and AgRP was tested. Specifically, PVH coinjection of a suprathreshold dose of NPY with a subthreshold dose of AgRP increased food intake above that of AgRP alone (54). Thus, given the colocalization and possible co-release of NPY and AgRP (20), both of their stimulatory effects on ingestive behavior, and apparent interaction in the PVH to stimulate food intake (54), it is possible that the two interact to stimulate appetitive (food foraging, hoarding) as well as consummatory (food intake) ingestive behaviors. Therefore, we conducted two experiments to address this hypothesis. In experiment 1 we asked: Do subthreshold doses of NPY and AgRP interact to increase ingestive behavior? In a second supporting functional neuroanatomical experiment to the first, we asked: Do subthreshold doses of NPY and AgRP interact to activate central neuronal activity? In terms of the latter study, we assessed c-Fos immunoreactivity (c-Fos-ir), an accepted indicator of neural activation (28).

**MATERIAL AND METHODS**

**Animals**

Adult male Siberian hamsters, ~2.5–3.5 mo old and weighing 35–45 g were obtained from our breeding colony. Hamsters were group housed and raised in a long-day photoperiod (16:8 h light-dark cycle; light onset at 0200) from birth until use in the experiments.

Food (5001; Purina, St. Louis, MO) and tap water were available ad libitum unless otherwise indicated. Room temperature was maintained at 21 ± 2.0°C. All procedures were approved by the Georgia State University Institutional Animal Care and Use Committee and are in accordance with Public Health Service and United States Department of Agriculture guidelines.

**Cannula Implantation, Injections, and Verification**

Cannulae were stereotaxically implanted into the 3V under isoflurane (Aerrane; Baxter Healthcare, Deerfield, IL) inhalation anesthesia as previously described (13). Briefly, each hamster had its head shaved and skull exposed. A guide cannula (26-gauge stainless steel; Plastics One, Roanoke, VA) was lowered stereotaxically [coordinates: level skull, anterior-posterior from bregma 0 mm, medial-lateral from midsagittal sinus 0 mm (the sinus was moved to avoid damage due to lowering of the cannula), and dorsal-ventral from the top of the skull ~5.0 mm] targeted for placement just above the 3V. The guide cannulae were secured to the skull using cyanoacrylate ester gel, 3/16 mm jeweler’s screws, and dental acrylic. A removable obturator sealed the opening in the guide cannulae throughout the experiment except during 3V injections. Hamsters received buprenorphine (0.2 mg/kg body mass) at 12 and 24 h postsurgery to minimize surgical discomfort and an apple slice to facilitate food and water intake. A 2-wk postsurgery recovery period occurred where the hamsters were housed in shoebox cages.

One week before each test day, each animal was mock-injected daily, where the obturator was removed, and the animal was lightly restrained for one minute to acclimate the animal to the intraventricular injection procedure. On test days, an inner cannula (33-gauge stainless steel; Plastics One) extending 0.5 mm beyond the guide cannula was connected to a Hamilton syringe via PE-20 tubing and inserted into the guide cannula. All hamsters were injected with 400 nl total volume of neurochemical and/or vehicle at light offset. Animals were lightly restrained by hand during the ~30-s injection, and the injection needle remained in place ~30 s before withdrawal, as we have done previously (13).

Following the last behavioral test in experiment 1, an injection of 400 nl of bromophenol blue dye was given to confirm placement of the cannula in the third ventricle. The animals were then given an overdose of pentobarbital sodium (100 mg/kg), transcardially perfused with 100 ml of heparinized saline followed by 125 ml of 4% paraformaldehyde in PBS, pH = 7.4. The brains were removed and then postfixed in 4% paraformaldehyde for a minimum of 2 days, followed by 30% sucrose for at least 2 days, replacing the sucrose solution after 24 h. Each brain was sliced manually (80 μm) for cannula verification. Cannulae were considered to be located in the 3V if the dye was visible in any part of this ventricle as revealed by light microscopy. A similar procedure was carried out for experiment 2, except no dye was injected. Site verification was confirmed by examining the cannula tract, and tissue was sliced at 35 μm and then stored in PBS at 5°C until tissue processing (see below). Only the data from animals with confirmed 3V cannulae placements were included in the analyses. No cannula was dislodged across either study.

