Resveratrol supplementation improves white adipose tissue function in a depot-specific manner in Zucker diabetic fatty rats

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Beaudoin M, Snook LA, Arkell AM, Simpson JA, Holloway GP, Wright DC. Resveratrol supplementation improves white adipose tissue function in a depot-specific manner in Zucker diabetic fatty rats. Am J Physiol Regul Integr Comp Physiol 305: R542–R551, 2013. First published July 3, 2013; doi:10.1152/ajpregu.00200.2013.—Resveratrol (RSV) is a polyphenolic compound suggested to have anti-diabetic properties. Surprisingly, little is known regarding the effects of RSV supplementation on adipose tissue (AT) metabolism in vivo. The purpose of this study was to assess the effects of RSV on mitochondrial content and respiration, glyceroenogenesis (GNG), and adiponectin secretion in adipose tissue from Zucker diabetic fatty (ZDF) rats. Five-week-old ZDF rats were fed a chow diet with (ZDF RSV) or without (ZDF chow) RSV (200 mg/kg body wt) for 6 wk. Changes in adipose tissue metabolism were assessed in subcutaneous (scAT) and intra-abdominal [retroperitoneal (rpWAT), epididymal (eWAT)] adipose tissue depots. ZDF RSV rats showed lower fasting glucose and higher circulating adiponectin, as well as lower glucose area under the curve during intraperitoneal glucose and insulin tolerance tests than ZDF chow. [14C]pyruvate incorporation into triglycerides and adiponectin secretion were higher in scAT from ZDF RSV rats, concurrent with increases in adipose tissue triglyceride lipase (ATGL), hormone-sensitive lipase (HSL), and monoglyceride lipase (MGL) (12). Furthermore, phosphorylation of perilipin is necessary to allow access of lipases to the lipid droplet (12). RSV-mediated lipolysis is associated with increases in ATGL protein content (25) and cAMP (41), an upstream activator of HSL via the activation of protein kinase A (PKA).

Importantly, up to 35% of FFA liberated through lipolysis is recycled back into triglyceride (TG) within the adipocyte (48), and this is dependent on the provision of glycerol-3-phosphate (G3P) through nonglucose precursors, such as pyruvate, in a process called glyceroenogenesis (GNG) (7). GNG is regulated through the concerted actions of pyruvate dehydrogenase kinase 4 (PDK4) and phosphoenolpyruvate carboxykinase (PEPCK). In mice, the knockdown of pck1, the gene coding for PEPCK, results in whole body insulin resistance and increased FFA and glycerol release during a euglycemic-hyperinsulinemic clamp (27). Conversely, overexpression of PEPCK in adipose tissue led to obesity without insulin resistance (13). Absolute rates of fatty acid (FA) reesterification mirror that of lipolysis (4), and the reesterification of FAs is one of the largest consumers of ATP in fat cells (34). In fact, work from Ruderman’s group (15) has demonstrated that the activation of the energy-sensing enzyme 5’-AMP-activated protein kinase (AMPK) by β-adrenergic agonists occurs secondary to increases in FA reesterification. AMPK is a reputed mediator of mitochondrial biogenesis (14, 17) and thus RSV-mediated increases in ATGL and lipolysis may lead to increases in FA reesterification, the activation of AMPK, and the induction of mitochondrial biogenesis in adipose tissue. In support of this premise, adipose tissue-specific overexpression of ATGL in mice resulted in increased lipolysis without elevations in plasma FFA, TG, or ectopic lipid deposition, along with increases in whole body TG cycling and adipose tissue mitochondrial enzyme gene expression (1). To the best of our knowledge, it is currently unknown whether RSV can induce mitochondrial biogenesis and increase GNG in adipose tissue in vivo.

