Unilateral renal denervation improves autonomic balance in conscious rabbits with chronic heart failure

Alicia M. Schiller, Karla K. V. Haack, Peter R. Pellegrino, Pamela L. Curry, and Irving H. Zucker

Department of Cellular and Integrative Physiology, The University of Nebraska Medical Center, Omaha, Nebraska

Submitted 30 May 2013; accepted in final form 28 August 2013

Unilateral renal denervation improves autonomic balance in conscious rabbits with chronic heart failure. Am J Physiol Regul Integr Comp Physiol 305: R886–R892, 2013. First published September 4, 2013; doi:10.1152/ajpregu.00269.2013.—A hallmark of chronic heart failure (CHF) is an increased sympathetic tone resulting in autonomic imbalance. Renal denervation (DNx) in CHF patients has resulted in symptomatic improvement, but the protective mechanisms remain unclear. We hypothesized in CHF, unilateral renal DNx would improve cardiac autonomic balance. The present study used conscious, chronically instrumented New Zealand White rabbits undergoing renal DNx prior to pacing-induced CHF. Four treatment groups were used: nonpace, non-DNx [Sham-Innervated (Sham-INV)], nonpace DNx (sham-DNx), pace non-DNx (CHF-INV) or pace DNx (CHF-DNx). We examined several markers indicative of autonomic balance. Baroreflex sensitivity and time domain heart rate variability (HRV) were both decreased in the CHF-INV group compared with sham-INV and were restored to sham levels by renal DNx. Power spectral analysis indicated an increase in low-frequency/high-frequency (LF/HF) ratio in the CHF-INV compared with the sham-INV, which was normalized to sham levels by DNx. To assess whether this was due to a withdrawal of sympathetic tone or an increase in parasympathetic tone, the heart rate response was measured after an intravenous bolus of metoprolol or atropine. Bradycardia induced by intravenous metoprolol (indicative of cardiac sympathetic tone) was exacerbated in CHF-DNx rabbits compared with sham-DNx but was normalized in CHF-DNx. Conversely, the tachycardia in response to intravenous atropine (indicative of cardiac vagal tone) was not improved in CHF-DNx vs. CHF-INV animals. Renal DNx also prevented the increase in circulating plasma NE seen in CHF-DNx rabbits. These results suggest renal DNx improves cardiac autonomic balance in CHF by a reduction of sympathetic tone.

Renal denervation can greatly influence renal function because the kidney is innervated by both effenter and afferent nerve fibers (8, 3). Activation of the renal efferent nerve fibers results in an increase in sodium reabsorption, renin secretion, and decreased renal blood flow and, therefore, may influence glomerular filtration rate (9). Activation of the renal afferent nerves has been implicated in the modulation of hypothalamic activity and the reflex influence of sympathetic outflow to organs, such as the heart and kidney (33). These mechanisms may be beneficial homeostatic measures during an initial decrease in cardiac output. However, a chronic decrease in cardiac output (such as in CHF) can lead to a vicious cycle, in which tonic increases in sympathetic outflow contribute to a state of autonomic imbalance and progression of the disease. Markers of autonomic imbalance, such as a depressed baroreflex function, reduction of heart rate variability (HRV), and elevated plasma NE have been observed (6, 11, 30) in CHF and other diseases with excessive sympathetic tone such as hypertension.

Previous studies from our laboratory have shown that there is a decrease in renal blood flow and an increase in ANG II type 1 receptor expression in the renal cortex of rabbits with CHF. Unilateral renal DNx effectively restored both of these parameters to normal in the DNx kidney (5). Therefore, we utilized this model to examine whether unilateral renal DNx evoked global changes in sympathetic tone and autonomic imbalance. On the basis of previous studies involving bilateral renal DNx in hypertension and the high level of sympathetic tone present in CHF (30), we hypothesized that renal DNx would improve autonomic balance in CHF by a withdrawal of excessive sympathetic tone. We examined sham and CHF rabbits to determine whether unilateral renal DNx impacts autonomic function and balance.

