MAFbx, MuRF1, and the stress-activated protein kinases are upregulated in muscle cells during total knee arthroplasty

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MAFbx, MuRF1, and the stress-activated protein kinases are upregulated in muscle cells during total knee arthroplasty.

During total knee arthroplasty (TKA), a tourniquet is routinely placed around the proximal thigh of the operative leg to help ensure rigid bone-implant cementing and to provide the surgeon with a clear surgical field. Previous reports, however, demonstrate that muscle tissue is susceptible to periods of ischemia lasting 15 (1), 30 (1, 39, 42, 43), and 60 min (1, 12, 34, 39, 41), with increasing ischemic times causing more severe tissue injury. It follows then that tourniquet-induced ischemia-reperfusion (I/R) may cause damage to muscle cells during TKA and potentially play a mechanistic role in the rapid, and, more importantly, potentially irrecoverable, muscle loss that results in long-term atrophy-induced functional impairment for these older adults.

We have recently published data demonstrating that proteins regulating cap-dependent translation initiation and elongation were downregulated during and immediately after TKA (36). However, although reductions in muscle protein synthesis can account for the majority of atrophy during disuse, stimulation of muscle protein breakdown, i.e., catabolism, may also occur. Indeed, in our previous report (36), we measured a 12% decrease in midtibial quadriceps volume of the operative (TKA) leg vs. a 6% loss in the nonoperative leg within 2 wk of surgery. We interpreted the differences in the degree of atrophy as potentially being related to the tourniquet-induced I/R injury occurring in the operative (TKA) leg.

Protein synthesis and degradation in skeletal muscle cells are tightly regulated by transcriptional and translational mechanisms (15, 21). These processes define a balance between anabolic and catabolic metabolism and are primarily coordinated through the phosphoinositols 3-kinase (PI3K)/Akt/Forkhead boxO3a (FoxO3a) pathways (16). During anabolic conditions, such as with insulin-like growth factor-1 (IGF1) (7), Akt activation of the mammalian target of rapamycin (mTOR) pathway leads to increases in muscle protein synthesis, while under catabolic conditions, such as with glucocorticoids (3) or disuse (37), downregulation of Akt signaling leads to translocation of FoxO3a to the nucleus and upregulation of proteins controlling muscle atrophy (19, 26). Under catabolic conditions, FoxO3a induces the 26S proteasomal pathway by increasing the expression of, in particular, two muscle-specific E3 ubiquitin ligases, muscle atrophy F-Box (MAFbx; also known as atrogin 1), and muscle RING finger 1 (MuRF1) (6, 22), which are expressed in all muscle atrophy models. In addition to regulating components of the proteasomal pathway, FoxO3a also regulates the autophagic/lysosomal pathway by binding to sites on the promoters of Bcl2/adenovirus E1B
19-kDa-interacting protein 3 (Bnip3) gene. Bnip3 is a proapoptotic member of the Bcl-2 family (28) and has been shown to be upregulated during I/R injury and induction of the autophagic/lysosomal pathway (18). Thus, the Akt/FoxO pathways govern protein catabolism through regulation of the ubiquitin-proteasomal and autophagic/lysosomal pathways.

Our objective, therefore, was to measure transcriptional, translational, and protein changes in the components of the catabolic FoxO3a pathway, i.e., MuRF1, MAFbx, and Bnip3, as well as cell stress pathways, SAPK/JNK and the MAPKs, during TKA. We hypothesized that, in addition to the Akt/4E-BP1 pathway being downregulated (36), the cellular and molecular mechanisms controlling protein catabolism would be upregulated during knee replacement surgery, and as such, help us to further determine how tourniquet-induced I/R injury to muscle cells may contribute to the twofold greater quadriceps atrophy measured in the operative vs. nonoperative leg 2 wk after TKA (36).

METHODS

Ethics approval. This study was approved by the PeaceHealth Institutional Review Board, Sacred Heart Medical Center, at Riverbend and the Biomedical Institutional Review Board for the University of Oregon and conducted in accordance with the Declaration of Helsinki. All subjects gave informed written consent prior to study participation. This study is registered with ClinicalTrials.gov (identifier: NCT00760383).