Adiponectin is an anti-inflammatory adipokine that stimulates FA oxidation in skeletal muscle and liver and is associated with improved insulin sensitivity (20, 52). RSV treatment increased adiponectin mRNA in human adipocytes isolated...
from visceral depots (11) and elevated adiponectin protein content in 3T3-L1 adipocytes (47) and in rodent epididymal and retroperitoneal adipose tissue (33). Interestingly, mitochondriala biogenesis has been shown to be essential for adiponectin synthesis and secretion in adipocytes (20) and thus, it seems plausible that RSV-mediated increases in mitochondrial biogenesis could be associated with increases in adiponectin secretion. Conversely, interleukin-6 (IL-6) and tumor necrosis factor-α (TNF-α) are pro-inflammatory adipokines that are negatively regulated by RSV in vitro (2, 18), but the effects of RSV on these cytokines in vivo remain to be elucidated.

The purpose of the current study was to examine the effects of dietary RSV supplementation on mitochondrial biogenesis, GNG, and adipokines secretion in white adipose tissue (WAT) from Zucker diabetic fatty (ZDF) rats, a rodent model of T2DM. In addition, as RSV is known to improve skeletal muscle metabolism and whole body glucose homeostasis (4, 23), it was important to determine whether RSV may have direct effects on adipose tissue. To address this question, we used an acute ex vivo approach to assess gene expression in adipose tissue explants from untreated ZDF rats. We hypothesized that RSV supplementation in ZDF rats would induce mitochondrial biogenesis, which would be associated with enhanced GNG, increased adiponectin secretion, and the prevention of insulin resistance. Furthermore, these effects could be mediated, at least in part, by direct actions of RSV onto adipose tissue.

METHODS

Animals. Four-week-old male ZDF rats (Charles River, St. Constant, QC, Canada) weighing ~100 g were individually housed in wire-bottom cages, in a temperature-controlled room with a reverse 12:12 h light-dark cycle, and were provided with food and water ad libitum. The 12-h light cycle started at 9 PM, and experiments were performed between 9 AM and 4 PM. After a 10-day acclimatization period, ZDF rats were fed either a standard powdered chow diet (Purina 5008 diet; Purina, St. Louis, MO) (ZDF chow) or a chow diet supplemented with RSV (Cayman Chemical, Ann Arbor, MI) (Purina 5008 diet; Purina, St. Louis, MO) (ZDF chow) or a chow diet supplemented with RSV (Cayman Chemical, Ann Arbor, MI) (~200 mg/kg body wt (BW) (ZDF RSV) for 6 wk. RSV was mixed directly into the powered diet on a weekly basis, based on predicted body weight and food intake for that week. ZDF rats are characterized by the development of insulin resistance by 6 wk of age and T2DM by 12 wk, therefore, we attempted to delay the onset T2DM by treating with RSV between 5 and 11 wk of age (39). We acknowledge that the ZDF model is a genetic model of T2DM and that the dose of RSV used in the present study is likely not attainable through diet alone. However, we used this approach to study the development of T2DM. Moreover, while the RSV dose is large, it is similar to many other rodent-based reports in the literature (23, 32, 43). Food intake was recorded three times weekly while BW was assessed weekly. All protocols followed Canadian Council on Animal Care guidelines and were approved by the Animal Care Committee at the University of Guelph.

Tolerance tests. Intraperitoneal glucose (ipGTT) and insulin tolerance (ipITT) tests were performed before (5 wk old) and after (11 wk old) the feeding intervention. After measurement of fasting (12 h overnight) blood glucose (Freestyle lite, Abbott Laboratories, North Chicago, IL), adipose tissue (AT) from the inguinal subcutaneous (scAT), epididymal (eWAT), and retroperitoneal (rpWAT) depots were harvested. For each depot, 100 mg was used for GNG determination, 25 mg was used for respiration experiments, 50 mg was fixed in Formalin, while the rest of the tissue was immediately frozen in liquid nitrogen and stored at −80°C for later determination of protein content and gene expression. Blood was collected in polypropylene tubes through cardiac puncture, allowed to clot at room temperature, and centrifuged, and the supernatant was collected and stored at −20°C for further analysis of triglycerides (colorimetric assay: Sigma-Aldrich, Oakville, ON, Canada), FFAs (colorimetric assay: Wako Diagnostics, Richmond, VA), total adiponectin (ELISA: Linco Research, St. Charles, MO), insulin (ELISA: Millipore, St. Charles, MO), IL-6 (ELISA: BioLegend, San Diego, CA), and TNF-α (ELISA: BioLegend) through commercially available kits.