METHODS

Animals. Experiments were carried out on 19 male New Zealand White rabbits ranging in weight from 3.7 to 3.9 kg (Charles River Laboratories, Wilmington, MA). All experiments were reviewed and approved by the University of Nebraska Medical Center Institutional Animal Care and Use Committee.

Animals underwent two separate surgeries and were included in two experimental groups when possible, as indicated in the experimental timeline shown in Fig. 1.

Surgical procedures. The model used to induce CHF and for renal DNx has been used previously in our laboratory (5). In brief, rabbits were instrumented with a radiotelemetry device placed in the right femoral artery (model TA11PA-C40; Data Sciences International, Minneapolis, MN) to measure pulsatile and mean arterial pressure (MAP) and heart rate (HR) in the conscious state. At the same time, a left thoracotomy was performed and a platinum wire-pacing electrode was placed on the left ventricle of the heart and a ground wire secured to the left atrial appendage. The chest was evacuated and closed. The pacing wires were tunneled beneath the skin and exited in

BILATERAL RENAL DENERVATION (DNx) has recently entered clinical practice as a successful therapy for drug-resistant hypertension (35). This treatment, using a radiofrequency catheter to ablate the renal nerves, chronically lowers arterial pressure while producing virtually no short-term side effects (2, 14). It is well established that most forms of hypertension are correlated with an overactive sympathetic nervous system, as indicated by elevated plasma norepinephrine (NE) or increases in muscle sympathetic nerve activity and increases in whole body NE spillover (27). There is anecdotal and recent evidence to suggest that renal DNx can reduce excessive sympathetic tone in hypertension (34, 17) in some cases. However, recent studies by Brinkmann et al. (4) and Hart et al. (16) suggest that this may not always be true. Other cardiovascular diseases, such as chronic heart failure (CHF), have a hallmark of increased sympathetic tone (27).
Surgery 2

for a minimum of 2 wk before any experiments were performed. The length of the artery and vein. The hilus of the kidney was cleared of microscopy, we carefully removed all visible renal nerves from the identified, and the renal artery and vein were isolated. Using surgical the midscapular region. The thoracotomy was closed in layers. No chest surgery from the renal denervation surgery. Experiments were then conducted but no denervation was performed, leaving the renal nerves intact; Dnx, denervation; HR, heart rate; BP, blood pressure. *

Fig. 1. Representative time line of the study design. Two weeks separated the chest surgery from the renal denervation surgery. Experiments were then carried out in the prepared or sham state. Following echo-based determination of the chronic heart failure (CHF) state, a similar set of experiments were carried out. Animals were euthanized at the conclusion of the experiments. the midscapular region. The thoracotomy was closed in layers. No earlier than 2 wk after this initial surgery, the renal DNX maneuver was performed. Through a left flank incision, the left kidney was identified, and the renal artery and vein were isolated. Using surgical microscopy, we carefully removed all visible renal nerves from the length of the artery and vein. The hilus of the kidney was cleared of fat and any visible nerve fibers. The rabbits were allowed to recover for a minimum of 2 wk before any experiments were performed.

Induction of CHF. CHF was induced by rapid ventricular pacing as previously described and performed in our laboratory (5, 22). Briefly, echocardiographic measurements (Acuson Sequoia 512 C; 4-MHz probe) were taken in conscious, quietly resting rabbits the day before the pacing protocol was initiated. Cardiac dimensions, volume, fractional shortening (FS), and ejection fraction (EF) were measured and calculated using standard formulas from the parasternal short axis view in the M mode configuration. A small external pacemaker of our own design was then attached to the exposed ventricular pacing wires and the rabbit fitted with a jacket to protect the unit and wires. The rabbits were paced at ~380 bpm for 1–2 wk, and then echocardiographic measurements were repeated on a weekly basis until an EF of <45% was observed. At this time, the postpace (i.e., CHF) experiments were started. The pacing protocol was the same for innervated (INV) and DNX animals. At the conclusion of the study, rabbits were euthanized with an overdose of pentobarbital sodium. The kidneys were quickly removed, placed on dry ice, and stored at ~80°C for future biochemical analysis.

