The Consequences of Gustatory Deafferentation on Body Mass and Feeding Patterns in the Rat.


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ABSTRACT

The contribution of orosensory signals, especially taste, on body mass, and feeding and drinking patterns in the rat was examined. Gustatory deafferentation was produced by bilateral transection of the chorda tympani, glossopharyngeal, and greater superficial petrosal nerves. Total calories consumed from sweetened milk diet and oil-chow mash by the nerve-transected rats significantly decreased relative to sham-operated controls, mostly attributable to decreases in bout number, but not size. Nevertheless, caloric intake steadily increased over the postsurgical observation period, but body mass remained below both presurgical baseline and control levels and did not significantly increase over this time. After the sweetened milk diet/oil-chow mash phase, rats received a series of sucrose preference tests. Interestingly, the nerve-transected rats preferred sucrose, and intake did not differ from controls, likely due to the stimulus sharing some non-taste chemosensory properties with the sweetened milk diet. The neurotomed rats initiated a greater number of sucrose licking bouts that were smaller in size and slower in licking rate, compared with control rats, and unlike in control rats, the latter two bout parameters did not vary across concentration. Thus, in the absence of gustatory neural input, body mass is more stable compared with the progressive trajectory of weight gain seen in intact rats, and caloric intake initially decreases but recovers. The consequences of gustatory neurotomy on processes that determine meal initiation (bout number) and meal termination (bout size) are not fixed and appear to be influenced by presurgical experience with the food stimulus coupled with its nongustatory chemosensory properties.
Keywords: ageusia; body mass regulation; taste loss; taste nerves
INTRODUCTION

Decades of research have shown that nutritional intake contributes to the development of human disease mainly by influencing the development of obesity and its associated conditions (e.g., cardiovascular disease, hypertension, type 2 diabetes mellitus) [e.g., (8, 28, 35)]. Collectively, these studies suggest that a majority of obesity-related diseases could be prevented by the adoption of healthier eating habits. In the act of feeding, specialized chemosensory, thermosensory, and mechanosensory receptors, positioned in the nasal and oropharyngeal cavities, provide information to the brain related to the macronutrient composition, caloric density, osmolarity, and potential toxicity of food ingested (1, 43, 49, 76). Chief among these neural signals are those provided by the gustatory system, which is the final guardian of the alimentary tract and provides a chemical analysis of stimulus components that do not typically have sufficiently high vapor pressures to effectively stimulate olfactory receptors and do not appreciably stimulate mechanoreceptors in a selective fashion. It is widely believed that these orosensory inputs, especially taste, integrate with other neural and endocrine signals related to the energy and hydromineral status of the animal to ultimately influence central neural circuits controlling food and fluid intake.

Orosensory inputs, particularly signals of gustatory origin, have been implicated in the control of meal size. For example, it is well known that when postingestive inputs to the CNS are substantially reduced, which can be accomplished via the use of sham feeding methodology, meal size increases considerably (e.g., 14, 46, 50). However, it is unknown what would happen to meal size and/or meal number in the absence of gustatory neural input. It is thus unclear whether taste input is necessary for normal
feeding patterns to be observed and for the normal maintenance of body mass. Indeed, if total intake is reduced by experimentally induced ageusia, it remains unclear whether this is due to disruption in processes that terminate meals or those that initiate them or both.

Despite the large body of evidence that indicates taste can influence nutritional intake, research demonstrating a definitive role for the gustatory system in the maintenance of body mass has been sparse (16). Here we sought to determine how combined transection of the chorda tympani nerve (CT), innervating the taste buds of the anterior tongue, the greater superficial petrosal nerve (GSP), innervating the taste buds of the palate, and the glossopharyngeal nerve (GL), innervating the taste buds of the posterior tongue would affect feeding and body mass regulation. These nerve transections effectively remove the input from ~90% of the taste receptor cells, which, in turn, degenerate after nerve transection (e.g., 41, 59). The remaining taste buds are found in the laryngeal epithelium (67), innervated by the superior laryngeal nerve, and are thought to be more involved in the protection of the airways (53). We reasoned that the functional consequences of this massive elimination of gustatory input would allow us to reveal whether taste signals were necessary for the maintenance of normal feeding patterns and body mass regulation.
Methods

Subjects

Twenty-five male Sprague-Dawley rats (Charles River Breeders, Wilmington, MA) with a mean weight of 416.1 ± 6.7 g (± S.E.M.) at the time of surgery, were individually housed in polycarbonate cages in a room with automatically controlled temperature (21.9 – 22.6 °C), humidity (34 – 69%) and a 12 h light/ 12 h dark lighting cycle. All experimental protocols were approved by the Institutional Animal Care and Use Committees at the University of Florida and at the Florida State University.

Surgery

Rats were anesthetized with a mixture of ketamine hydrochloride (125 mg/kg body mass) and xylazine hydrochloride (5 mg/kg body mass) delivered via intramuscular injection. Supplemental doses were administered as necessary. Rats received one of the following surgeries: combined bilateral transection of the CT, GSP, and GL (TRIPLEx); bilateral transection of the CT and exposure of the GSP (CTx); combined bilateral exposure of both the CT and GL (SHAM). The GL was exposed after an incision was made in the ventral neck and the sublingual and submaxillary salivary glands, the omohyoid, sternohyoid, and posterior belly of the digastric muscle were all retracted revealing the nerve running medial to the external wall of the bulla. In animals receiving SHAM surgery, the nerve was left uncut and in the TRIPLEx animals ~8-10 mm of the nerve was removed on both sides. The incision was then closed with nylon suture. In the TRIPLEx rats an incision was also made dorsal to the pinna which
transected the external ear canal which was retracted laterally. The muscles overlying the bone were blunt dissected and the bony meatus was enlarged with a dental drill. The tympanic membrane and the middle ear ossicles were removed as well as a large section of the CT. The tensor tympani muscle was then avulsed exposing a ridge of temporal bone, a small piece of which was removed with microforceps revealing the GSP which was then transected and the ends cauterized. Wound clips were used to close the incision. In animals receiving SHAM surgery, the pinna and ear canal was transected and the muscle surrounding the bony meatus was blunt dissected and retracted and the tympanic membrane was punctured. For animals in the CTx group, only the tympanic membrane, the middle ear ossicles, and a large section of the CT were removed. The inclusion of the CTx group, in addition to serving as an additional surgical trauma control group, helped to control for any putative effects of the loss of parasympathetic efferents to the sublingual and submaxillary glands on body mass. For three days following surgery the animals received daily injections of Penicillin G Procaine suspension (30,000 units sc) and Ketorolac Tromethamine (2 mg/kg body mass sc).

Behavioral procedure

Body mass measures were taken daily in TRIPLEx, SHAM, and CTx rats. TRIPLEx or SHAM rats were placed in specially designed cages described elsewhere (55) with some modifications. These cages allowed continuous monitoring of chow and fluid ingestion in 6-s time bins over 23-h daily test sessions. Powdered laboratory chow (Purina Laboratory Chow 5001, St. Louis, MO) was provided ad libitum in a food jar
placed in the feeding compartment of the cage. When the rat entered the feeding area, its head interrupted an infrared beam and these beam-break signals were recorded by a computer. On the other side of each cage was a stainless steel rack that held two fluid bottles. Licks on the drinking spouts activated an electronic contact circuit which was registered by a computer. The CTx rats were included in the design as a body mass control group and thus were not maintained in the testing apparatus, but had access to the same foods and fluids as the other two groups. Because there were a limited number of cages in the feeding and drinking monitor, all of the animals in the groups could not be tested at the same time, but rather, the testing was conducted in two phases with animals from both the SHAM and TRIPLEx groups included in each phase. All of the rats in the CTx group were tested in the second phase.