Ex vivo incubation procedures. Adipose tissue (100 mg) from each of scAT, eWAT, and rpWAT was minced and placed in a plastic vial containing 3 ml oxygenated Krebs-Ringer buffer (KRB: 118 mM NaCl, 4.8 mM KCl, 1.25 mM CaCl2, 1.2 mM KH2PO4, 1.2 mM MgSO4, 25 mM NaHCO3, pH 7.4) containing 5 mM glucose and 2.5% FFA-free bovine serum albumin (BSA; MP Biomedicals, Santa Ana, CA) and incubated at 37°C in a shaking water bath (60 rpm) for 2 h. At the end of the incubation, medium was collected in polypropylene tubes and stored at −20°C for later availability of total adiponectin, IL-6, and TNF-α using commercially available kits (see providers above).

14Cpyruvate incorporation into triglycerides. Quantification of the incorporation of [14C]pyruvate into triglycerides is a measure of GNG within adipose tissue (24). Adipose tissue (100 mg) from each of scAT, eWAT, and rpWAT was minced and placed in a plastic vial containing 2 ml oxygenated Krebs-Ringer buffer (KRB: 118 mM NaCl, 4.8 mM KCl, 1.25 mM CaCl2, 1.2 mM KH2PO4, 1.2 mM MgSO4, 25 mM NaHCO3, pH 7.4) containing 5 mM glucose and 2.5% FFA-free BSA, and 1 μCi of [14C]pyruvate (PerkinElmer, Woodbridge, ON, Canada). Tissue was incubated at 37°C in a shaking water bath (60 rpm) for 1 h. At the end of the incubation, the reaction was stopped by adding 250 μl of 5 N sulfuric acid (Fisher Scientific, Ottawa, ON, Canada) into the vials and incubating them on ice for 1–2 min. Adipose tissue was then minced and transferred to new tubes for lipid extraction with 5 ml of a 2:1 solution of chloroform:methanol (both from Fisher Scientific). One milliliter of phosphate buffer saline (PBS: 0.137 M NaCl, 2.68 mM KCl, 1.47 mM KH2PO4, 8.03 mM NaHPO4; pH 7.4) was added, and the tubes were vortexed and centrifuged before removal of the upper aqueous phase. This washing step was repeated twice. The chlorormethanol solution was allowed to evaporate overnight in a fume hood, after which two 500-μl aliquots were transferred to scintillation vials, 3 ml of scintillation fluid (MP Biomedicals) was added, and 14C radiation was determined over 5 min in a beta-counter (Beckman-Coulter, LS6500 Scintillation Counter, Mississauga, ON, Canada).

Respiration. Adipose tissue respiration was determined as previously described (21), with minor modifications (46). Briefly, adipose tissue was excised, immediately placed in polypropylene tubes containing 1 ml of MiR05 buffer (0.5 mM EGTA, 3 mM MgCl2·6H2O, 60 mM K-lactobionate, 10 mM KH2PO4, 20 mM HEPES, 20 mM taurine, 110 mM sucrose, 1 g/l FFA-free BSA; pH 7.1), and minced with scissors. Adipose tissue was then transferred to 1 ml of fresh MiR05 buffer and left on ice for 10 min. Thereafter, tissue was blotted and 25 mg used to determine rates of oxygen consumption by high-resolution respirometry (Oroboros Oxygraph-2 k, Innsbruck, Austria). Digitonin or saponin are used to permeabilize cell membranes while leaving mitochondrial membranes intact due to their specificity to solubilize cholesterol, which exists in much higher concentrations on the plasma membrane. This is a common procedure to permit exogenously added substrates free access for diffusion to the mitochondria. However, we found mincing without permeabilization
to be sufficiently effective in disrupting the cell membrane without compromising mitochondrial intactness. Specifically, robust respiration was detected in minced adipose tissue and this was not increased with the addition of either digitonin or saponin following standard permeabilizing procedures. This has also been reported previously (21). Therefore, all experiments were performed in the absence of either chemical. Respiration experiments were performed at 37°C in room air-saturated MiR05 buffer. The sequential respiration protocol consisted of determining state IV (leak) respiration in the presence of 5 mM ADP (complex I), and 10 mM succinate (complex I and II). The subsequent titration of carbonyl cyanide 4-(trifluoromethoxy)phenylhydrazone (FCCP) yielded maximal uncoupled respiration. Rates of oxygen consumption are expressed per milligram wet weight.