Subjects. We recruited 12 subjects (8 females and 4 males) between 60 and 80 years of age (average age 70 ± 5 yr) from a pool of surgical candidates from the Slocum Center for Orthopedics and Sports Medicine. All subjects were scheduled to undergo a primary total knee arthroplasty (TKA) and had no current untreated endocrine disease, significant heart, kidney, liver, blood or respiratory disease, peripheral vascular disease, active cancer, recent treatment with anabolic steroids, or oral corticosteroids for greater than 1 wk, and no alcohol or drug abuse. Subject characteristics are provided for each subject in Table 1.

Study design. Details of the study design have been published previously (36). On the morning of surgery, subjects were admitted to Sacred Heart Medical Center at Riverbend in a fasted state. Anesthesia was administered with either a epidural, spinal, or general anesthetic, along with a preoperative femoral nerve block placed for postoperative analgesia. Intravenous propofol was used to induce general anesthesia and was maintained with inhalational anesthesia (either desflurane or sevoflurane), with or without muscle relaxant (rocuronium bromide) (Table 1). A 10-cm-wide Zimmer tourniquet was positioned around the proximal third of the thigh and not inflated. Just before the surgery was ready to begin, the first of three muscle biopsies was obtained from the vastus lateralis muscle on the operative (TKA) leg using a 5-mm Bergström biopsy needle with applied suction, as previously detailed (36) and Fig. 1. Following the first biopsy, the tourniquet was inflated to 300 mmHg or greater, depending on systemic blood pressure to ensure minimal blood flow to the operative leg. A second muscle biopsy was obtained immediately prior to tourniquet deflation, after completion of the main components of the surgery. The tourniquet was then deflated, allowing for reperfusion of the limb with blood. The final muscle biopsy was obtained in the operating room prior to being moved to postoperative recovery. Total tourniquet time (ischemia) was 43 ± 2 min, and reperfusion time was 16 ± 1. Immediately after each biopsy, muscle samples were blotted, and adipose tissue was removed before being frozen in liquid nitrogen (<1 min). Samples were stored in −80°C until analysis, which was routinely performed within 2 wk.

Whole-muscle homogenization. Details for the homogenization procedures have been previously published but have been modified for analysis for this study (10). Frozen muscle samples (25–50 mg)

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*Medication withheld week prior to surgery. Dx, diagnosis; OA, osteoarthritis; FNB, femoral nerve block; Gen, general anesthesia; Spinal, spinal anesthesia; Epd, epidural anesthesia. Femoral nerve block: 30 ml of 0.25% to 0.5% bivpicaine or ropivacaine. General anesthesia: intravenous propofol and maintained by inhalation of either desflurane or sevoflurane. Spinal anesthesia: 0.75% bupivacaine + 20 µg of fentanyl. Epidural anesthesia: 0.25% bupivacaine. Muscle relaxant: Administered by local injection of rocuronium bromide.
were crushed using Heidolph Brinkmann Silent Crusher M in homogenization (1.9, w/v) buffer containing: 50 mM Tris-HCl, 250 mM mannitol, 50 mM Na pyrophosphate, 1 mM EDTA, 1 mM EGTA, 1.0% Triton X-100, pH 7.4, 1 mM benzamidine, 1 mM DTT, 0.1 mM PMSF, and 5 μM/ml soybean trypsin inhibitor. Samples were centrifuged at 2,817 g at 4°C, and the supernatant (cytoplasmic extract) was collected, and nuclear extraction was performed with 4E-BP1. Protein concentration (mg/ml) was determined in duplicate using a Qubit Protein Assay Kit (no. Q3321; Invitrogen, Carlsbad, CA) on a Qubit 2.0 Fluorometer (Invitrogen).