Intake of powered food and water (deionized reverse-osmosis) from the two bottles were measured for four days. After this period, oil-chow mash (2 parts Canola corn oil + 5 parts powdered chow) was introduced in place of the powered chow. After one week of experience with the oil-chow mash, water in one of the fluid bottles was replaced with a sweetened milk diet (1 part sweetened condensed milk (Borden’s) + 2 parts water + 1 mL multiple vitamins (Polyvisol)), prepared fresh daily. Both nutritional regimens, the oil-chow and the milk diet, were chosen because they are lubricated and thus their consumption was less likely to be affected by any postsurgical complications associated with potential decreases in salivation. Moreover, these are highly palatable foods and have been shown to be effective at motivating rats to eat, as well as at maintaining the health of nerve-transected rats (e.g., 5, 24, 60, 63). The bottles were rinsed daily and then refilled with fresh solutions. The position of the bottles was
switched every 24 h. On day 20 or 21 of the experiment, rats had surgery. Animals were fed the oil-chow and sweetened milk diet for 44 days postsurgery before being moved back to the powered chow diet and water access for 4 days. Baseline measures were operationally defined as those gathered the last 2 complete presurgical days for each rat.

After day 48, rats were then tested for sucrose preference using a two-bottle test across an ascending series of concentrations of sucrose vs. water in two consecutive 23-h tests per concentration. During this phase, rats also had *ad libitum* access to oil–chow mash. The position of the bottles was switched every 24 h. Sucrose was presented across the days in ascending order of concentration (0.0625, 0.125, 0.25, 0.5 and 1.0 M sucrose; Sigma Aldrich, St. Louis, MO).

**Data analysis**

Licks on the drinking spout and photobeam interruptions at the food jar were measured in consecutive 6-s bins. Thus, in a 23-h period there were 13,800 bins from which to reconstruct the drinking and feeding patterns of each rat with the aid of customized software. A drinking bout (for the sweetened milk diet and sucrose) was defined as starting with three licks in a 6-s bin and ending when fifty 6-s bins (i.e., 5 min) had passed with no licking; a drinking bout had to contain a minimum of 10 licks to be included in the analysis. A feeding bout was defined as starting with 3-s of photobeam interruption in a 6-s bin and ending with fifty 6-s bins (5 min) without beam breaks; a feeding bout had to last 10-s to be included in the analysis. Similar bout criteria have been successfully used in previous studies (2, 11, 19, 55, 68). Estimated fluid spillage,
based on the measurement from bottles placed on empty cages, was subtracted from the intake values. We did not detect any significant spillage of food.

Caloric intake for the oil–chow mash was calculated by multiplying the amount consumed by 4.97 kcal/g. The latter value was based on both the oil (9 kcal/g) and chow (3.36 kcal/g; physiological fuel value) caloric contribution relative to the mixture proportions of the oil–chow mash (i.e., 2 parts oil, 5 parts chow). Caloric intake for the sweetened milk diet was calculated by multiplying adjusted amount consumed by 1.1 kcal/g. This value was derived by adjusting the caloric value of the sweetened condensed milk (3.3 kcal/g) relative to the mixture proportions of the sweetened milk diet (1 part sweetened condensed milk, 2 parts water). For the sweetened milk diet and sucrose, to calculate bout size, we divided total intake by total licks to get an mL/lick value. We then converted each bout size from licks to mLs. Food bout size was calculated by taking total intake and dividing by total beam break time to get a g/s value and then multiplying this value by the beam break time of a given bout. The rate of ingestion during bouts was derived by dividing bout size by bout duration. Only bouts that lasted ≥ 1 min were included in the analysis of bout rate.

Body mass, calories ingested, as well as food intake pattern variables (e.g., bout number, bout size, etc.) were measured during the entire presurgical period and for three weeks postsurgically. Presurgical baselines for the various measures were operationally defined as the mean dependent values two days before surgery for each rat and were compared across groups with the use of t-tests (see Table 3). The effects of surgery were compared by subtracting the postsurgical values for a given variable from the presurgical baseline value for each animal resulting in a change from
presurgical baseline score. This controlled for minor individual differences in the presurgical value for the variables. The mean change from presurgical baseline scores for a given variable were analyzed by performing linear regression for each surgical group over the 21 day postsurgical observation period. This produced a y-intercept, that represented the initial effect of the surgery and slope that represented the change in the scores over postsurgical days. Additionally, the initial impact of surgery was also assessed between the groups through the use of t-tests that compared the mean scores for a given variable averaged over the first two postsurgical days. Recovery from surgery was assessed in a similar fashion except the scores from last two days of postsurgical observation period were averaged and analyzed.

All variables from the sucrose phase of the experiment were also compared using two-way ANOVAs. Posthoc analyses were conducted using t-tests and by one-way or two-way ANOVA. For all tests conducted during this experiment, p ≤ 0.05 was set as the statistical rejection criterion. Bonferroni adjustments were also performed to correct for multiple related comparisons.

One rat in the TRIPLEx group died shortly after surgery. Data from one other TRIPLEx rat had to be excluded from the analysis because of technical problems with its cage precluding reliable measurement of behavior. We also excluded one CTx rat from the analysis of body mass because histological analysis was not conducted for this animal, but it should be noted that its trajectory of body mass gain over the course of the experiment was consistent with that of the other rats in the group. Thus the final sample sizes, for all experiments, are as follows: TRIPLEx (n=7), SHAM (n=7), and CTx (n=8) rats.
**Histology**

At the completion of behavioral testing, the rats were deeply anesthetized with sodium pentobarbital and transcardially perfused with saline followed by 10% buffered formalin. The tongue, soft palate, and nasoincisor ducts (NID) were extracted and stored in 10% buffered formalin. The anterior two-thirds of the tongue was soaked briefly in 0.5% methylene blue and then rinsed in a purified water bath. The epithelium was stripped from the underlying muscle and connective tissue and placed between two glass slides for inspection using a standard light microscope. The total number of pale staining fungiform papillae was counted as were those that possessed a taste pore indicated by a dark blue dot. The circumvallate papilla of the posterior tongue and the NID were embedded in paraffin and sectioned (10 μm) on a rotary microtome, mounted, stained with hematoxylin and eosin, and coverslipped. All tissues were coded so that taste bud counts could be conducted by an experimenter who was blind to the exact surgical treatment of a given animal.
RESULTS

HISTOLOGY

The histological results for the groups are summarized in Table 2. Rats in the TRIPLEx group had very few, if any, remaining taste buds in the representative taste fields examined from the oral cavity when compared with the SHAM rats (anterior taste pores: $t(12) = 36.13, p < 0.0001$; circumvallate papillae: insufficient variance for test; NID: $t(12) = 10.60, p < 0.0001$). The rats in the CTx group had normal numbers of taste buds in the circumvallate papillae ($p = 0.25$), innervated by the GL, and in the NID ($p = 0.73$), innervated by the GSP, but had very few intact taste pores in the fungiform papillae. The CTx and TRIPLEx did not differ in either the number of intact taste pores ($p = 0.13$) or the number of intact fungiform papillae ($p = 0.68$) found on the anterior tongue, but both groups significantly differed from the SHAM group (all $Ps < 0.0001$). These results indicate that the nerve transections were successful and that no significant regeneration occurred.