Animals were randomly divided into two primary experimental groups: a sham-operated group (Sham-INV), in which radiotelemetry and left ventricular pacing leads were placed and in which the kidney was manipulated but no denervation was performed, leaving the renal nerves intact. The second group also had radiotelemetry and pacing leads placed on the left ventricle, but the renal nerves were removed (Sham-DNX). After the prepace (Sham) experiments were performed, the animals were then subjected to rapid ventricular pacing to produce the CHF-INV and CHF-DNX groups (Fig. 1). Although most animals were able to complete the entire study and be included in Sham and CHF groups, a small subset of animals were unable to pace continuously or encountered other technical difficulties making paired analysis not possible.

Baroreflex control of HR. During all experiments, the protocols were performed in an identical fashion for all groups with the exception that the CHF groups had their external pacemakers turned off for ~20 min before the start of the experiment. For the baroreflex protocol, rabbits were placed inside of a Plexiglas box in a quiet, dimly lit laboratory, and an intravenous catheter was placed in a marginal ear vein. The animals were allowed a 15-min acclimation period before placement of the intravenous catheter. Baseline measurements of HR and MAP were recorded from the radiotelemetry unit during this time. The last 5 min of this recording was used to calculate baseline values. Baroreflex function was assessed by alterations in arterial pressure following an intravenous infusion of sodium nitroprusside (100 μg/kg) at a rate of 0.5 ml/min until MAP reached ~50 mmHg (~3–4 min). An infusion of phenylephrine (80 μg/kg) was started at the same rate and continued until the MAP reached ~100 mmHg (~4 min).

Arterial baroreflex curves were constructed by taking an average HR every 5 s from the lowest MAP to the highest MAP achieved in each experiment. The data were fit to a logistic regression, as previously described by Kent et al. (21) and used by our laboratory in previous studies (15, 20). The following equation was used to fit the data: HR = A/[1 + exp(B(MAP − C))] + D, where A is the HR range, B is the slope coefficient, C is the pressure at the midpoint of the range (BP50), D is the minimum HR, and MAP is mean arterial pressure. The peak slope (or maximum gain) was determined by taking the first derivative of the baroreflex curve as described by Slope = A × B × exp[(B(MAP − C))]2/(1 + exp[B(MAP − C)]2).

The mean values (see Table 1 and Fig. 4A) of each parameter generated from individual curves in each group of rabbits were used to derive the composite curves presented in Fig. 3, B and C.

Table 1. Parameters used to generate composite baroreflex curves in Fig. 3, B and C

<table>
<thead>
<tr>
<th></th>
<th>Min HR, bpm</th>
<th>BP50, mmHg</th>
<th>Slope coefficient, l/(s·slope)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sham-INV</td>
<td>183.99 ± 19.9</td>
<td>76.40 ± 2.5</td>
<td>0.12 ± 0.01</td>
</tr>
<tr>
<td>CHF-INV</td>
<td>247.88 ± 15.1*</td>
<td>75.70 ± 7.9</td>
<td>0.11 ± 0.03</td>
</tr>
<tr>
<td>Sham-DNX</td>
<td>115.24 ± 29.7*</td>
<td>67.24 ± 6.0</td>
<td>0.06 ± 0.01*</td>
</tr>
<tr>
<td>CHF-DNX</td>
<td>39.52 ± 24.6†</td>
<td>93.01 ± 11.0†</td>
<td>0.05 ± 0.01*</td>
</tr>
</tbody>
</table>