**Foraging and Hoarding Apparatus**

Hamsters were acclimated for 1 wk, before and after cannula implantation, in specially designed foraging/hoarding apparatuses as previously described (15) that would serve as their housing for the duration of the experiment. More specifically, two cages were connected with polyvinylchloride tubing system (38.1 mm ID and ~1.52 m long) with corners and straight-aways for vertical and horizontal climbs. The top, or food cage, was 456 × 234 × 200 mm (length × width × height) and equipped with a water bottle, running wheel (524 mm circumference), and food pellet dispenser. The diet (75 mg pellets, Bio-Serve, Frenchtown, NJ) and tap water were available ad libitum, unless otherwise noted. The bottom, or burrow cage, was
Measurement of Foraging, Food Hoarding, and Food Intake

Foraging (pellets earned) was defined as the number of pellets delivered upon completion of the requisite 10 wheel revolutions. Food hoarding (pellets hoarded) was defined as the number of pellets found in the bottom burrow cage in addition to those removed from the cheek pouches. For the 10 wheel revolutions per pellet group, food intake (pellets eaten) was defined as pellets earned minus pellets left in the top cage (surplus pellets) and pellets hoarded. For the nonwheel running contingent food access groups, food intake (pellets eaten) was defined as pellets given (400 pellets) minus pellets left in the top cage and pellets hoarded. An electronic balance used to weigh the food pellets was set to parts measurement; thus one 75-mg food pellet = 1 and fractions of a pellet were computed by the scale.

Experiment 1: Do Subthreshold Doses of NPY and AgRP Interact to Increase Ingestive Behavior?

Performing pilot experiments, we were able to determine subthreshold doses of both NPY (0.0176 nmol) and AgRP (0.01 nmol) that did not induce any of the three behaviors above that of saline controls, with the lowest doses producing behavioral increases being NPY: 0.088 nmol and AgRP: 0.05 nmol; that is, we are using doses five times lower than the lowest effective dose that stimulated these behaviors for each peptide. After those determinations, 42 Siberian hamsters were trained in the foraging/hoarding apparatuses and were then separated into three foraging groups: 10 revolutions/pellet (10REV; access to food contingent upon running 10 wheel revolutions per pellet), free wheel [FW; food was available noncontingently (not earned), but the running wheel was active (locomotor activity control group)], or blocked wheel [BW; food was available noncontingently (not earned), but the running wheel was blocked (sedentary control group)]. The groups were balanced for percent body mass change and average hoard size. On test days, animals were injected at light-offset and then were perfused at 90 min or 14 h postinjection and processed for c-Fos-ir (sc-52; Santa Cruz Biotechnology, Santa Cruz, CA) using our previously described methods for immunohistochemistry (37). The 90-min time point postinjection was chosen based on the predominant increases in food intake seen during that interval with 3V injection of suprathreshold doses of NPY (13, 18, 32) or of AgRP (17), as well as experiment 1. The 14-h time point was chosen because it was the midpoint of the interval ending at 4 h (the latest postinjection interval where food hoarding was not increased) and the interval where food hoarding was first increased (4–24 h) based on the data from experiment 1 (see below). c-Fos-ir was then counted across each entire structure using light microscopy with the counter blind to the treatment regimen. Specific nuclei were selected a priori based upon the available literature where NPY or AgRP injection resulted in c-Fos-ir and any other nuclei that appeared to have increased c-Fos-ir during quantification. To reduce the probability of counting the same neuron twice, Abercrombie’s correction factor was used (1).

Statistical Analyses

In experiment 1, raw data for each individual animal were transformed into percent change from saline before statistical analyses according to the formula: [\((x - \text{saline})/\text{saline}\) * 100], where x is the value measured in response to peptide treatment (0.0176 nmol NPY, 0.01 nmol AgRP, or 0.0176 nmol NPY + 0.01 nmol AgRP) and saline is the variable response to saline treatment for each behavior and time point. For all measures of food intake, foraging and food hoarding in experiment 1, the data are graphed as the mean percent difference from saline treatment (100%) ± SE. No statistical comparisons are reported across time within a test day or among foraging treatments, as there were no significant differences after the analysis of the full data set using repeated-measures ANOVA. Therefore, analyses only were run within time points using two-way ANOVA. c-Fos-ir data from experiment 2 were analyzed using one-way ANOVA. All statistical analyses were run using NCSS (version 2007; Kaysville, UT). Exact probabilities and test values were omitted for simplicity and clarity of presentation. Differences were considered statistically significant if P < 0.05.

RESULTS

Experiment 1: Do Subthreshold Doses of NPY and AgRP Interact to Increase Ingestive Behavior?