BODY MASS

A two-way ANOVA (day x group) revealed that the groups did not differ significantly in regard to body mass during the week before surgery ($p = 0.74$) and that there was no interaction ($p = 0.68$; Table 3). However, after surgery, mean body mass values were significantly lower in TRIPLEx rats compared to both SHAM and CTx rats (Figure 1). A two-way ANOVA revealed a main group effect for body mass ($F(2, 20) = 11.22, p = 0.0005$), an effect of postsurgical day ($F(36, 720) = 118.24, p < 0.0001$), as well as a significant interaction ($F(72, 720) = 11.77, p < 0.0001$; Figure 1).
A separate two-way ANOVA revealed that the SHAM and CTx groups did not differ from one another in body mass (p = 0.82). We did observe an effect of postsurgical day (F(36, 468) = 148.66, p < 0.0001; Bonferroni-adjusted α = 0.0167) but there was no significant interaction (p = 0.99). These results indicate that the main group effect observed in the plenary ANOVA were driven by the sustained reduction in body mass exhibited in the TRIPLEx rats which significantly differed from the other groups (SHAM: F(1, 13) = 12.57, p = 0.004; CTx: F(1, 14) = 21.05, p = 0.0005; Bonferroni-adjusted α = 0.0167). Indeed, TRIPLEx rats never reached SHAM body mass levels. Moreover, these animals never recovered presurgical baseline body mass over the entire course of the postsurgical experimental time frame (> 5 weeks). In fact, we fit a line to the mean body mass values for postsurgical days 10 through 25, when the same presurgical diets were offered (i.e., both the sweetened milk and oil-chow diets) and when the rate of weight gain appeared to be relatively stable, and found that the rate of gain (i.e., the slope) was 2.65 g/day (p < 0.0001) for TRIPLEx rats and was 5.32 g/day (p < 0.0001) for SHAM rats. At these trajectories, which appeared to be stable over the 16 day observation period, in 3 additional month’s time, the predicted difference in body mass between the two groups would be a striking 359 g. Obviously, this observation is predicated on the assumption that these rates would be maintained over the three month period.

**CALORIC INTAKE**

**Total Calories**
The loss in body mass in the TRIPLEx group resulted, at least in part, from the fact that the animals consumed fewer calories than they did presurgically. Presurgical baseline caloric intake did not differ between the TRIPLEx and SHAM groups (p = 0.87; Table 3). Results from the analysis of linear regression fits of the change in caloric intake relative to presurgical baseline across postsurgical days (Figure 2) revealed that surgery had an initial impact in both groups, as estimated by the y-intercepts (b) which significantly differed from 0 (SHAM: $b = -46.7$ kcal, $t(19) = -12.8$, $p < 0.0001$; TRIPLEx: $b = -96.1$ kcal, $t(19) = -20.9$, $p < 0.0001$). Nevertheless, it is clear that the initial drop in intake was greater in the TRIPLEx group compared with the SHAM rats as confirmed by a t-test of the mean change total caloric intake on the first 2 postsurgical days relative to presurgical baseline ($t(12) = 2.62$, $p = 0.023$; Table 3).

The slopes ($m$) from the linear regression fit of change in total caloric intake relative to presurgical baseline across postsurgical days (Figure 2) had a significant rise in both the SHAM and TRIPLEx groups (SHAM: $m = 1.82$ kcal/day, $t(19) = 6.25$, $p < 0.0001$; TRIPLEx: $m = 3.20$ kcal/day, $t(19) = 8.75$, $p < 0.0001$). A t-test conducted of the mean values of the change in total caloric intake relative to baseline on postsurgical days 20 and 21 indicated that the significant difference between the groups that was present initially after surgery had disappeared ($p = 0.09$), although the TRIPLEx rats did appear to be lagging behind the SHAM group in their recovery of caloric intake (Table 3).

**Percentage of Calories Ingested Between the Two Diets**
An inspection of Figure 3 suggests that the majority of caloric intake in both groups came from the sweetened milk diet postsurgically. For each group, more than 50% of total caloric intake came from the milk diet, presurgically, and this percentage did not differ between the groups (p = 0.49; TRIPLEx = 0.62%, SHAM = 0.56%). After surgery, this preference for the sweetened milk diet increased considerably. Mean total caloric intake coming from the milk diet, averaged across all 21 days, increased significantly in both groups relative to presurgical baseline (both Ps < 0.02; TRIPLEx = 83%, SHAM = 72%).

**Calories from the Sweetened Milk Diet**

Presurgical baseline caloric intake from the sweetened milk diet did not differ between the TRIPLEx and SHAM groups (p = 0.48; Table 3). Again, the linear regression analysis revealed that although surgery had an initial impact by decreasing caloric intake from sweetened milk diet in both groups (Figure 2), the drop was almost 3 times greater in the TRIPLEx animals (SHAM: \( b = -16.7 \) kcal, \( t(19) = -10.3, p < 0.0001; \) TRIPLEx: \( b = -47.45 \) kcal, \( t(19) = -19.5, p < 0.0001 \)). A t-test conducted on the mean change on the first 2 postsurgical days relative to presurgical baseline confirmed that the initial drop in caloric intake from the sweetened milk diet was significantly greater in TRIPLEx rats relative to controls (\( t(12) = 3.23, p = 0.007; \) Table 3).

The slopes from the linear regression fit of the change in sweetened milk diet caloric intake relative to baseline across postsurgical days (Figure 2) had a significant rise in both the SHAM and TRIPLEx groups (SHAM: \( m = 1.62 \) kcal/day, \( t(19) = 12.65, p < 0.0001; \) TRIPLEx: \( m = 2.02 \) kcal/day, \( t(19) = 10.44, p < 0.0001 \)). Despite the fact that
sweetened milk diet intake progressively increased across the postsurgical test period for both groups, the TRIPLEx rats still significantly differed from the SHAM rats in terms of their recovery of sweetened milk diet caloric intake on postsurgical days 20 and 21 ($t(12) = 2.25, p = 0.04$; Table 3).

**Calories from the Oil-Chow Diet**

Presurgical baseline caloric intake from the oil-chow mash did not differ between the TRIPLEx and SHAM groups ($p = 0.79$; Table 3). The linear regression analysis of the change in caloric intake from the oil-chow mash relative to baseline across postsurgical days (Figure 2) revealed that the y-intercepts were significantly lower than 0 in both groups (SHAM: $b = -28.0 \text{ kcal}$, $t(19) = -9.62$, $p < 0.0001$; TRIPLEx: $b = -49.43 \text{ kcal}$, $t(19) = -17.4$, $p < 0.0001$). Although this change in oil-chow caloric intake appeared to be greater in TRIPLEx rats relative to that observed in the SHAM rats, a $t$-test run on mean values from postsurgical days 1 and 2 indicated that the groups did not significantly differ ($p = 0.40$; Table 3).