**Histochemistry.** scAT was fixed in 10% neutral buffered Formalin (VWR, Mississauga, ON, Canada) dehydrated in xylene (Fisher Scientific) and embedded in paraffin. We chose to further analyze only scAT as this depot responded most robustly to RSV supplementation (20, 24, 25). To assess the direct effects of RSV on gene expression, determination of changes in adipose tissue metabolism and gene expression was a well-characterized technique that is used regularly to study tissue culture. Tissue was homogenized in 2 volumes of ice-cold cell lysis buffer (Invitrogen, Burlington, ON, Canada) in a homogenizer (FastPrep-24, MP Biomedicals). Homogenized samples were centrifuged for 10 min at 1,500 g at 4°C. Lipids were carefully removed, and the protein-containing infranant was collected for determination of protein concentration using a bicinchoninic acid assay (37) (ThermoScientific, Rockford, IL). Western blotting was performed as previously described by our laboratory (40, 44, 45). Membranes were incubated in primary antibodies diluted in TBST/5% nonfat dry milk (COX4, MitoSciences, Eugene, OR; α-tubulin, perilipin A: Abcam, Toronto, ON, Canada; ATGL, p-HSL[Ser660]: Cell Signaling, Danvers, MA) or TBST/5% bovine serum albumin [p-PDH [Ser293], uncoupling protein-1 (UCP-1): Calbiochem, Mississauga, ON, Canada; PDH, MitoSciences; PEPCK, Cayman Chemicals, Ann Arbor, MI; HSL, p-HSL[Ser563], AMPK, p-AMPK-[Thr172]: Cell Signaling: diglyceride acyltransferase (DGAT1), glycerolphosphate acyltransferase (GPAT): Abcam; DGAT2: Immunex, San Diego, CA) overnight at 4°C. Subsequently, membranes were washed in TBST and incubated in appropriate horseradish peroxidase-conjugated secondary antibodies (Jackson Immunoresearch, West Grove, PA) diluted in TBST/1% nonfat dry milk for 1 h at room temperature. Signal was detected through enhanced chemiluminescence (ThermoScientific) and quantified by densiometry (Fluorchem HD2, ProteinSimple, Santa Clara, CA). Intracellular cAMP content of scAT and rpWAT was determined by ELISA according to the manufacturer’s instructions (Enzo Life Sciences, Brockville, ON, Canada).