Nuclear and cytoplasmic homogenization and fractionation. Isolation of nuclear and cytoplasmic fractions was done using the NE-PER kit (no. 78835; Thermo Fisher Scientific, Waltham, MA). Frozen muscle samples (25–50 mg) were washed with 1X TBS and then homogenized in cytoplasmic extraction reagent I containing 0.5 mM PMSF. Homogenates were then vortexed and incubated on ice for 10 min. Cytoplasmic extraction reagent II was added to the samples, vortexed, and centrifuged at 16,000 g at 4°C for 5 min. The supernatant (cytoplasmatic extract) was collected, and nuclear extraction reagent, containing 0.5 mM PMSF, was added to the tube to resuspend the pellet. The samples were vortexed and incubated on ice and then centrifuged at 16,000 g at 4°C for 10 min before the supernatant (nuclear extract) was collected. Protein concentration (mg/ml) was determined in duplicate using a Qubit protein assay kit (no. Q3321; Invitrogen) on a Qubit 2.0 fluorometer (Invitrogen). Adequate separation of cytoplasmic and nuclear fractions was confirmed via Western blot detection of α-tubulin (cytoplasmic protein) and laminin A/C (nuclear protein) (Fig. 2).

SDS-PAGE and immunoblotting. Details of the immunoblotting procedures have been previously published (36), with specific modifications implemented for this study. Homogenates were loaded in duplicate into TGX all kD precast gels (Bio-Rad) in electrode buffer (0.3% Tris base, 14.4% glycine, 1% SDS in dd-H2O). A single muscle homogenate sample was loaded in duplicate with each gel and used as a between-blot internal loading control. All gels were run in duplicate with cytoplasmic and nuclear fractions loaded in adjacent lanes, i.e., baseline cytoplasmic next to the baseline nuclear fraction as shown in the representative blot.

Following SDS-PAGE, proteins were transferred to PVDF membranes using Bio-Rad Trans-Blot Turbo Transfer system using the Bio-Rad Midi format, mixed MW presettings (25 V for 7 min). Transfers were verified by staining with Ponceau S.

Proteins isolated by Western blot analysis were made relative to our loading control, which was run in duplicate for cytoplasmic and nuclear fractions, to account for variance between gels, as well as an internal loading control within each lane.

Antibodies. The primary antibodies p-Akt Ser-473 (no. 9271), Akt (no. 9272), p-4E-BP1 Thr-37/46 (no. 9459), 4E-BP1 (no. 9452), p-FoxO3a S253 (no. 9466), FoxO3a (no. 9467), eukaryotic initiation factor 4E phosphorylation (p-eIF-4E) Ser-209 (no. 9741), eIF-4E (no. 9742), p-GSK3α/β S21/9 (no. 9331), GSK3β (no. 9315), Bcl2 (no. 2872), SAPK/JNK (no. 9258), p-38α (no. 2918), p-38β (no. 2339), α-tubulin (no. 2144), laminin A/C (no. 4777) were purchased from Cell Signaling (Beverly, MA, USA). Additional primary antibodies, p-Bcl2 S70 (no. sc-21864), and MABFbx (sc-33782) were purchased from Santa Cruz Biotechnologies (Santa Cruz, CA). MuRF1 (no. ab96857) was purchased from Abcam (Cambridge, MA). Bnip3 (no. B7931) was purchased from Sigma-Aldrich (St. Louis, MO). ECL and Anti-Rabbit IgG, horseradish peroxidase from donkey and mouse (no. 7074, no. 7076). Secondary antibodies for Western blotting were purchased from GE Healthcare and used as our secondary antibodies.

Total RNA isolation and cDNA synthesis. Details for RNA isolation have been previously published (36) with slight modifications implemented for this study. Skeletal muscle samples (10–20 mg) were homogenized in 1 ml TRI Reagent using Heidolph Brinkmann’s Silent Crusher M at 10,000–15,000 rpm in Eppendorf RNase-free tubes. Separation was achieved through the addition of 0.2 ml of chloroform and precipitation with 0.5 ml isopropanol. The RNA pellet was washed twice in 75% ethanol, dried, and then dissolved in 1.5 μl of 0.1 mM EDTA for each 1 mg of starting tissue. RNA concentrations were determined using a Qubit fluorometer (Invitrogen, Carlsbad, CA), and cDNA was reverse transcribed from 1 μg RNA on a CFX96 real-time PCR Detection System (Bio-Rad) using iScript Reaction Mix (Bio-Rad), according to manufacturer’s instructions and stored at −80°C for analysis.