The postsurgical caloric intake from oil-chow significantly increased in the TRIPLEx group ($m = 1.17 \text{ kcal/day}$, $t(19) = 5.19$, $p < 0.0001$), but stayed relatively stable in the SHAM rats ($p = 0.41$). The mean change in caloric intake from oil-chow on postsurgical days 20 and 21 relative to baseline did significantly differ between the groups ($p = 0.82$; Table 3). Thus, surgery had an initial impact in both groups and although statistically speaking it was not more severe in the TRIPLEx rats than the SHAM rats, the former group nonetheless did statistically increase its oil-chow intake at a greater rate across the 3 week postsurgical observation period.
SWEETENED MILK DIET MEAL PATTERNS

Bout Size

The size of milk bouts taken by the animals presurgically did not differ between the TRIPLEx and SHAM groups (p = 0.42; Table 3). The analysis of the linear regression fits of the change in milk bout size relative to baseline by postsurgical day (Figure 4) indicated that there was a significant decrease initially for both groups (SHAM: $b = -0.73 \text{ ml}, t(19) = -4.58, p = 0.0002$; TRIPLEx: $b = -0.96 \text{ ml}, t(19) = -4.84, p = 0.0001$). A t-test conducted on the mean change in milk bout size relative to baseline comparing the two groups on days 1 and 2 was not significant (p = 0.52) indicating that the nerve transections had no additional impact on bout size relative to SHAM surgery (Table 3).

Additionally, results from this analysis revealed that milk bout size significantly increased across postsurgical days (SHAM: $m = 0.12 \text{ ml/day}, t(19) = 9.71, p < 0.0001$; TRIPLEx: $b = 0.14 \text{ ml/day}, t(19) = 8.96, p < 0.0001$). The mean change in milk bout size on postsurgical days 20 and 21 relative to baseline did not significantly differ between the groups (p = 0.88; Table 3).

Bout Number

The number of sweetened milk diet bouts initiated by the animals presurgically did not differ between the TRIPLEx and SHAM groups (p = 0.27). The analysis of the linear regression fits of the change in milk bout number relative to baseline by postsurgical day (Figure 4) indicated that there was a significant decrease initially for
both groups (SHAM: $b = -1.79$, $t(19) = -3.76$, $p = 0.001$; TRIPLEx: $b = -7.32$, $t(19) = -9.90$, $p < 0.0001$). A t-test conducted on the means from the first 2 days postsurgically confirmed that the initial drop in the number of sweetened milk diet bouts was significantly greater in TRIPLEx rats relative to controls ($t(12) = 2.46$, $p = 0.03$; Table 3).

Interestingly, the postsurgical decrease in milk bout number was relatively stable in both groups as indicated by slopes that did not significantly differ from 0 (SHAM: $p = 0.98$; TRIPLEx: $p = 0.18$). However, a t-test conducted on the mean change in milk bout number relative to baseline indicated that the greater decrease initially observed for the TRIPLEx rats in comparison to the SHAM rats just missed statistical significance on postsurgical days 20 and 21 ($p = 0.066$; Table 3).

**Bout Rate**

The rate of the sweetened milk diet ingestion within bouts did not appear to be affected by surgery. The presurgical baseline milk bout ingestion rate did not differ between the TRIPLEx and SHAM groups ($p = 0.20$; Table 3) and there was no significant change from baseline milk bout ingestion rate after surgery as indicated by the fact that the y-intercepts from the linear regression analysis (Figure 4) did not significantly differ from 0 (SHAM: $p = 0.82$; TRIPLEx: $p = 0.56$). Likewise, a t-test conducted on the mean change in milk bout ingestion rate on postsurgical days 1 and 2 relative to baseline did not detect any significant difference between the groups ($p = 0.96$; Table 3).

The slopes of the linear regression fits of the change in milk bout ingestion rate by postsurgical day did not significantly differ from 0 in either the SHAM or the TRIPLEx
group (SHAM: p = 0.05; TRIPLEx: p = 0.21; Figure 4) suggesting that bout ingestion rate remained relatively stable. However, the mean change in milk bout ingestion rate relative to baseline on postsurgical days 20 and 21 was significantly lower in the TRIPLEx group compared with controls (t(12) = 3.11, p = 0.009; Table 3).

OIL-CHOW DIET MEAL PATTERNS

Bout Size

The size of oil-chow bouts taken by the animals presurgically did not differ between the TRIPLEx and SHAM groups (p = 0.69; Table 3). The analysis of the linear regression fits of the change in oil-chow bout size by postsurgical day (Figure 4) indicated that there was a significant decrease initially for both groups (SHAM: \( b = -0.24 \) g, \( t(19) = -3.13, p = 0.008 \); TRIPLEx: \( b = -0.59 \) g, \( t(19) = -6.71, p = 0.00001 \)). However, a t-test did not detect any significant group difference in the mean change in oil-chow bout size based on the first two postsurgical days (t(12) = 1.00, p = 0.34; Table 3).

Although the TRIPLEx group progressively increased oil-chow bout size (\( m = 0.03 \) g, \( t(19) = 5.11, p = 0.0002 \)) across the postsurgical observation period and the SHAM group did not (p = 0.27), there was also no difference between the groups in the mean change in bout size on postsurgical days 20 and 21 relative to baseline (p = 0.12; Table 3).

Bout Number

Presurgical baseline oil-chow bout number did not differ between the TRIPLEx and SHAM groups (p = 0.67; Table 3). The analysis of the linear regression fits of the
change in oil-chow bout number relative to baseline by postsurgical day (Figure 4) indicated that there was a significant decrease initially for both groups (SHAM: $b = -4.18$, $t(19) = -8.52$, $p < 0.0001$; TRIPLEx: $b = -9.25$, $t(19) = -10.4$, $p < 0.0001$). The initial drop in bout number was about twice as great in the TRIPLEx rats compared with SHAM controls as assessed on postsurgical days 1 and 2 ($t(12) = 2.342$, $p = 0.037$: Table 3).

Furthermore, the results from the linear regression analysis revealed that oil-chow bout number significantly increased across postsurgical days in the TRIPLEx group ($m = 0.20$, $t(19) = 2.85$, $p = 0.01$) but was relatively stable in the SHAM controls ($p = 0.60$). Accordingly, there was also no difference between the groups in the mean change in bout number on postsurgical days 20 and 21 relative to baseline ($p = 0.18$; Table 3).

**Bout Rate**

Presurgical baseline oil-chow ingestion rate did not differ between the TRIPLEx and SHAM groups ($p = 0.18$) and there was no significant change from baseline oil-chow ingestion rate after surgery as indicated by the fact that the y-intercepts from the linear regression analysis (Figure 4) did not significantly differ from 0 (SHAM: $p = 0.15$; TRIPLEx: $p = 0.08$). This was confirmed by a lack of difference between the groups in the postsurgical change in oil-chow ingestion rate as assessed on postsurgical days 1 and 2 ($p = 0.12$). Furthermore, the slopes of the linear regression fits of the change in oil-chow ingestion rate relative to baseline by postsurgical day did not significantly differ from 0 in either the SHAM or the TRIPLEx group (SHAM: $p = 0.99$; TRIPLEx: $p = 0.75$)
suggesting that, like what was observed with milk diet, ingestion rate remained relatively stable. There was also no difference between the groups in the postsurgical change in oil-chow ingestion rate relative to baseline as assessed on postsurgical days 20 and 21 (p = 0.18).