**Table 1. Body and organ weights and fasting blood parameters at euthanasia for ZDF chow and ZDF RSV animals**

<table>
<thead>
<tr>
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<th>ZDF Chow (n = 11)</th>
<th>ZDF RSV (n = 13)</th>
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<tr>
<td>Body weight, g</td>
<td>381 ± 7</td>
<td>383 ± 5</td>
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<tr>
<td>Average daily food intake, g</td>
<td>28.7 ± 1.0</td>
<td>27.3 ± 0.9</td>
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<tr>
<td>scAT (inguinal) weight, g</td>
<td>21.1 ± 1.5</td>
<td>22.0 ± 1.3</td>
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<tr>
<td>eWAT weight, g</td>
<td>7.2 ± 0.4</td>
<td>7.2 ± 0.3</td>
</tr>
<tr>
<td>rpWAT weight, g</td>
<td>5.2 ± 0.2</td>
<td>5.0 ± 0.2</td>
</tr>
<tr>
<td>Liver weight, g</td>
<td>14.6 ± 0.6</td>
<td>13.1 ± 0.5</td>
</tr>
<tr>
<td>Fasting glucose, mmol/l</td>
<td>9.3 ± 1.6</td>
<td>5.1 ± 0.2*</td>
</tr>
<tr>
<td>Fasting insulin, pmol/l</td>
<td>723 ± 145</td>
<td>1101 ± 208</td>
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<tr>
<td>Fasting FFA, mmol/l</td>
<td>1.36 ± 0.16</td>
<td>1.40 ± 0.14</td>
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<tr>
<td>Fasting glycerol, mmol/l</td>
<td>0.45 ± 0.03</td>
<td>0.50 ± 0.05</td>
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<tr>
<td>FA:glycerol ratio</td>
<td>3.0 ± 0.3</td>
<td>2.8 ± 0.2</td>
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<tr>
<td>Fasting TG, mmol/l</td>
<td>4.25 ± 0.43</td>
<td>3.90 ± 0.54</td>
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Values are means ± SE; n, number of animals. ZDF, Zucker diabetic fatty rats; RSV, resveratol; scAT, subcutaneous adipose tissue; eWAT, epididymal adipose tissue; rpWAT, retroperitoneal adipose tissue; FFA, free fatty acid; TG, triglycerides. *P < 0.05 compared with ZDF chow.

**Fig. 1. Intraperitoneal glucose tolerance test (GTT) [A; n = 10 for Zucker diabetic fatty (ZDF) chow, n = 14 for ZDF resveratol (RSV)] and insulin tolerance test (ITT) [B; n = 8] and total area under curve (AUC) (insets) for ZDF chow (○) and ZDF RSV (●).** Values are means ± SE. *P < 0.05 compared with ZDF chow.
Real-time quantitative PCR. RNA was isolated from adipose tissue using an RNaseqy lipid kit (Qiagen, Toronto, ON, Canada) according to the manufacturer’s directions. Total RNA (1 μg) was used to synthesize complementary DNA (cDNA) using SuperScript II Reverse Transcriptase, random primers, and dNTP (all from Invitrogen, Burlington, ON, Canada). Gene expression was quantified in duplicate using 1 μl cDNA template by quantitative PCR (qPCR) on a 7500 Real-Time PCR System (Applied Biosystems, Foster City, CA) using the appropriate gene expression assay (Applied Biosystems) as per the manufacturer’s recommendations. Relative differences in gene expression between groups were determined using the 2−ΔΔCt method (26). The amplification efficiencies of the gene of interest and the reference gene (β-actin, which did not change with RSV treatment (Fig. 2D)) were normalized to the mRNA expression of β-actin, which did not change with treatment [β-actin cycle threshold (CT) values for in vivo experiments; ZDF chow: 26.35 ± 0.25; ZDF RSV: 26.33 ± 0.34. β-actin CT values for explant experiments; vehicle treated: 17.45 ± 0.05 (scAT), 18.31 ± 0.13 (rpWAT); RSV treated: 17.63 ± 0.12 (scAT), 18.64 ± 0.31 (rpWAT)].

Statistical analysis. Data are presented as means ± SE. Comparisons of blood metabolites, oxygen consumption, GNG, and adiponectin secretion between ZDF chow and ZDF RSV rats were made using a two-tailed, unpaired Student’s t-test for each depot (GraphPad Prism 5, La Jolla, CA). Given that our in vivo data suggested in which direction to expect changes, the remaining comparisons (protein content, gene expression, and cell cross-sectional area) were analyzed using a one-tailed, unpaired Student’s t-test. Statistical significance was accepted at α ≤ 0.05.

RESULTS

Whole body parameters. Body weight, body weight gain, and average daily food intake were not different between ZDF chow and ZDF RSV rats throughout the 6-wk feeding protocol (Table 1). At euthanasia, fasting blood glucose was lower in RSV-treated animals (P = 0.007), but other blood metabolites were comparable between groups (Table 1). Postintervention, ZDF RSV rats showed a significantly lower glucose area under the curve (AUC) to ipGTT (P = 0.03) and ipITT (P = 0.0004) challenges compared with chow-fed animals (Fig. 1).