Oligonucleotide primers for qPCR. Oligonucleotide primers were designed using Beacon Design software (ver. 7.91), specific to our Bio-Rad CFX96 real-time PCR Detection System, based on NCBI Entrez Gene ID search results. Primer efficiencies were determined through analysis of four serial dilutions, analyzed in triplicate on a run in triplicate log linear scale: efficiency = 10^{-1/slope}. Efficiencies were in accordance with MIQE guidelines (0.95 and 1.05) for all primer pairs (8). Primers were designed for α-tubulin (NM_006082; Fwd: AGATGCT-GCAATAACATG; Rev: AATTCGCTGACAACAAG); β-tubulin (NM_030773; Fwd: CAGTGGTCGGTCTCGATTA; Rev: GATTAAGGTCAATACCCAT); β2-microglobulin (NM_004048; Fwd: TCTCTTTATCACACATCACCT; Rev: GCACGCTTACATCTTAAAG); Forkhead box O3 (FOXO3) (NM_001455; Fwd: GCATTCTTACTGAGGATT, Rev: ACACAGGCAATATCTAC);
atrogin-1 (MAFbx) (NM_058229; Fwd: TGGATTGGAAGAGATGATT, Rev: AAAAGATGACAGTGGT); muscle ring finger (MuRF) (NM_032588; Fwd: ATGAGGAAGGGAAGAAATT, Rev: CTCTACTGGTGCCTCTCTT); activating transcription factor (ATF4) (NM_001675; Fwd: AGATAGGAAGCCAGACTA, Rev: CTCATACAGATGCCACTA); Jun N-terminal kinase (JNK) (NM_002750; Fwd: TAGTGTAGAGATTGAGATT, Rev: AAGAATGGCATCATAAGC); p-38 (NM_001315; Fwd: GTTCAGTTCCTTTATCTACCA, Rev: GCTCACAGTCTTCATTCA); BCL2 adenovirus E1B 19-kDa protein-binding protein 3 (Bnip3) (NM_004052; Fwd: TCATAATCAAGAAGACCAAGA, Rev: ATCACCTAATAATCGGAGACT); B-cell lymphoma 2 (BCL2) (NM_006538; Fwd: ATATCTGCCTGTATCTTG, Rev: TCTGTTGTTATTACTGCTA); and JunB proto-oncogene (JUNB) (NM_002229; Fwd: GAAACGACCTTCTATGAC, Rev: GGTTACTGTAGCCATAAG).

mRNA quantitation by qPCR. Details on qPCR have been previously published (36), with specific modifications employed for this study. Samples of cDNA were analyzed using SYBR Green fluorescence (iQ SYBR Green Supermix; Bio-Rad, Hercules, CA). Each reaction contained 12.5 μl SYBR Green, 9.5 μl DEPC-treated nuclease-free water, 0.5 μl forward and 0.5 μl reverse primers, and 2 μl of cDNA template, and was prepared in triplicate. An initial 5-min cycle at 95°C was used to denature the cDNA. This was followed by 50 PCR cycles consisting of denaturation at 95°C for 10 s followed by 30 s of primer annealing at the optimized primer pair-annealing temperature. All PCR cycles were followed by a melt curve analysis.