BODY MASS REGRESSION ANALYSIS

To facilitate comparison of the body mass and microstructural data analyses, we conducted a regression analysis of body mass on the same 21 postsurgical days for which microstructural data were collected. Presurgical baseline body mass values did not differ between the groups (p = 0.95; Table 3). The analysis of the linear regression fits of the change in body mass relative to baseline by postsurgical day (Figure 5) indicated that there was a significant decrease initially for all groups (SHAM: b = -21.6 g, t(19) = -8.19, p < 0.0001; CTx: b = -28.1 g, t(19) = -12.3, p < 0.0001; TRIPLEx: b = -47.9 g, t(19) = -9.85, p < 0.0001). The initial drop in body mass was more than twice as great in the TRIPLEx rats compared with SHAM and CTx controls as assessed on postsurgical days 1 and 2 (F(2,19) = 6.60, p = 0.007; Table 3); posthoc tests revealed that the TRIPLEx group differed from both the SHAM and CTx groups (both Ps < 0.027) and that the SHAM and CTx groups did not differ from one another (p = 0.21).

The results from the linear regression analysis revealed that body mass significantly increased across postsurgical days in both the SHAM and CTx groups (SHAM: m = 4.59 g/day, t(19) = 21.85, p < 0.0001; CTx: m = 5.04 g/day, t(19) = 27.85, p < 0.0001) but not in the TRIPLEx cut group (m = 0.36 g/day, t(19) = 0.92, p = 0.37). Indeed, there was a significant difference between the groups in the change in body
mass relative to presurgical baseline on postsurgical days 20 and 21 ($F(2, 19) = 64.78, p < 0.0001$; Table 3); posthoc tests revealed that the TRIPLEx group differed from both the SHAM and CTx groups (both $Ps < 0.0001$) and that the SHAM and CTx groups did not differ from one another ($p = 0.83$).

SUCROSE

Surprisingly, both groups of animals displayed robust preference for sucrose in two-bottle intake tests. Although the SHAM group exhibited slightly greater preference than did the TRIPLEx rats, the latter group still chose to drink sucrose primarily over water at all concentrations. A two-way ANOVA revealed a main group effect for sucrose preference ($F(1,12) = 10.16, p = 0.0078$; Figure 6), an effect of sucrose concentration ($F(4, 48) = 6.75, p = 0.0002$), as well as a significant interaction ($F(4, 48) = 7.17, p = 0.0001$). Posthoc analysis revealed that rats in the SHAM group displayed significantly greater preference for sucrose at the 0.0625 and 0.5 M concentrations with the latter not surviving Bonferroni correction ($0.0625: t(12) = 3.40, p = 0.0053, 0.5: t(12) = 2.56, p = 0.03$; Figure 6; Bonferroni-adjusted $\alpha = 0.01$).

A two-way ANOVA revealed that the groups did not differ in their total intake of sucrose ($p = 0.33$), although the TRIPLEx rats trended towards lower levels of ingestion (Figure 6). There was an effect of concentration ($F(4, 48) = 16.85, p < 0.0001$) on sucrose intake, but no significant interaction was observed ($p = 0.82$).

Although both groups of rats displayed robust preference for and intake of sucrose, SHAM rats consumed the sugar solutions in an entirely different manner. Rats in the SHAM group took significantly larger sucrose bouts, relative to rats in the
TRIPLEx group. A two-way ANOVA revealed a main effect of group for sucrose bout size \((F(1,12) = 6.901, p = 0.022; \text{Figure 7})\) and a main effect of concentration \((F(4, 48) = 3.375, p = 0.016)\). No significant interaction was observed \((p = 0.09)\). Consistent with the effect of the nerve transections on sucrose bout size, a two-way ANOVA revealed that the TRIPLEx rats had a significantly slower rate of sucrose drinking \((\text{mL/min})\) within bouts compared with the SHAM group \((F(1, 12) = 15.5, p = 0.002; \text{Figure 7})\). There was also a significant effect of concentration on rate of drinking within sucrose bouts \((F(4, 48) = 5.1, p = 0.0016)\), as well as a significant interaction \((F(4, 48) = 3.2, p = 0.0196)\). Interestingly, the within bout drinking rate for SHAM rats increased as a function of concentration \((\text{one-way ANOVA: } F(4, 24) = 5.69, p = 0.0023; \text{Bonferroni-adjusted } \alpha = 0.025)\) and the higher concentrations approached twice the value seen for TRIPLEx rats, which, in turn, stayed relatively constant across the concentration range \((\text{one-way ANOVA: } p = 0.25; \text{Figure 7}; \text{Bonferroni-adjusted } \alpha = 0.025)\).

Apparently, the TRIPLEx rats compensated for their significantly decreased sucrose bout size by initiating a significantly larger number of sucrose bouts compared with the SHAM group. A two-way ANOVA for bout number revealed main effects of group \((F(1,12) = 10.641, p = 0.0068; \text{Figure 7})\) and concentration \((F(4, 48) = 21.55, p < 0.0001)\), as well as a significant interaction \((F(4, 48) = 5.93, p = 0.00058)\). Posthoc analysis revealed that the TRIPLEx group took more bouts when sampling the three highest concentrations of sucrose tested \((\text{all } p < 0.006; \text{Figure 6}; \text{Bonferroni-adjusted } \alpha = 0.01)\).
DISCUSSION

There were four principal outcomes in this experiment. First, rats that had the gustatory branches of the seventh and ninth cranial nerves transected lost body mass initially but then stabilized, even though total caloric intake steadily increased over the postsurgical observation period. Second, the absence of lingual and palatal gustatory input resulted in an initial drop in caloric intake that was effected primarily through decreases in bout number, not bout size, when rats were offered sweetened milk diet and oil-chow to which they had been presurgically exposed. Third, despite the loss of histologically confirmed oral gustatory input, TRIPLEx rats displayed elevated intake of and preference for sucrose solution similar to that observed in sham-operated controls. Finally, although the TRIPLEx rats did not differ from SHAM rats in their sucrose intake, the manner by which the two groups ingested the sugar solution varied remarkably and did not emulate what was observed with the milk diet. The conceptual ramifications of each of these findings are discussed below.

Impact of Gustatory Neurotomy on Body Mass Regulation and Caloric Intake.

Rats in the TRIPLEx group lost more body mass after surgery and stabilized at a lower level compared with the two other surgical control groups. The greater initial loss of body mass by the TRIPLEx rats can be explained by the greater drop in total caloric intake postsurgically relative to the SHAM rats. In the absence of lingual and palatal taste input, accounting for 90% of the taste bud population, body mass was more stable compared with the more progressive course of weight gain observed in controls. What
is noteworthy about the relatively lower body mass maintained during the postsurgical observation period in TRIPLEx rats is that it occurred even though total caloric intake steadily increased (Figure 2). Thus, we speculate that, in addition to having effects on feeding, the loss of oral gustatory signals, at least those provided by the seventh and ninth cranial nerves, has additional effects on nutrient absorption and/or energy expenditure. It would be instructive to explicitly test this in future studies.