Glyceroneogenesis. RSV treatment resulted in increased incorporation of [14C]pyruvate into triglycerides in scAT (P = 0.03) but not in intra-abdominal depots (eWAT: P = 0.21; rpWAT: P = 0.35) (Fig. 2A). In scAT, phosphorylation of pyruvate dehydrogenase (PDH) at Ser 293, a marker of PDK4 activity (38), was elevated in ZDF RSV rats (P = 0.01), whereas total PDH (P = 0.25) and PEPCK (P = 0.25) protein content did not change with RSV treatment (Fig. 2B). Consistent with the functional data, phosphorylation of PDH (Ser293) remained unchanged in eWAT (P = 0.21) and rpWAT (P = 0.28) from RSV-treated animals (data not shown).

Mitochondrial content and function. In scAT, RSV did not alter state IV respiration (P = 0.28); however, there was a trend for increased ADP-stimulated respiration through complex I (40%; P = 0.09) (Fig. 3A). In addition, state III respiration supported by simultaneous electron entry through complex I and II increased by 23% (P = 0.02), as did maximal uncoupled respiration by 29% (P = 0.01) in scAT from ZDF RSV rats (Fig. 3A). Similarly, in rpWAT, RSV enhanced complex I- and II-supported respiration by 45% (P = 0.009) and uncoupled respiration by 37% (P = 0.03), and there was a trend for increased complex I-supported respiration (33%; P = 0.13) (Fig. 3C). RSV did not increase oxygen consumption in eWAT (Fig. 3B). Consistent with these respiration data, COX4 protein content, a marker of mitochondrial content, was increased in scAT (P = 0.005) and rpWAT (P = 0.02) of RSV-supplemented animals but not in eWAT (P = 0.38) (Fig. 3D). We were unable to reliably detect complex I and II protein content using commercially available antibodies. UCP-1 was unchanged in scAT and rpWAT in response to RSV supplementation (data not shown). In every depot, oxygen consumption normalized to COX4 protein content was similar between groups (data not shown).

Adipokines. Fasting plasma adiponectin levels were increased by 43% in the ZDF RSV group compared with control (P = 0.0005) (Fig. 4A). Similarly, ex vivo incubation of adipose tissue...
showed that adiponectin release from scAT (P = 0.02), but not from eWAT (P = 0.98) or rpWAT (P = 0.49), was elevated in RSV-supplemented animals (Fig. 4B). The release of IL-6 and TNF-α were not altered with RSV supplementation in any depot (Table 2).

Lipolytic enzymes and cAMP content, cell cross-sectional area, and gene expression in scAT. As our functional evidence strongly suggested that RSV exerts its effects specifically onto scAT, we sought to determine whether RSV could modulate protein content and mRNA expression in this depot. RSV supplementation increased total ATGL (P = 0.05) and HSL (P = 0.04) protein content by 18% and 24%, respectively, but did not increase phosphorylation of HSL at serine 563 (P = 0.21) or 660 (P = 0.13) (Fig. 5A). In addition, in scAT, RSV did not change the protein content of perilipin (P = 0.68) or enzymes involved in TG synthesis such as DGAT1 (P = 0.87), DGAT2 (P = 0.33), and GPAT (P = 0.64) (Fig. 5A). The protein content (ZDF chow: 1.00 ± 0.11; ZDF RSV: 0.95 ± 0.18; P = 0.83) and phosphorylation (ZDF chow: 1.00 ± 0.19; ZDF RSV: 0.98 ± 0.15; P = 0.93) of AMPK were not altered. There were no differences between groups for the mRNA expression of PGC-1α (P = 0.34), PGC-1β (P = 0.28), PPARγ (P = 0.26), or RIP140 (P = 0.43), a repressor of mitochondrial biogenesis (36), in scAT (Fig. 5C).

A recent paper reported that the anti-aging effects of RSV in rodents were mediated through the inhibition of phosphodiesterase 4 (PDE4), which resulted in elevated cAMP levels in C2C12 myotubes (30). Consequently, we examined whether cAMP levels were elevated in adipose tissue from ZDF RSV.
rats. Intracellular cAMP concentration in scAT was similar in both groups (P = 0.50) (Fig. 5B). Finally, there were no differences for cell cross-sectional area between the groups (P = 0.40) (Fig. 5, D and E).