Wheel running. Subthreshold doses of NPY or of AgRP given alone did not stimulate wheel running activity that was not coupled to foraging (i.e., FW group, Fig. 1, A and B, respectively), suggesting there was not nonspecific stimulation of locomotor activity by these low doses of neuropeptides.
when given alone. When the subthreshold doses of NPY and AgRP were coinjected into the 3V, FW group wheel revolutions increased significantly during the 0–1 h postinjection interval (700%, P < 0.05; Fig. 1C), however.

Foraging. 3V injection of subthreshold doses of either NPY (Fig. 1D) or of AgRP (Fig. 1E) did not significantly increase foraging. When the subthreshold doses of each were given together, however, foraging was significantly increased compared with saline control injections during the 0- to 1-h postinjection interval (700%, P < 0.05; Fig. 1F). Foraging condition alone did not affect food intake.

Food intake. Subthreshold doses of either NPY (Fig. 2A) or of AgRP (Fig. 2B) did not stimulate food intake in any treatment group during any time interval with the exception of a ~600% increase during the first hour in the BW group when given only AgRP (P < 0.05; Fig. 2B). 3V injection of both NPY and AgRP together significantly increased food intake during the first hour for all foraging groups (~400–900%) compared with saline (P < 0.05; Fig. 2C). Foraging condition alone did not affect food intake.

Food hoarding. 3V injection of either NPY (Fig. 3A) or of AgRP (Fig. 3B) did not significantly increase food hoarding in any group at any time. Subthreshold doses of NPY and AgRP given together, however, significantly increased food hoarding during the 4–24 h postinjection interval for all foraging groups (~500–700%) compared with saline (P < 0.05; Fig. 3C). In addition, NPY + AgRP significantly increased food hoarding in the 10REV group during day 2 (~700%, P < 0.05; Fig. 3C). Foraging condition alone did not affect food hoarding.

Experiment 2: Do Subthreshold Doses of NPY and AgRP Interact to Activate Central Neuronal Activity?

Administration of subthreshold NPY or of subthreshold AgRP alone did not lead to a significant increase in c-Fos-ir at either time period compared with their saline-injected counterparts (Fig. 4). Significant increases in c-Fos-ir occurred at 90 min (Fig. 4A) and 14 h (Fig. 4B) postinjection of NPY + AgRP compared with saline (Figs. 5 and 6, respectively). Specifically, after 90 min, c-Fos-ir was significantly increased in the PVH (magnocellular and parvocellular regions; Fig. 5, A–D) and the pFA of the lateral hypothalamus (Fig. 5, E–H) compared with the saline-treated group (P < 0.05). Other areas that have increased c-Fos-ir in response to food deprivation (42), AgRP (23, 57), or NPY (44), such as the ventromedial nucleus of the

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Fig. 1. Mean ± SE % difference from saline of wheel revolutions in response to 3rd ventricular (3V) injection of 0.0176 nmol neuropeptide Y (NPY; A), 0.01 nmol agouti-related protein (AgRP; B), or 0.0176 nmol NPY + 0.01 nmol AgRP (C) in Siberian hamsters with no foraging requirement and a freely moving wheel (Free Wheel Revolutions) and 0.0176 nmol NPY (D), 0.01 nmol AgRP (E), or 0.0176 nmol NPY + 0.01 nmol AgRP (F) in Siberian hamsters with a 10 wheel revolutions per pellet foraging requirement (10 Rev). * P < 0.05 vs. saline controls.
limited to the regions described above, but other regions did not have significant changes in c-Fos-ir.

DISCUSSION

NPY and AgRP are important orexigenic factors underlying ingestive behavior in a variety of conditions and species (for review see Ref. 6). Here we found that 3V coinjection of subthreshold doses of NPY and AgRP resulted in significant increases in ingestive behaviors as well as in neural activation in brain nuclei shown to be involved in the response to food

Fig. 2. Mean ± SE % difference in food intake in response to 3V injection of 0.0176 nmol NPY (A), 0.01 nmol AgRP (B), or 0.0176 nmol NPY + 0.01 nmol AgRP (C) with no foraging requirement and a stationary wheel (Blocked Wheel, white bars), hamsters with no foraging requirement and a freely moving wheel (Free Wheel, gray bars), and hamsters with a 10 wheel revolutions per pellet foraging requirement (10 Rev, black bars). *P < 0.05 within foraging condition compared with saline controls.