Our results are consistent with the literature regarding the effects of gustatory neurotomy on body mass and food intake. For example, previous research has shown that when the CT, GL, and the pharyngeal branch of the vagus (PBV) nerves are bilaterally transected, rats do indeed lose significant levels of body mass, and, depending on the nature and composition of the diet to which the animals have access, these rats do not recover their baseline levels of body mass until anywhere between one to five weeks (29, 42). Similar results were seen when these nerves were cut in addition to the anterior palatine nerve, which supplies the taste buds of the incisive papilla in the rat (71). The interpretation of the findings from these studies must be tempered by the fact that there is no evidence that the PBV innervates taste buds, but it does provide substantial motor innervation to the pharynx (12, 32), which could have a significant impact upon feeding behavior and body mass. The possibility of a motor component is supported by the observation that “control” rats in the latter study, that only had the PBV nerve bilaterally transected, lost more weight than any of the “gustatory” control groups (rats that had either the CT, GL, or anterior palatine nerve bilaterally transected) and had a similar, but significantly different, weight loss as rats with combined transections (71).
In the current report, the TRIPLEx group did not recover baseline levels of body mass over the entire course of the five week experiment. Jacquin et al. (29) reported that after having the CT, GL, and the PBV nerves bilaterally transected, the rats, on a “cafeteria” diet, recovered to baseline levels at a mean of 34 days. Moreover, in this same report, a small number of nerve transected rats, given access to pablum, displayed hyperphagia and recovered normal intake and body mass within 10 days, suggesting that the characteristics of the diet (e.g., caloric content, nutrient content, palatability level, texture, viscosity) can interact with the nature of the neural deficit to influence body mass regulation. It should be noted, however, that a postmortem histological analysis of oral tissue was not performed in this, or in any of the other reports on the effects of gustatory deafferentation on body mass (29, 42, 71). Thus, it is uncertain whether the observed outcomes (e.g., return to baseline/control body mass levels) were the result of compensatory physiological mechanisms, remaining gustatory sensory innervation of the oral cavity (i.e., incomplete nerve transections, GSP, superior laryngeal branch of the vagus), or simply the regeneration of the transected nerves which would result in the reestablishment of some gustatory function (e.g., 23, 59). That said, in a report by Grill and Schwartz in which the neurotomies were histologically confirmed, rats that had bilateral CT and GL nerve transections, maintained a significant weight loss, relative to controls, 20 days postsurgery (25). These rats, however, only had access to standard laboratory chow. As alluded to above, it has been demonstrated that rats deprived of gustatory neural input that are given standard chow, lose substantially greater amounts of body mass than do rats maintained on a more palatable, calorically dense diet. Yet, rats in the current report were given a very
palatable diet and still did not return to baseline weight levels highlighting the importance of both diet and neurophysiology in the maintenance of body mass.

In addition to innervating the taste buds of the fungiform papillae on the anterior two-thirds of the tongue, the CT nerve also contains parasympathetic fibers that innervate the submaxillary, sublingual, and accessory salivary glands (12, 57). Likewise, the GL contains parasympathetic fibers that innervate the von Ebner's gland (7, 33). Thus, transection of the CT and GL could affect the salivary content of the oral cavity, and, in turn, possibly influence food intake and body mass. However, both the sublingual and submaxillary salivary glands receive sympathetic innervation via other nerves, and the lingual branch of the trigeminal nerve, which was left intact, also supplies some parasympathetic innervation (22, 27). Moreover, the rats in the CTx group did not lose body mass relative to SHAM controls. Nevertheless, parasympathetic fibers from the GSP, in part, innervate the mucosal glands of the palate (32) and when transection of this nerve is combined with that of the other gustatory nerves, the functional status of the oral epithelium could be adversely affected and we cannot entirely dismiss that this contributed to the effects observed here. That said, there are multiple reports that changes in the saliva content of the oral cavity, via the complete removal of some or all of these salivary glands, while increasing chow bout duration, has no impact upon daily food and fluid consumption or body mass gain (20, 56, 65, 69).

It is important to acknowledge that the GSP and GL possess somatosensory afferent fibers innervating the palate and posterior tongue respectively. It has been shown that trigeminal deafferentation of the anterior tongue leads to significant
decreases in intake and body mass (30). Accordingly, we cannot entirely dismiss the possibility that a decrease in oral somatosensory signals in the TRIPLEx rats contributed to the outcomes observed here. That said, the lingual branch of the trigeminal nerve was left intact and thus somatosensory input from the anterior two-thirds of the tongue was maintained in the TRIPLEx rats. Another consideration is that although the combined transection of the CT, GSP, and GL results in loss of 90% of taste bud population, the superior laryngeal branch of the vagus was left intact; this nerve innervates the few taste buds residing in the laryngeal epithelium. However, these taste buds are thought to be involved with protection of the airways (6, 15, 53).

**Impact of Gustatory Neurotomy on Feeding and Drinking Patterns**

After surgery, the majority of total calories ingested, from both surgical groups, came from sweetened milk diet consumption (see Figure 2). The neurotomy-induced decrease in sweetened milk diet intake can be explained by a reduction in bout frequency, but not in bout size (see Figure 4). A similar outcome was observed for the oil-chow mash diet.

In the context of feeding patterns, bout initiation can be considered as governed by purely appetitive processes, defined as the actions that bring the animal to the goal. Bout size can be considered to be controlled by both appetitive and consummatory processes, with, perhaps, a bias towards the latter, which can be defined as the final behavior of the appetitive sequence stimulated by contact of the stimulus with the receptors. This distinction is important because it has been shown through a variety of pharmacological, endocrine, and neural manipulations that mechanisms controlling bout
initiation are dissociable from those controlling bout size (39, 40, 54). Given that
gustatory signals have been implicated in the control of meal size as clearly shown by
the concentration-dependent increases in sucrose intake in sham-feeding preparations
(46, 72), it was surprising that bout size of neither sweetened milk diet or oil-chow mash
was affected by the neurotomy. It was also especially surprising that the drinking rate
during sweetened milk diet bouts did not differ between the TRIPLEx and control rats, at
least initially after surgery, given that ingestion rate has been shown to be influenced by
stimulus palatability (e.g., 13, 62, 74). We speculate that these outcomes are related to
the presurgical experience that the animals had with these diets. As such, the animals
had ample opportunity to learn about the nongustatory features of the sweetened milk
and oil-chow mash (olfactory, textural, visual) and associate them with the beneficial
nutritive properties of diets. In other words, it is possible that the conditioned palatability
based on the nongustatory cues of the diets were sufficient to maintain bout size and
bout rate. In contrast, bout number was appreciably affected initially by the neurotomy
indicating a significant consequence of the removal of gustatory signals on the
appetitive drive to approach nutritive sources and this resulted in a decrease in caloric
consumption. However, bout number tended to increase over the 21 day postsurgical
observation period, suggesting a recovery of appetitive drive, as did bout size in the
TRIPLEx rats and this led to a steady increase in caloric intake, but could not be based
on oral gustatory signals. Regardless of the underlying mechanism, it is clear, that
under some conditions, a normal meal size can be maintained in the absence of
gustatory neural input from the tongue and palate.
Impact of Gustatory Neurotomy on Sucrose Intake, Preference and Drinking Patterns.