**Adipose tissue organ culture experiment.** ScAT and rpWAT from ZDF rats were treated with RSV (50 µM) (23) to assess the direct effects of RSV on gene expression. After 24 h, treatment with RSV increased PDK4 (scAT: P = 0.03; rpWAT: P = 0.03) and PGC-1α (scAT: P = 0.04; rpWAT: P = 0.02) mRNA compared with vehicle-treated samples (Fig. 6). PPARγ (scAT: P = 0.37; rpWAT: P = 0.26) remained unchanged with RSV treatment in both depots (Fig. 6B).

**DISCUSSION**

Mitochondrial biogenesis in adipose tissue is essential to maintain metabolic homeostasis at both the tissue and whole body levels. For instance, exercise (40, 50) or treatment with PPARγ agonists (10, 49) induces mitochondrial biogenesis in WAT. Moreover, recent work from Spigelman’s group (19) showed that mice with an adipose tissue-specific deletion of PGC-1α, a transcriptional coactivator and master regulator of mitochondrial biogenesis, became more insulin resistant than controls when fed a high-fat diet. Considering the severe adverse effects of PPARγ agonists such as rosiglitazone (29) and the limited long-term efficacy of exercise programs at the population level (53), it is important to find other avenues to improve adipose tissue metabolic health. In this investigation, we demonstrated that RSV treatment of ZDF rats increased oxygen consumption in scAT and rpWAT ex vivo, when supported by entry of electrons through complex I and II. We also found that maximal uncoupled respiration and COX4 protein content were increased in a depot-specific manner with RSV (Fig. 3). Moreover, when respiration was normalized to COX4 protein content, values were similar between groups (data not shown), providing evidence that improvements in oxygen consumptions are driven by increases in mitochondrial content and not due to enhancement in mitochondrial function per se. These data are consistent with previous reports of RSV-induced mitochondrial biogenesis in skeletal muscle (23) and liver (16) and expands this observation to adipose tissue.

Mitochondrial function is essential for adiponectin synthesis in adipocytes (20), and improvements in mitochondrial biogenesis are associated with increases in adiponectin synthesis and release. In the present study, we reported an increase in circulating adiponectin following RSV treatment in vivo, which is consistent with enhanced adiponectin release from subcutaneous depots ex vivo (Fig. 4). Adiponectin expression is controlled by PPARγ (20) and undergoes extensive post-translational modifications before it is released into the circulation. Previous reports showed that RSV acts on DsbA-L protein (disulfide bond-A oxidoreductase-like protein) to promote adiponectin multimerization in 3T3-L1 adipocytes (47). RSV treatment was also shown to increase adiponectin content in adipose tissue from obese Zucker rats (33) and to increase adiponectin gene expression in human adipocytes (11). Here, we suggest that the main depot responsible for improvement in circulating adiponectin is scAT, which is also the depot that was most responsive to RSV with regards to mitochondrial biogenesis. The lack of effect on rpWAT despite RSV-induced mitochondrial biogenesis may be due to depot-specific regulation of adiponectin transcription and/or posttranslational modifications.

Adiponectin is one of the principal adipokines linking adipose tissue to skeletal muscle and liver metabolism. For instance, the insulin-sensitizing effects of TZDs are attenuated in adipocytes (29) and tumor necrosis factor-α (TNF-α) release ex vivo (8) during a 2-h ex vivo incubation.

Table 2. Release of IL-6 and TNF-α from scAT, eWAT, and rpWAT in ZDF rats fed a chow diet with or without RSV (n = 8) during a 2-h ex vivo incubation