Fig. 3. Mean ± SE % difference in food hoarding in response to 3V injection of 0.0176 nmol NPY (A), 0.01 nmol AgRP (B), or 0.0176 nmol NPY + 0.01 nmol AgRP (C) with no foraging requirement and a stationary wheel (Blocked Wheel, white bars), hamsters with no foraging requirement and a freely moving wheel (Free Wheel, gray bars), and hamsters with a 10 wheel revolutions per pellet foraging requirement (10 Rev, black bars). *P < 0.05 within foraging condition compared with saline controls.
The PVH (magno- and parvocellular regions) and pFA had increased c-Fos-ir at both threshold doses of NPY and AgRP. The PVH (paraventricular nucleus of the hypothalamus-magnocellular region; PVH-M, paraventricular nucleus of the hypothalamus-parvocellular region; VMH, ventromedial hypothalamus; ARC, arcuate nucleus; sZI, sub-zona incerta; pFA, perifornical area; CeA, central nucleus of the amygdale; AP, area postrema; NTS, nucleus of the tractus solitaries). *P < 0.05 compared with saline controls.

To our knowledge, only two other studies have examined the interaction of subthreshold doses of orexigenic factors on increasing food intake in rodents (46, 54). In the first, a subthreshold dose of AgRP was combined with a suprathreshold dose of NPY injected together in the PVH, resulting in increased food intake above AgRP alone, suggesting an interaction (54). NPY and AgRP have been coinjected in sheep at suprathreshold doses; however, there was no interaction such that food intake is not different than when the same dose of each is applied alone (51), possibly due to a ceiling effect on food intake or perhaps to species differences. In the second study of subthreshold doses of orexigenic factors interacting to affect food intake, subthreshold doses of NPY increased food intake when combined with either subthreshold orexin A or subthreshold melanin concentrating hormone (46). By stark
Fig. 5. Representative photographs of c-Fos-ir in response to 3V injection of saline, 0.0176 nmol NPY, 0.01 nmol AgRP, or 0.0176 nmol NPY + 0.01 nmol AgRP in the hypothalamic paraventricular nucleus after 90 min (A, a–d; ×10 magnification), the pFA after 90 min (B, e–h; ×10 magnification with box delineating the region).
Fig. 6. Representative photographs of c-Fos-ir in response to 3V injection of saline, 0.0176 nmol NPY, 0.01 nmol AgRP, or 0.0176 nmol NPY + 0.01 nmol AgRP in the sZI after 14 h (A, a–d; ×10 magnification with box delineating the region), and the central nucleus of the amygdala after 14 h (B, e–h; ×10 magnification).
In our second experiment, we expanded our behavioral data and tested for neural activation, as assayed by c-Fos-ir, in response to single acute 3V injection of subthreshold doses of either NPY or AgRP as well as their combination. We recognize the caveats of c-Fos-ir as an indicator of neural activation (28) in that the absence of proof is not the proof of absence given that some neurons use other immediate-early genes and that neuronal inhibition does not trigger neuronal expression of c-Fos. Therefore, we interpret the present findings knowing they are suggestive, at best, but nevertheless a commonly used, accepted, and starting point for determining brain sites affected by a given manipulation.

Food deprivation and ghrelin both increase c-Fos-ir in laboratory rats and mice, as might be expected given their role in triggering various ingestive behaviors. Food deprivation causes increases in c-Fos-ir indicating neural activation [e.g., (41, 42, 48)], with the nuclei showing increases in c-Fos-ir varying depending upon the species, strain, and phenotype of the animal examined. Despite the variation, a pattern of c-Fos-ir emerges with the ARC and PVH (magnocellular and parvicellular regions), typically showing labeling and occasionally the CeA. Exogenous ghrelin administration triggers c-Fos-ir in the ARC NPY/AgRP expressing neurons [e.g., (9, 49, 52)] as well as promoting increases in NPY and AgRP mRNA expression [e.g., (29, 34)]. Therefore, it is not surprising that central NPY or AgRP administration increases c-Fos-ir [e.g., (23, 57)]. In the present experiment, at our first time point, 90 min postinjection, the increased c-Fos-ir occurred in a distribution pattern similar to that of central NPY [e.g., (36, 56)] or AgRP [e.g., (23, 57)] given alone by others in different species assayed within 2 h after injection of either peptide. In brief, in the present finding, only when acute coinjection of subthreshold doses of both AgRP and NPY did c-Fos-ir increase 90-min postinjection in the PVH and pFA, but not additional areas (sZI, VMH, ARC, CeA, NTS, and AP) shown by others in other species to be responsive to NPY and/or AgRP at concentrations above threshold (23, 36, 56, 57) or in the present study (sZI and CeA) at the later time point postinjection (14 h).