Values are expressed as means ± SE. CHF, chronic heart failure; INV, innervated; Dnx, denervation; HR, heart rate; BP, blood pressure. *P < 0.05 vs. Sham-INV, †P < 0.05 vs. CHF-INV, n = 5–7/group.
Table 2. Baseline hemodynamic parameters in Sham-INV, CHF-INV, Sham-DNx, and CHF-DNx rabbits

<table>
<thead>
<tr>
<th></th>
<th>Sham-INV</th>
<th>CHF-INV</th>
<th>Sham-DNx</th>
<th>CHF-DNx</th>
</tr>
</thead>
<tbody>
<tr>
<td>Body weight, kg</td>
<td>3.7 ± 0.1</td>
<td>3.8 ± 0.2</td>
<td>3.9 ± 0.1</td>
<td>3.9 ± 0.1</td>
</tr>
<tr>
<td>HR, beats/min</td>
<td>216 ± 10</td>
<td>258 ± 11*</td>
<td>196 ± 3.0</td>
<td>247 ± 5.0*</td>
</tr>
<tr>
<td>MAP, mmHg</td>
<td>75.4 ± 5.3</td>
<td>65.5 ± 1.9*</td>
<td>17.7 ± 0.5</td>
<td>15.9 ± 0.5</td>
</tr>
<tr>
<td>LVEDD, mm</td>
<td>16.8 ± 0.3</td>
<td>17.7 ± 0.5</td>
<td>15.9 ± 0.5</td>
<td>18.2 ± 0.4</td>
</tr>
<tr>
<td>LVESD, mm</td>
<td>11.6 ± 0.3</td>
<td>14.2 ± 0.6*</td>
<td>8.6 ± 2.0</td>
<td>17.7 ± 0.5*</td>
</tr>
<tr>
<td>LVD Vol, ml</td>
<td>8.2 ± 0.5</td>
<td>9.5 ± 0.7</td>
<td>8.2 ± 0.5</td>
<td>9.9 ± 0.5</td>
</tr>
<tr>
<td>LVsVol, ml</td>
<td>3.22 ± 0.1</td>
<td>5.4 ± 0.6*</td>
<td>3.1 ± 0.2</td>
<td>6.1 ± 0.3*</td>
</tr>
<tr>
<td>FS, %</td>
<td>31.1 ± 1.1</td>
<td>19.8 ± 1.2*</td>
<td>32.0 ± 1.4</td>
<td>17.4 ± 1.2*</td>
</tr>
<tr>
<td>Ejection fraction, %</td>
<td>62.4 ± 1.0</td>
<td>38.9 ± 1*</td>
<td>62.4 ± 1.0</td>
<td>43.5 ± 2.0*</td>
</tr>
</tbody>
</table>

Values are expressed as means ± SE. MAP, mean arterial pressure; LVD, left ventricle; LVEDD, left ventricular end-diastolic diameter; LVESD, left ventricular end-systolic diameter; LVD, left ventricle diastolic; LVs, left ventricle, systolic; FS, fractional shortening. *P < 0.05 vs. respective sham group. n = 7/group.

for NE. To normalize the NE content to protein concentration, a protein assay kit (Pierce, Rockford, IL) was used according to the manufacturer’s directions. At the end of the study, NE tissue levels were compared between the left (DNx) and right (INV) kidney in three rabbits.

During the course of both sham and CHF experimental periods, blood samples were obtained by placing a catheter in a central ear artery. Approximately 3 ml of whole blood was collected and immediately placed on ice in a heparinized collection tube. No more than 30 min later, the blood was centrifuged at 1,000 g for 15 min. The plasma was collected and stored at −80°C. Plasma NE levels were measured using the same ELISA procedure as described above.

Statistical analysis. Data are expressed as the means ± SE. All HR and MAP data were obtained with the use of a 16-channel Powerlab (model 16 SP; ADInstruments). Differences between groups were determined with a one-way ANOVA followed by a post hoc analysis (model 16 SP; ADInstruments). Differences between groups were determined with a one-way ANOVA followed by a post hoc analysis (model 16 SP; ADInstruments). Differences between groups were determined with a one-way ANOVA followed by a post hoc analysis (model 16 SP; ADInstruments).