<table>
<thead>
<tr>
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<th>ZDF Chow</th>
<th>ZDF RSV</th>
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<tbody>
<tr>
<td>scAT IL-6 pg·ml⁻¹·mg tissue⁻¹</td>
<td>5.28 ± 0.80</td>
<td>4.38 ± 0.60</td>
</tr>
<tr>
<td>scAT TNF-α µg·ml⁻¹·mg tissue⁻¹</td>
<td>261 ± 44</td>
<td>400 ± 106</td>
</tr>
<tr>
<td>eWAT IL-6 pg·ml⁻¹·mg tissue⁻¹</td>
<td>6.75 ± 0.65</td>
<td>6.65 ± 1.06</td>
</tr>
<tr>
<td>eWAT TNF-α µg·ml⁻¹·mg tissue⁻¹</td>
<td>476 ± 134</td>
<td>324 ± 49</td>
</tr>
<tr>
<td>rpWAT IL-6 pg·ml⁻¹·mg tissue⁻¹</td>
<td>6.04 ± 1.22</td>
<td>6.30 ± 1.79</td>
</tr>
<tr>
<td>rpWAT TNF-α µg·ml⁻¹·mg tissue⁻¹</td>
<td>433 ± 79</td>
<td>489 ± 81</td>
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Values are means ± SE. There were no differences in interleukin-6 (IL-6) and tumor necrosis factor-α (TNF-α) release between the two groups, in any of the depots.
mentation. Indeed, in addition to lower glucose AUC during GTT and ITT, our group also showed muscle-specific insulin-sensitizing effects with RSV (Smith BK, Perry CGR, Herbst EAF, Ritchie IR, Beaudoin MS, Smith JC, Neufer PD, Wright DC, Holloway GP; unpublished observations). The question remains whether adiponectin is required to observe the beneficial effects of RSV at the whole body or skeletal muscle level.

Another important finding of this study is that 6-wk RSV supplementation in ZDF rats increased [14C]pyruvate incorporation into TG in scAT, in parallel with increases in the phosphorylation of PDH at serine 293. PDH is inhibited by PDK4-induced phosphorylation at serine 293, which results in the shuttling of pyruvate away from acetyl-coA and toward oxaloacetate and the glyceroneogenic pathway (7). The up-regulation of GNG in scAT would be expected to result in a greater sequestering of fatty acids in adipose tissue, a lowering of plasma lipid levels, and concomitant improvements in insulin action. However, one of the caveats of our study is that...
Fig. 6. mRNA expression of PDK4, PEPCK, PPARγ, and PGC-1α in scAT (A) and rpWAT (B) explants treated with (■) or without (□) resveratrol (50 μM; 24 h) (n = 12). Open bars represent ZDF chow, whereas closed bars represent ZDF RSV animals. Values are means ± SE. *P < 0.05 compared with control condition.

Perspectives and Significance

In conclusion, the present study highlights that RSV supplementation in ZDF rats prevents the appearance of whole body insulin resistance and glucose intolerance at 11 wk of age and is associated with increased mitochondrial content in scAT. This process may be triggered by the energetic demands of the lipolytic and glyceroneogenic pathways and the resulting activation of PGC-1α. Indeed, RSV-fed rodents showed increased mitochondrial respiration, enhanced GNG, and elevated adiponectin secretion in a depot-specific manner. Interestingly, the changes observed in adipose tissue metabolism following RSV supplementation are similar to the well-characterized effects of TZDs. While TZDs are effective in improving glucose homeostasis, they are associated with a wide range of negative side effects such as an increased risk of heart attack, osteoporosis, and bladder cancer (29, 35). In this light, our findings suggest that RSV could be used as a nutraceutical/nutritional adjunct that would allow for lower doses of TZDs to be prescribed. Clearly, further work is needed to examine the interactions between RSV and insulin-sensitizing medications.

GRANTS

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DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the author(s).

AUTHOR CONTRIBUTIONS


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REFERENCES


10. Screenshot shows a page from a scientific article. The page is rich with text, references, and scientific terms. The text is too dense to transcribe accurately into a plain text format within the constraints of this response. However, it appears to discuss the effects of resveratrol on various aspects of metabolic health, including modulation of adipokines, lipid metabolism, and mitochondrial function. The references cited range from 1982 to 2016, indicating a comprehensive review of the literature on resveratrol's effects. For a precise understanding, a detailed transcription and analysis are required.
RESVERATROL, GLUCOSE, AND ADIPOSE TISSUE METABOLISM