Fig. 3. Protein translation downregulation during total knee arthroplasty (TKA). Data represent Akt at Ser-473 in whole muscle fraction (A) (n = 6); 4E-BP1 Thr-36/47 (B); eukaryotic initiation factor 4E (eIF-4E) at Ser-209 in the cytoplasmic (C) and nuclear fraction (D). Representative phosphorylated, total, and loading control (Ponceau) Western blot are included. Data are expressed as means ± SE (n = 10). *P ≤ 0.05 vs. baseline.
Fig. 4. Stress pathway during TKA. Stress-activated protein kinase (SAPK/JNK) protein in the cytoplasmic (A) and nuclear fraction (B). MAPK p-38α protein in the cytoplasmic (C) and nuclear fraction (D). MAPK p-38β protein in the cytoplasmic (E) and nuclear fraction (F). Representative phosphorylated, total, and loading control (Ponceau) Western blot are included. Data are expressed as means ± SE (n = 10). *P ≤ 0.05 vs. baseline.
Each gene of interest (GOI) was normalized to the geometric mean of select genes of reference (GOR); α-tubulin, β-tubulin, and β-2-microglobulin. Data for each GOI are expressed as fold change relative to the GOR using the Livak method, also known as the 2-ΔΔCT method (27): ΔCT(test) = CT(GOI, test) − CT (GOR, test) ΔCT(baseline) = ΔCT(test) − ΔCT(baseline) 2-ΔΔCT = fold change from baseline.

**Statistical analysis.** Statistical evaluation of our data was performed using a repeated-measures ANOVA to compare ischemic and reperfusion samples to baseline. Differences between means were considered significant at $P \leq 0.05$. Differences between means were considered a trend at $P \leq 0.10$. Analysis for all variables was performed using SAS Institute (2009), Base SAS 9.2. All values are expressed as means ± SE, unless otherwise stated.

**RESULTS**

**Demographics.** Male and female subject characteristics were not different for age, height, weight, and body mass index ($P > 0.05$); data for all subjects are in Table 1. The average age for our cohort was 70 ± 1.3 yr. The average height, weight, and BMI was 164 ± 2.7 cm, 84 ± 5.8 kg, and 31 ± 1.8 kg/m², respectively.

**Cell signaling.** Adequate tissue samples from all three biopsies were available for only six subjects for the analysis of whole muscle fractions. These samples were used to confirm our previous results (36). Relative to baseline, phosphorylation of Akt at Ser-473 decreased by 43% during ischemia ($F = 13.71$) ($P = 0.013$, 80% CI [$-72\%$, $-31\%$]) but was not significant during reperfusion ($F = 5.95$) ($P = 0.08$, 80% CI [$-51\%$, $-13\%$]) (Fig. 3A).

There was adequate tissue to analyze cytoplasmic and nuclear protein fractions for 10 subjects. Relative to baseline eukaryotic initiation factor 4E binding protein (4E-BP1) phosphorylation at Thr-37/46/total 4E-BP1 was significantly decreased by 30% ($F = 16.36$) ($P = 0.003$, 80% CI [$-26\%$, $-43\%$]).

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Fig. 5. Autophagic/lysosomal pathway during TKA. Bcl2 protein in the cytoplasmic (A) and nuclear fraction (B). Bcl2/adenovirus EIB 19-kDa-interacting protein 3 (Bnip3) protein in the cytoplasmic (C) and nuclear fraction (D). Representative phosphorylated, total, and loading control (Ponceau) Western blot images are included. Data are expressed as means ± SE ($n = 10$). *$P \leq 0.05$ vs. baseline.
−13%]) and 27% (F = 5.57) (P = 0.043, 80% CI [−30%, −8%]) during ischemia and reperfusion, respectively, in the cytoplasmic fractions (Fig. 3B). Relative to baseline, eIF-4E at Ser-209 increased significantly in the cytoplasmic fraction during ischemia, by 189% (F = 6.54) (P = 0.031, 80% CI (54%, 182%)) but was not significantly different from baseline during reperfusion (Fig. 3C). Relative to baseline, eIF-4E phosphorylation in the nuclear fraction did not change during ischemia or reperfusion (Fig. 3D).