RESULTS

Baseline hemodynamics. One to two weeks after rapid ventricular pacing, rabbits exhibited signs of impaired cardiac function, as validated by echocardiography. Left ventricular end-systolic dimensions and left ventricular end-systolic volume were increased in both CHF groups compared with their respective Sham group. EF and FS were also decreased in all CHF animals compared with their respective Sham group (Table 2). As reported previously in this model of CHF, the rapid ventricular pacing protocol produces an increased HR and decreased MAP in CHF compared with Sham rabbits (5, 20). Renal DNx did not alter any of the baseline hemodynamic parameters.

Renal cortical NE content. The renal DNx surgical maneuver performed in our laboratory has previously been validated to be an effective means of removing the renal nerves (5) by showing a significant decrease in renal cortical NE levels in the DNx kidney compared with the contralateral innervated side.

In the current study, we found a pronounced decrease in renal cortical NE content of the left/DNx kidney compared with the contralateral kidney in the same animals, indicating complete and prolonged renal DNx (Fig. 2A).

Circulating plasma NE levels. To evaluate whether unilateral renal DNx could reduce global sympathetic tone, circulating plasma NE was measured (Fig. 2B). As expected, the CHF-INV group showed an increase in plasma NE compared with Sham-INV rabbits. Renal DNx prevented the increase in plasma NE in CHF rabbits.

Arterial baroreflex function. We evaluated baroreflex function as it is known to be depressed in CHF and other states of autonomic imbalance (11, 27, 30). Representative tracings from arterial baroreflex experiments are shown in Fig. 3A. Composite arterial baroreflex curves for HR and MAP are shown in Fig. 3B. Interestingly, baroreflex function was enhanced by renal DNx treatment in both treatment groups (i.e., sham and CHF states). The Sham-DNx group (left) exhibited an increased baroreflex function compared with the Sham-INV. The depressed baroreflex function observed in the CHF-INV group was enhanced in the CHF-DNx group (right). Mean data for HR range and maximal gain are shown in Fig. 4. Renal DNx increased baroreflex HR range in the Sham-DNx group compared with Sham-INV (Fig. 4A). As expected, the CHF-INV group exhibited a decrease in HR range compared with Sham-INV; however, this was restored to Sham levels in the CHF-DNx group. Similar results were observed for the maximal baroreflex gain (peak slope; Fig. 4B), indicating that renal DNx enhances baroreflex function and abolishes the blunted baroreflex response normally observed in CHF-INV animals.
Heart rate variability. Time and frequency domain of HRV was used as a reflection of autonomic balance in the heart. Therefore, we examined the effects of renal DNx on this parameter in animals with and without CHF. Fig. 5A shows representative Poincaré plots visualizing beat-to-beat HRV generated from ECG recordings from all groups of rabbits, and Fig. 5B shows summary HRV (SDNN). In CHF-INV rabbits, there was a profound depression in HRV compared with Sham-INV. However, in the CHF-DNx group, HRV was restored to sham levels. Renal DNx in the sham group did not affect HRV. Power spectral analysis was also performed (Fig. 5C) to examine any changes in the frequency domain of HRV. Frequency domain analysis indicated an increase in the LF/HF power spectral density ratio in the CHF-INV group compared with both sham groups. This increase in ratio was prevented in the CHF-DNx group. While the total power of the HRV spectrum was not different between groups by one-way ANOVA ($P = 0.11$), there was a trend toward lower total power in the CHF-INV group (36.8 ± 11.5 ms²) compared with both their CHF-DNx counterparts (219.3 ± 51.1 ms²), as well as both the Sham-INV (273.8 ± 65.0 ms²) and Sham-DNx (381.8 ± 104.8 ms²) groups.