The typical time course used for assaying c-Fos-ir as a marker of neural activation is within the first 2 h of treatment, with longer intervals seldom used, likely because of the classic immediate-early characteristic of this marker [23–24 h; (23, 57)] as well as the likelihood of secondary sites of action showing neuronal activation. Using later time points, however, can permit assessment of behaviors or treatments that differ in their temporal signature, for example, the lingering increases in the orexigenic effects of suprathreshold acute central injection of AgRP (23, 57). In laboratory rats, a single AgRP injection causes neuronal activation that is still apparent after 23–24 h in several brain areas (pFA, ARC, PVH, CeA, and NTS) and differs, in part, from the c-Fos-ir at 2 h postinjection (PVH, DMN). Here we used a second time point that was 14 h after treatment to find brain regions that could be uniquely activated at a time when food hoarding is maximally elevated, but not food intake. At the later time point (14 h), there was a different c-Fos-ir profile than that after 90 min. Specifically, c-Fos-ir increased in the PVH (magnocellular and parvicellular regions), sZI, pFA, and CeA, and not in the ARC, VMH, NTS, or AP. Individual injection of subthreshold doses of NPY or of AgRP did not increase c-Fos-ir compared with saline, a result expected given the behavioral data. Therefore, these data provide evidence that NPY and AgRP interact to stimulate areas of the brain known to be activated by neurochemical controllers of energy intake/expenditure (i.e., NPY, AgRP, ghrelin). In addition, the data offer possible discrete locations for the control of different portions of the ingestive behavior sequence. Specifically, the sZI and CeA exhibit an interesting pattern of c-Fos-ir, with increased immunoreactivity coinciding with the increase in food hoarding. For example, taken at face value, it could be argued that the combination of NPY and AgRP together act to increase food intake/foraging and hoarding through unique structures and/or messaging systems. The present data supports previous data that AgRP can act via the CeA to cause the extended increases in ingestive behavior (23, 57) and leads to a hypothesis positing that administration of a subthreshold dose of NPY potentiates the effect of a subthreshold dose of AgRP, or vice versa, on food hoarding perhaps involving the CeA. A more direct examination of these sites is warranted to further elucidate the mechanism by which food intake and hoarding are dissociated in response to food deprivation and the accompanying physiological signals.

It is possible that the different patterns of c-Fos-ir seen between the two points is due to the different time of day that the tissue was collected, but saline-induced c-Fos-ir is not different between the two time points. Another possible explanation for the increased c-Fos-ir is the increased food intake in the NPY + AgRP-treated animals. This is a difficult potential confound to eliminate experimentally, where to our knowledge the choices are to clamp food intake of NPY + AgRP animals to that of controls (pair feeding) as done by Zheng et al. (57) in their animals injected with AgRP 3V and sampled 23 h later or have the animals food deprived for the entire period. Both add the effects of caloric restriction/deprivation into the mix of the peptide injections, however. Thus, the perfect control escapes us.

In previous ingestive behavioral studies, we aimed, in part, to determine the individual roles of NPY and AgRP, and much of the work is detailed above (17, 18). Our present findings, as well as those of others (23, 24), have led to a possible alternative explanation of our initial work, especially regarding melanotan II (MTII), a MC 3/4R agonist, administration after a food deprivation (31). In brief, 3V administration of MTII only partially attenuates the increases in food hoarding after a 48-h food deprivation, but MTII was able to inhibit ghrelin-induced increases in food hoarding (30). The differences in efficacy of MTII to inhibit food hoarding was initially interpreted as being due to other food deprivation-specific factors even though food deprivation does cause increased ghrelin release. This interpretation is certainly possible (8, 25), but it overlooks the gradual increase in serum concentrations of ghrelin that occurs over the duration of the food deprivation, and this increase may have already initiated the AgRP/MC...
NPY/AgRP COINJECTION STIMULATES APPETITIVE BEHAVIORS

In the present experiments, we demonstrated, for the first time, that the coadministration of subthreshold doses of NPY and AgRP are able to interact to increase food intake and hoarding and neural activation in the PVH, pFA, sZI, and CeA; other brain regions were examined for increases in c-Fos-ir, but nothing of note was observed. The increase in food intake was within the first hour, and food hoarding was not increased until 4–24 h and 2 days postinjection. The c-Fos-ir pattern was unique to the time at which the tissue was collected and combined with recent hoarding-related c-Fos-ir data in Mongolian gerbils in brain reward circuitry (55); it may be possible to further dissect the mechanism(s) underpinning the temporal disassociation of food intake and food hoarding.