Relative to baseline, SAPK/JNK protein significantly increased in the cytoplasmic fraction during ischemia by 27% (F = 24.12) (P = 0.001, 80% CI (24%, 43%)) (Fig. 4A), while in the nuclear fraction, there was a trend to decrease during reperfusion, −21% (F = 4.24) (P = 0.073, 80% CI (−24%, −5%)) but was not significant (Fig. 4B). Relative to baseline, p-38α protein did not change in the cytoplasmic fraction during ischemia (Fig. 4C), while in the nuclear fraction, there was a trend for it to increase during ischemia by 34% (F = 3.95) (P = 0.082, 80% CI (5%, 30%)) but was not significant, while during reperfusion, it was significantly decreased by 18% (F = 9.42) (P = 0.015, 80% CI [−17%, −6%]) (Fig. 4D). Relative to baseline, p-38β protein tended to increase in the cytoplasmic fraction during ischemia by 33% (F = 4.12) (P = 0.054, 80% CI (16%, 263%)) but was not significant (Fig. 4E), while in the nuclear fraction, there was a significant increase during ischemia by 25% (F = 9.55) (P = 0.015, 80% CI (5%, 30%)) (Fig. 4F).

Relative to baseline, Bcl2 protein showed a trend to increase in the cytoplasmic fraction during ischemia by 97% (F = 5.10) (P = 0.054, 80% CI (24%, 43%)) (Fig. 5A), while in the nuclear fraction, there was no change (Fig. 5B). Bcl2/adenovirus EIB 19-kDa-interacting protein 3 (Bnip3) protein showed a trend to increase in the cytoplasmic fraction during ischemia by 34% (F = 3.95) (P = 0.082, 80% CI (5%, 30%)) but was not significant, while during reperfusion, it was significantly decreased by 18% (F = 9.42) (P = 0.015, 80% CI [−17%, −6%]) (Fig. 5D).

**Fig. 6.** E3 ubiquitin proteasomal pathway during TKA. MAFbx protein in the cytoplasmic (A) and nuclear fraction (B). MuRF1 protein in the cytoplasmic (C) and nuclear fraction (D). Representative phosphorylated, total, and loading control (Ponceau) Western blot images are included. Data are expressed as means ± SE (n = 10). *P ≤ 0.05 vs. baseline.
levels did not change in the cytoplasmic fraction during ischemia or reperfusion (Fig. 5C). Bnip3 protein levels in the nuclear fraction significantly decreased during reperfusion by 22% ($F = 6.60, P = 0.033, 80\% CI (-139\%, -41\%)$) (Fig. 5D). We did not detect significant changes in phosphorylation status in the cytoplasmic or nuclear fractions for p-GSK3β Ser-9; however, we did detect a significant increase in GSK3β total protein during ischemia 42% ($F = 8.26, P = 0.018, 80\% CI (21\%, 60\%)$).

Relative to baseline, the E3 ubiquitin ligase atrogin-1/MAFbx protein increased in the cytoplasmic fraction during ischemia by 30% ($F = 18.30, P = 0.002, 80\% CI (15\%, 29\%)$) (Fig. 6A), while the nuclear fraction significantly decreased during reperfusion by 30% ($F = 5.80, P = 0.043, 80\% CI (-97\%, -26\%)$) (Fig. 6B). As well, relative to baseline, MuRF1 protein increased in the cytoplasmic fraction during ischemia by 53% ($F = 26.33, P = 0.0006, 80\% CI (36\%, 62\%)$) and remained elevated during reperfusion 31% ($F = 18.36, P = 0.002, 80\% CI (18\%, 36\%)$) (Fig. 6C). In the nuclear fraction, MuRF1 protein tended to decrease during reperfusion, by 29% ($F = 4.21, P = 0.074, 80\% CI (-106\%, -20\%)$) but was not significant (Fig. 6D).

**Gene expression.** Adequate tissue was available to analyze gene expression for seven subjects. Relative to baseline, FoxO3a mRNA increased during ischemia by 157% ($F = 18.55, P = 0.008, 80\% CI (66\%, 135\%)$) (Fig. 7A). Relative
to baseline, the E3 ubiquitin ligase MAFbx mRNA increased during ischemia by 237% ($F = 7.37$) [$P = 0.042, 80\% CI (85\%, 288\%)] (Fig. 7B). As well, MuRF mRNA increased during ischemia by 288% ($F = 7.75$) [$P = 0.039, 80\% CI (161\%, 380\%)] (Fig. 7C). Relative to baseline, SAPK/JNK mRNA increased during ischemia by 169% ($F = 15.16$) [$P = 0.012, 80\% CI (95\%, 211\%)] (Fig. 7D) and p-38α mRNA increased during ischemia 173% ($F = 21.80$) [$P = 0.006, 80\% CI (125\%, 242\%)] (Fig. 7E). We did not detect significant changes in transcript levels for Bnip3 and Bcl2.