Vagal and sympathetic components of resting heart rate. To further investigate whether the changes observed in baroreflex function and HRV were due to a withdrawal of sympathetic tone, an increase in parasympathetic tone or both, we evaluated the change in resting HR in response to autonomic blockade (Fig. 6). In CHF-INV animals, the bradycardia in response to
metoprolol was enhanced compared with the Sham-INV group (Fig. 6A). However, this exaggerated response was attenuated in the CHF-DNx animals and restored to the response seen in the Sham-INV group. There were no differences between the Sham-INV and Sham-DNx groups. When the same protocol was performed following atropine (Fig. 6B), we observed a decreased tachycardia in the CHF-INV compared with Sham-INV group. This same trend was observed in the CHF-DNx groups, indicating no differences in the vagal contributions to resting HR in the CHF-DNx group.

DISCUSSION

The results of the present study show that unilateral renal DNx is sufficient to reduce local and global sympathetic outflow (Fig. 2), as evident by a reduction in both renal cortical NE levels and plasma NE levels and improved autonomic balance (Figs. 3–5) in CHF. Our data suggest that renal DNx evokes these protective effects mainly by mediating a reduction in sympathetic tone (Fig. 6).

Over the past several years, a growing body of literature has focused on the use of renal DNx as a therapy for drug-resistant hypertension (12, 23, 28, 32). These studies point to a profound and prolonged reduction in arterial pressure, and there is also some evidence to suggest a reduction in sympathetic outflow (31). Because increased sympathoexcitation characterizes both hypertension and CHF, the efficacy of this procedure in the setting of CHF has begun to be evaluated more extensively. Recent studies have shown benefit for using bilateral renal DNx in both animal and patient models of CHF (7, 19). The REACH pilot study provides evidence that in patients with chronic systolic heart failure, bilateral renal DNx increases exercise tolerance and provides patients with symptomatic improvement. Surprisingly, MAP in these CHF-DNx patients was maintained and did not decrease over a period of 6 mo (7). Given, however, that previous studies in hypertensive patients indicated that bilateral renal DNx continually lowered arterial pressure for up to 24 mo, additional studies will have to be performed in the CHF cohort to confirm that blood pressure is

---

**Fig. 5.** A: representative Poincaré plots showing the relationship between the R-R interval and the next R-R interval for resting HR in each of the four groups studied. B: time domain of heart rate variability (HRV) representing the standard deviation of all NN intervals. C: frequency domain of HRV representing the PSD ratio of low frequency/high frequency (LF/HF). Values in B and C are expressed as means ± SE. *P < 0.05 vs. Sham-INV. †P < 0.05 vs. CHF-INV. n = 5–7.

---

**Fig. 6.** Sympathetic and vagal components of resting HR. Change in HR in quietly resting rabbits after an intravenous bolus of metoprolol (A) or atropine (B). Values are expressed as means ± SE. *P < 0.05 vs. Sham-INV. †P < 0.05 vs. CHF-INV. n = 5–7.
maintained for a longer time frame post-renal DNx (34). Previous studies from our laboratory have shown that in CHF, there is an increased level of ANG type 1 receptor and a deceased level of ANG type 2 receptor in the renal cortical vasculature of CHF rabbits (5). Unilateral renal DNx was able to locally restore this imbalance in the DNx kidney. Therefore, we investigated whether these local changes produced by unilateral renal DNx were also producing global changes in autonomic balance in CHF. The current study provides evidence that DNx of a single kidney restores autonomic balance and improves baroreflex function; future studies comparing the potential additive effects of unilateral and bilateral renal DNx, as well as unilateral renal DNx, as an alternative to bilateral renal DNx should be explored.