Surprisingly, despite histologically confirmed massive gustatory deafferentation, the TRIPLEx group displayed a robust, albeit slightly attenuated, preference for sucrose and did not differ from SHAM rats in their absolute sucrose intake. High intake of (34, 42, 70, 71) or preference for (25, 42, 70) sucrose and other sweeteners by rats with various combinations of gustatory neurotomies has been reported previously. However, in prior work, we have demonstrated that after the bilateral transection of the CT, GL and GSP, behavioral responsiveness to sucrose across a broad range of concentrations is effectively eliminated (61). These prior findings were based on rats that had the same gustatory neurotomy performed using the same surgical procedures as the rats described in the current report, but sucrose responsiveness was assayed using a brief-access taste test that minimizes postingestive influences on concentration-dependent licking during a given trial (see 58 for a detailed explanation). Thus, these data suggest that postingestive factors, possibly interacting with the substantial experience of the animals with sweet-tasting stimuli (i.e., sweetened condensed milk), and not residual gustatory input from superior laryngeal nerve, drove the sucrose preference observed here. This hypothesis is supported by data gathered with T1R3 knockout mice. T1R3 is a protein that serves as a component of the canonical sweet taste receptor, which mediates the perception of sweeteners in mammals (75). In long-term intake tests, these animals initially show greatly reduced preference for sucrose across a broad range of concentrations, relative to wild type controls, but with repeated testing eventually display hearty preferences for the sugar (75, 77, 78). Indeed, other types of
knock-out mice missing critical taste transduction components can display significant preference for sweeteners, including sucrose, if they have had prior testing experience with the compound (e.g., 51). Thus, it would appear that normal gustatory function is not necessary for sucrose preferences as measured in long-term intake tests to be expressed by rodents.

Interestingly, despite having relatively normal preference and intake of sucrose, the manner in which the TRIPLEx rats ingested this taste stimulus was entirely different from the way that SHAM rats did. Relative to SHAM rats, TRIPLEx rats had significantly smaller sucrose drinking bouts and compensated for this decrease by initiating a significantly greater number of bouts over the 23-hour period. Apparently, the loss of gustatory neural input influences the ingestion of sucrose by decreasing the maintenance of licking during bouts likely because of the absence of excitatory signals. This could be viewed as a decrease in the orosensory-based hedonic effectiveness of the stimulus (55). In support of this interpretation, the bout drinking rate, thought to reflect the motivational state of the animal (e.g., 13, 55, 62), was significantly lower in TRIPLEx rats relative to SHAM controls and was not dependent on sucrose concentration. Thus, the prior experience with the sweetened condensed milk diet pre- and postsurgically, during which animals could learn that calories were available from the fluid source that was distinguishable from water (not necessarily based on taste), was likely responsible for increasing the probability at which the animals returned to the sucrose drinking spout to initiate a bout. It is noteworthy that the sweetened milk diet contained about 18% sugar and there is evidence that sucrose has a detectable odor starting at concentrations of 0.03 M (~1.3%, (47)). One possible reason that SHAM
animals had a lower frequency of bout initiation could arguably be due to the greater postingestive inhibition that they experienced from the larger bouts they had that maintained satiety longer.

Although it is possible that the nerve transections affected the oromotor competence of the animals, Spector et al. (61), found that TRIPLEX rats had interlick intervals, a measure often used to assess the status of the putative central pattern generator for licking, that were not significantly different from controls. Moreover, while the animals in that study displayed virtually flat sucrose concentration-response functions in a brief access test, their maximum licks across all of the trials was no different from sham-operated animals. Finally, the bout drinking rate observed for the sweetened milk diet was greater than that for sucrose in the TRIPLEX rats demonstrating that these animals had the capacity to display higher rates of ingestion.

The profile of sucrose drinking patterns in TRIPLEX rats is opposite to what we observed when the animals were presented with sweetened milk diet after surgery (i.e., no difference in bout size and a decrease in bout number). The differences appear to relate to the chemical composition of the stimuli (pure sugar solution vs. liquid mixture of fat, protein, sugar, minerals, and vitamins) and the extensive presurgical experience the rats had with the sweetened milk diet vs. the lack of prior experience with pure sucrose solutions. The nongustatory cue of the sucrose is only one component of the complex milk diet, which could in some sense be considered a conditioned stimulus, and while it is sufficient to drive choice and intake, it is apparently insufficient to drive drinking bout size and rate in marked contrast to the control rats. Such results highlight the important
distinction between measures of preference and intake and measures of the organization of licking itself.

**Perspectives**

On a final note, it is worth considering some clinical ramifications of these findings. First, there is the well-documented decrease in appetite associated with cancer treatments. This decrease in food intake can impact upon a broad number of factors including tissue healing, energy levels, mood, social interactions, and overall quality of life (17, 38). Although it is difficult to completely disambiguate the contributions of the disease, as well as the nonspecific consequences of the therapy including effects on salivation, a growing body of current research suggests that taste loss is a contributing factor to the decrease in appetite observed in cancer patients (4, 37, 38, 66). For example, it has been demonstrated by numerous laboratories that alterations in chemosensory function in cancer patients predict dietary intake patterns, as well as quality of life (e.g., 9, 21). The use of radiation and/or cytotoxic agents to combat the cancer is thought to be the major cause of taste dysfunction in cancer patients (17, 18, 31, 64, 73) via influences on taste bud cell survival, replacement and proliferation (3, 44, 45, 48). Collectively, these data provide powerful support for the role of taste in maintaining eating habits and quality of life. Current treatment for taste loss due to cancer treatment consists primarily of dietary manipulation focused on enhancing the flavor of food with more intense tastes (17, 18). The results presented here would suggest that, in addition to presenting more intense flavors, giving these patients a familiar palatable caloric supplement, particularly a liquid diet such as Ensure, before
treatment begins may help to mitigate the influence of cytotoxic cancer treatments on the subsequent intake of the caloric supplement during treatment.

Another clinically relevant observation is that the postsurgical difference in body mass between the SHAM and TRIPLEX rats resembled the magnitude of the difference seen between control rats and those that have received Roux-en-Y gastric bypass (e.g., 10, 26, 36, 52). Currently, gastric bypass is considered the most effective treatment to cause long-term weight loss far surpassing what is seen with pharmacological interventions or dietary restriction regimens. These data provide strong support for the important contribution of the gustatory system in the regulation of body mass and although, obviously, gustatory neurotomy would not be a viable treatment option for obesity, the development of more selective pharmacological interventions targeting the gustatory pathways could have some potential utility in curbing hyperphagia and promoting body mass regulation.
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DISCLOSURES

C.D. Dotson, C.L. Colbert, M. Garcea, J.C. Smith, and A.C. Spector have no conflicts of interest.

AUTHOR CONTRIBUTIONS

A.C.S. and J.C.S. designed research; C.L.C. and M.G. conducted research; C.D.D., C.L.C., J.C.S., and A.C.S. analyzed data; C.D.D. and A.C.S. wrote the paper. A.C.S. had primary responsibility for final content.
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Figure 1:
Body mass values assessed for SHAM, CTx, and TRIPLEx groups measured over the entire surgical period (means ± SEM).