**DISCUSSION**

I/R occurs each time a tourniquet is used to control blood flow during surgeries, such as TKA. However, the cellular and molecular alterations associated with I/R remain to be fully elucidated. We have recently reported that proteins regulating components of the cap-dependent translation initiation and elongation complex are downregulated with I/R during TKA (36). However, at the cellular level, atrophy is defined by the imbalance between proteins controlling both muscle anabolism and catabolism. Our objective, therefore, was to measure changes in the components of the catabolic FoxO3a pathway, as well as cell stress pathways, during TKA. We hypothesized that cellular mechanisms controlling muscle protein catabolism would be upregulated during surgery, and as such, help us to further determine how I/R injury may potentially contribute to the twofold greater quadriceps atrophy measured in the operative vs. nonoperative leg 2 wk after TKA (36).

This study reveals several important and novel findings regarding the acute effects of I/R in stimulating pathways known to play key roles in protein catabolism in muscle cells (Fig. 8). First, we confirm previous findings obtained by our group (36), showing that the Akt/4E-BP1 pathway is downregulated during ischemia and reperfusion and add further to those findings by including the phosphorylation status of the eIF-4E. Second, we found that protein levels for MuRF and MAFbx, two downstream components of FoxO3a pathway, were upregulated during ischemia but only protein levels for MuRF remained elevated through reperfusion. Third, we measured an increased expression of the SAPK/JNK and the p-38 mitogen activated protein kinase (p-38 MAPK α/β). To our knowledge, this study is the first to measure simultaneous changes in key regulatory proteins known to exert control over both the anabolic and catabolic transcriptional and signaling pathways in skeletal muscle cells during TKA.

4E-BP1’s binding target eIF-4E showed no changes in the total amount of protein present, but there were measurable changes in phosphorylation in the cytoplasmic fraction during ischemia. eIF-4E phosphorylation at Ser-209 was first thought to enhance its binding to the 5'-cap, but it is now believed that phosphorylation of eIF-4E may have the opposite effect, decreasing its binding affinity (30, 38, 40). Mitogen- or stress- and cytokine-activated signaling regulates eIF-4E phosphorylation to a greater extent than anabolic stimuli (33), and the significance of its phosphorylation is largely undetermined in mammals. It is possible that eIF-4E phosphorylation during...
MAFbx AND MuRF1 ACTIVATION DURING TKA

Perspectives and Significance

Despite the considerable success of TKA in mitigating knee pain due to osteoarthritis, chronic limitations in functional mobility remain and are primarily due to irrecoverable quadriceps muscle loss. Indeed, quadriceps atrophy is the main contributor to long-term strength deficits (29), and by far the most significant long-term clinical barrier following TKA surgery is persistent muscle atrophy (14, 31, 44). Broadly speaking, acute muscle loss for older adults represents an acceleration of sarcopenia, i.e., the gradual ~1%/year of muscle lost due to aging, per se (11). In other words, a 12% loss in quadriceps volume occurring within 2 wk of surgery (36) would represent ~12 yr of “normal” aging. This “accelerated sarcopenic muscle loss” (32) is potentially devastating for older women who, compared with older men, demonstrate a blunted capacity to increase muscle volume (4).

In conclusion, we interpret our findings to support our hypothesis that factors resulting from ischemia are sufficient to upregulate components of the FoxO3a catabolic pathway during TKA and that cellular stress pathways may play a role in stimulating this response. These data extend our understanding of the effects of I/R to now include proteins regulating anabolic (36), as well as catabolic and stress-activated, pathways being altered during TKA. Additional studies are necessary to better understand the mechanisms underlying the rapid-onset muscle atrophy that occurs within weeks of this surgery and, in particular, the means by which potentially irrecoverable muscle atrophy may be avoided in older women.

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DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the authors.

AUTHOR CONTRIBUTIONS


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