The rapid ventricular pacing protocol resulted in reduced cardiac function validated by a significant decrease in EF by echocardiography. It is of interest that renal DNx in our model did not lead to improvements in cardiac function (Table 1), even though it appears that cardiac and global sympathetic tone is decreased. These results are consistent with previous data from our laboratory using this model (4). The reasons for this are not readily apparent; however, these effects may be model specific, as cardiac pacing was continued throughout the duration of the study by an external stimulus. Therefore, improvements in cardiac function are unlikely to be observed unless the pacing paradigm is discontinued. Secondly, it is possible that the effects of renal DNx on cardiac function take a longer period of time to manifest. This aspect of renal DNx was not evaluated in this study. For future studies, adaptations to our model protocol could be implemented to better observe changes in cardiac function. We are also planning future studies in which renal DNx will be performed after the development of CHF to further increase the clinical relevance. Technical challenges in reanesthetizing sick CHF rabbits to perform renal DNx prevented collection of these data at the current time.

We and others have consistently shown that baroreflex function is impaired in the setting of CHF and may contribute to the sympathoexcitatory state (11, 22, 26). In the current study, the baroreflex HR range and maximal gain were increased in CHF animals following renal DNx. These data point to important differences in baroreflex-evoked changes in HR compared with resting baseline HR measurements, as renal DNx did not influence resting HR values in either the Sham or CHF groups. This may also reflect the fact that animals were continuously paced throughout the duration of the study. In the present study, renal cortical NE measurements taken at the conclusion of the study or ~6 wk after renal DNx show virtually no NE present in the renal cortex, indicating little or no sympathetic innervation and no efferent reinnervation over this period of time (Fig. 2). We did not evaluate the presence of afferent innervation and the conclusion of the study and, therefore, cannot definitively say that afferent function was not restored over this period of time or make conclusions regarding the effect of efferent or afferent input separately.

If some of the effects of renal DNx are mediated by interruption of renal afferent nerves, this may lead to a reflexive decrease in sympathetic outflow, as increases in afferent nerve activity can evoke increases in efferent renal sympathetic nerve activity (10). It is not known whether there are compensatory changes in contralateral efferent renal nerve activity in the time frame studied in our protocol. Removal of afferents from one kidney is likely to affect a sympathetic excitatory reflex that may be more active in the setting of CHF, as has been shown for cardiac sympathetic afferents (37).

We observed an increase in baroreflex function in the Sham-DNx group compared with the Sham-INV group when we used a pharmacological approach to assess baroreflex function (Fig. 3B). Therefore, it is possible that there is a withdrawal in sympathetic tone or an increase in vagal tone in the sham state following DNx. However, changes in resting HR in response to atropine and metoprolol do not support this notion. Further investigation is required to determine whether renal DNx changes autonomic balance under nonpathophysiological conditions.

Perspectives and Significance

The current study shows that three independent measures of sympathovagal balance indicate that unilateral renal DNx prevents cardiac autonomic imbalance and decreased baroreflex function in rabbits with pacing-induced CHF. Autonomic blocker data demonstrate that this is through a sympatholytic effect. Unilateral renal denervation also had no effect on MAP in our model. This study raises the possibility that unilateral renal DNx may be sufficient to observe beneficial therapeutic effects in CHF. This is an important clinical application as a reduction in MAP in CHF, even if accompanied with a reduction in sympathetic tone, would not be a desirable outcome. The clinical implications of this finding suggest that unilateral renal DNx therapy may be effective in reducing both global and local sympathetic tone in CHF patients.

ACKNOWLEDGMENTS

We thank Ms. Johnnie F. Hackley for her excellent technical assistance in performing the echocardiographic measurements.

GRANTS

This work was supported by a National Institutes of Health National Heart, Lung, and Blood Institute grant PO1 HL-62222. A. M. Schiller was partially supported by an American Heart Association predoctoral fellowship, Midwest affiliate.

DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the authors.

AUTHOR CONTRIBUTIONS

Author contributions: A.M.S. and P.L.C. performed experiments; A.M.S. and P.R.P. analyzed data; A.M.S., K.K.V.H., P.R.P., and I.H.Z. interpreted data.

REFERENCES

RENAL DENERVATION AND AUTONOMIC BALANCE IN HEART FAILURE