Figure 2:
Daily total calories (left), calories from the sweetened milk diet (middle), and calories from oil-chow mash (right) ingested by SHAM and TRIPLEx groups measured for three weeks postsurgery.

Figure 3:
The percentage of total calories ingested from the sweetened milk diet measured for three weeks postsurgery.

Figure 4:
Daily bout size (left), bout number (middle), and bout rate (right) when ingesting the sweetened milk diet (top panel) and the oil-chow mash (bottom panel) by SHAM and TRIPLEx groups measured for three weeks postsurgery.

Figure 5:
Body mass values assessed for SHAM and TRIPLEx groups measured for three weeks postsurgery.

Figure 6:
Sucrose preference vs. water (left) and intake (right) as a function of concentration for SHAM and TRIPLEx groups (means ± SEM).

Figure 7:
Bout size (left), bout number (middle), and bout rate (right) as a function of concentration for SHAM and TRIPLEx groups when ingesting sucrose during preference testing (means ± SEM).
<table>
<thead>
<tr>
<th>Test Day</th>
<th>Food available</th>
<th>Fluid (other than water*) available</th>
</tr>
</thead>
<tbody>
<tr>
<td>1 - 5</td>
<td>Powdered chow</td>
<td>Nothing</td>
</tr>
<tr>
<td>6 - 11</td>
<td>chow + oil</td>
<td>Nothing</td>
</tr>
<tr>
<td>12 - 44</td>
<td>chow + oil</td>
<td>Sweetened milk diet</td>
</tr>
<tr>
<td>45 - 46</td>
<td>chow + oil</td>
<td>Nothing</td>
</tr>
<tr>
<td>47 - 48</td>
<td>chow + oil</td>
<td>Water</td>
</tr>
<tr>
<td>49 - 50</td>
<td>chow + oil</td>
<td>0.06 M Sucrose</td>
</tr>
<tr>
<td>51 - 52</td>
<td>chow + oil</td>
<td>0.125 M Sucrose</td>
</tr>
<tr>
<td>53 - 54</td>
<td>chow + oil</td>
<td>0.25 M Sucrose</td>
</tr>
<tr>
<td>55 - 56</td>
<td>chow + oil</td>
<td>0.5 M Sucrose</td>
</tr>
<tr>
<td>57 - 58</td>
<td>chow + oil</td>
<td>1.0 M Sucrose</td>
</tr>
</tbody>
</table>

* Water in one bottle was always available.
### Table 2: Histology Results

<table>
<thead>
<tr>
<th>Group</th>
<th>Pores in Anterior Tongue</th>
<th>Number of Intact Fungiform Papillae</th>
<th>% Intact Fungiform Papillae Containing Pore</th>
<th>Number of Taste Buds in CV</th>
<th>Number of Taste Buds in NID</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sham</td>
<td>148.2 ± 4.05</td>
<td>156.1 ± 4.03</td>
<td>94.9 ± 0.95</td>
<td>374.8 ±10.21</td>
<td>108.8 ± 10.60</td>
</tr>
<tr>
<td>Sham + CTx</td>
<td>9.0 ± 3.60</td>
<td>88.3 ± 7.76</td>
<td>8.8 ± 3.15</td>
<td>399.5 ± 18.86</td>
<td>113.9 ± 9.84</td>
</tr>
<tr>
<td>TRIPLEX</td>
<td>3.5 ± 1.12</td>
<td>84.0 ± 6.59</td>
<td>4.06 ± 1.14</td>
<td>0.0 ± 0.00</td>
<td>2.1 ± 0.88</td>
</tr>
<tr>
<td>Measure</td>
<td>Mean of the Last Two Presurgical Days (i.e., Presurgical Baseline)</td>
<td>Mean Change from Presurgical Baseline on the First Two Postsurgical Days</td>
<td>Mean Change from Presurgical Baseline on the Last Two Postsurgical Days</td>
<td></td>
<td></td>
</tr>
<tr>
<td>--------------------------------</td>
<td>--------------------------------------------------------------------</td>
<td>------------------------------------------------------------------------</td>
<td>------------------------------------------------------------------------</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Sham</td>
<td>TRIPLEx</td>
<td>CTx</td>
<td>P value</td>
<td>Sham</td>
</tr>
<tr>
<td><strong>Caloric Intake</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total calories</td>
<td>127.64 ± 13.11</td>
<td>130.33 ± 9.25</td>
<td></td>
<td>0.87</td>
<td>-101.52 ± 11.35</td>
</tr>
<tr>
<td>Milk diet calories</td>
<td>71.09 ± 9.75</td>
<td>80.24 ± 8.03</td>
<td></td>
<td>0.48</td>
<td>-19.24 ± 6.61</td>
</tr>
<tr>
<td>Milk diet bout #</td>
<td>14.14 ± 1.89</td>
<td>16.86 ± 1.40</td>
<td></td>
<td>0.27</td>
<td>-3.43 ± 1.16</td>
</tr>
<tr>
<td>Milk diet bout size</td>
<td>4.57 ± 0.41</td>
<td>4.01 ± 0.53</td>
<td></td>
<td>0.42</td>
<td>-0.45 ± 0.44</td>
</tr>
<tr>
<td>Milk diet bout rate</td>
<td>1.89 ± 0.14</td>
<td>1.65 ± 0.10</td>
<td></td>
<td>0.20</td>
<td>-0.12 ± 0.09</td>
</tr>
<tr>
<td>Oil-chow diet calories</td>
<td>54.51 ± 11.03</td>
<td>50.87 ± 7.83</td>
<td></td>
<td>0.79</td>
<td>-37.39 ± 12.51</td>
</tr>
<tr>
<td>Oil-chow diet bout #</td>
<td>13.36 ± 1.37</td>
<td>14.14 ± 1.16</td>
<td></td>
<td>0.67</td>
<td>-5.50 ± 1.37</td>
</tr>
<tr>
<td>Oil-chow diet bout size</td>
<td>0.69 ± 0.12</td>
<td>0.63 ± 0.11</td>
<td></td>
<td>0.69</td>
<td>-0.43 ± 0.12</td>
</tr>
<tr>
<td>Oil-chow diet bout rate</td>
<td>0.25 ± 0.03</td>
<td>0.31 ± 0.03</td>
<td></td>
<td>0.18</td>
<td>-0.07 ± 0.04</td>
</tr>
<tr>
<td><strong>Body Mass</strong></td>
<td>415.86 ± 11.85</td>
<td>415.86 ± 11.85</td>
<td></td>
<td>0.95</td>
<td>-2.00 ± 3.22</td>
</tr>
</tbody>
</table>

All values are mean ± standard error.

* Posthoc tests revealed that the TRIPLEx group differed from both the SHAM and CTx groups (both Ps < 0.027) and that the SHAM and CTx groups did not differ from one another (p = 0.21).

# Posthoc tests revealed that the TRIPLEx group differed from both the SHAM and CTx groups (both Ps < 0.0001) and that the SHAM and CTx groups did not differ from one another (p = 0.83).
Percent of Total Calories Ingested from the Sweetened Milk Diet