Perspectives and Significance

The prevalence of overweight/obese individuals continues to rise and with it the incidence of many secondary health consequences, such as, but not limited to, type 2 diabetes, stroke, heart disease, and some types of cancer (e.g., for review see Ref. 26). The control of energy balance is of paramount importance in the onset/maintenance/reversal of obesity, and this has been a focus of much research. Despite the considerable gains in understanding the underlying mechanisms controlling the consummatory phase of ingestive behavior, relatively little is known about the appetitive phase. We have recently reviewed the relevance of food hoarding to normal human ingestive behavior as well as in pathological conditions (e.g., Prader-Willi Syndrome), including overweight/obesity (for review, see Refs. 6 and 7). The present data address both phases of ingestive behavior using an animal model not unlike golian gerbils in brain reward circuitry (55); it may be possible for a deeper understanding of the behavioral sequence leading to energy imbalance. Using doses of NPY and AgRP in tandem that, on their own, are subthreshold to cause food deprivation-like induced increases in ingestive behavior, we were able to show temporal separation of food intake and food hoarding with distinctive patterns of neural activation (i.e., c-Fos-ir). These data, showing the interaction of two neuroactive substances that increase these appetitive behaviors, is another example of the interaction of neuroactive substances that affect ingestive behaviors, such as the interaction of cholecystokinin and leptin (2), cholecystokinin and estradiol (22), or amylin and leptin (43), although those interactions are examples of substances that decrease consummatory ingestive behavior (eating). Only one previous study showed increases based on combination of subthreshold doses of two orexigenic factors (46). Collectively, these interactions demonstrate the complex interplay between neuroactive factors that have been shown to be at least sufficient, and in some cases necessary, for the control of ingestive behaviors. Such cross-talk has the potential for therapeutic treatments of appetitive and/or consummatory ingestive behaviors.

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Present address of Erin Keen-Rheinhart: Department of Biology, Susquehanna University, Selinsgrove, PA 17870.

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In the present experiments, we demonstrated, for the first time, that the coadministration of subthreshold doses of NPY and AgRP are able to interact to increase food intake and hoarding and neural activation in the PVH, pFA, sZI, and CeA; other brain regions were examined for increases in c-Fos-ir, but nothing of note was observed. The increase in food intake was within the first hour, and food hoarding was not increased until 4–24 h and 2 days postinjection. The c-Fos-ir pattern was unique to the time at which the tissue was collected and combined with recent hoarding-related c-Fos-ir data in Mongolian gerbils in brain reward circuitry (55); it may be possible to further dissect the mechanism(s) underpinning the temporal disassociation of food intake and food hoarding.

Perspectives and Significance

The prevalence of overweight/obese individuals continues to rise and with it the incidence of many secondary health consequences, such as, but not limited to, type 2 diabetes, stroke, heart disease, and some types of cancer (e.g., for review see Ref. 26). The control of energy balance is of paramount importance in the onset/maintenance/reversal of obesity, and this has been a focus of much research. Despite the considerable gains in understanding the underlying mechanisms controlling the consummatory phase of ingestive behavior, relatively little is known about the appetitive phase. We have recently reviewed the relevance of food hoarding to normal human ingestive behavior as well as in pathological conditions (e.g., Prader-Willi Syndrome), including overweight/obesity (for review, see Refs. 6 and 7). The present data address both phases of ingestive behavior using an animal model not unlike golian gerbils in brain reward circuitry (55); it may be possible for a deeper understanding of the behavioral sequence leading to energy imbalance. Using doses of NPY and AgRP in tandem that, on their own, are subthreshold to cause food deprivation-like induced increases in ingestive behavior, we were able to show temporal separation of food intake and food hoarding with distinctive patterns of neural activation (i.e., c-Fos-ir). These data, showing the interaction of two neuroactive substances that increase these appetitive behaviors, is another example of the interaction of neuroactive substances that affect ingestive behaviors, such as the interaction of cholecystokinin and leptin (2), cholecystokinin and estradiol (22), or amylin and leptin (43), although those interactions are examples of substances that decrease consummatory ingestive behavior (eating). Only one previous study showed increases based on combination of subthreshold doses of two orexigenic factors (46). Collectively, these interactions demonstrate the complex interplay between neuroactive factors that have been shown to be at least sufficient, and in some cases necessary, for the control of ingestive behaviors. Such cross-talk has the potential for therapeutic treatments of appetitive and/or consummatory ingestive behaviors.

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